

ORIGINAL ARTICLE

Conservation priorities of Iberoamerican pig breeds and their ancestors based on microsatellite information

O Cortés¹, AM Martínez², J Cañón¹, N Sevane¹, LT Gama³, C Ginja^{4,5}, V Landi², P Zaragoza⁶, N Carolino⁷, A Vicente^{7,8}, P Sponenberg⁹ and JV Delgado² for the BioPig Consortium¹⁰

Criollo pig breeds are descendants from pigs brought to the American continent starting with Columbus second trip in 1493. Pigs currently play a key role in social economy and community cultural identity in Latin America. The aim of this study was to establish conservation priorities among a comprehensive group of Criollo pig breeds based on a set of 24 microsatellite markers and using different criteria. Spain and Portugal pig breeds, wild boar populations of different European geographic origins and commercial pig breeds were included in the analysis as potential genetic influences in the development of Criollo pig breeds. Different methods, differing in the weight given to within- and between-breed genetic variability, were used in order to estimate the contribution of each breed to global genetic diversity. As expected, the partial contribution to total heterozygosity gave high priority to Criollo pig breeds, whereas Weitzman procedures prioritized Iberian Peninsula breeds. With the combined within- and between-breed approaches, different conservation priorities were achieved. The Core Set methodologies highly prioritized Criollo pig breeds (Cr. Boliviano, Cr. Pacifico, Cr. Cubano and Cr. Guadalupe). However, weighing the between- and within-breed components with F_{ST} and $1-F_{ST}$, respectively, resulted in higher contributions of Iberian breeds. In spite of the different conservation priorities according to the methodology used, other factors in addition to genetic information also need to be considered in conservation programmes, such as the economic, cultural or historical value of the breeds involved.

Heredity (2016) **117**, 14–24; doi:10.1038/hdy.2016.21; published online 30 March 2016

INTRODUCTION

Pig breeds located in the Iberian Peninsula are mainly classified as Mediterranean and Celtic types. Mediterranean-type pigs (generally grouped in the Iberian trunk) include several breeds, sometimes divided into lineages, that display a high degree of phenotypic heterogeneity, even though they show a certain degree of genetic proximity (Gama *et al.*, 2013). Celtic-type pigs derive from northern–central European pig breeds (Royo *et al.*, 2007), and it is documented that Chinese germplasm was introduced in this group in the distant past (McGlone and Pond, 2003). On the other hand, the Mediterranean-type pig is assumed to be the pre-extant type in Iberian Peninsula. Currently, Mediterranean pigs are mainly located in the southwest of the Iberian Peninsula, in a Mediterranean ecosystem traditionally known as ‘Dehesa’, whereas Celtic pig types have been raised in the Peninsula’s northern part. The second trip (1493) of Columbus took horses, cattle, sheep, goats and pigs from the Iberian Peninsula to America for the first time. The first pigs were taken to Haiti and subsequent arrivals to the American continent favoured their widespread expansion throughout Latin America under a wide range of environmental conditions, thus giving rise to the populations currently known as Criollo pigs. Currently, 17% of the world’s pigs are

located in South America where they are widespread from Mexico to Southern Argentina and from sea level to 4500 m of altitude (Benítez, 2000). Criollo pig breeds are characterized by their sexual precocity, poor reproductive efficiency, long lactations, low weaning weights but a better hardiness than commercial pig breeds (Linares *et al.*, 2011). As a consequence of their distribution in many rural areas under diverse environmental conditions, pig production plays a key role in the social economy as a source of food and income in Latin America. Furthermore, Criollo pig breeds form part of the social traditions that reinforce local communities and their identities. There is a growing awareness of the importance of these breeds in terms of adaptive traits to specific (often extreme) environments and as genetic reservoirs, in addition to their historical or cultural value that plays a key role in the sustainable development of marginal rural areas (Ponzoni, 1997; Oldenbroek, 1999; Ruane, 1999). High levels of genetic diversity decrease the probability of extinction of a breed, as they give the breed a better chance of adapting to environmental and/or production system changes in the future (Fisher, 1930). It is therefore necessary to implement policies focussed on the conservation of genetic diversity, even more so in cases in which breed viability is threatened by inbreeding derived from reproductive isolation and

¹Departamento de Producción Animal, Facultad de Veterinaria, Madrid, Spain; ²Departamento de Genética, Universidad de Córdoba, Córdoba, Spain; ³CIISA, Faculdade de Medicina Veterinária, Universidade de Lisboa, Lisboa, Portugal; ⁴Ce3C—Centro de Ecologia, Evolução e Alterações Ambientais, Faculdade de Ciências, Universidade de Lisboa, Lisboa, Portugal; ⁵CIBIO-InBIO—Centro de Investigação em Biodiversidade e Recursos Genéticos, Universidade do Porto, Campus Agrário de Vairão, Vairão, Portugal; ⁶Laboratório de Genética Bioquímica, Facultad de Veterinaria, Universidad de Zaragoza, Zaragoza, Spain; ⁷Instituto Nacional de Investigação Agrária e Veterinária, I.P. Unidade Estratégica de Investigação e Serviços de Biotecnologia e Recursos Genéticos Polo de Investigação da Fonte Boa, Vale de Santarém, Portugal; ⁸Escola Superior Agrária, Instituto Politécnico de Santarém, Vale de Santarém, Portugal and ⁹Regional College of Veterinary Medicine, Virginia Tech, Blacksburg, VA, USA

Correspondence: O Cortés, Departamento de Producción Animal, Facultad de Veterinaria, Universidad Complutense de Madrid, Madrid 28040, Spain.

E-mail: ocortes@vet.ucm.es

¹⁰Members of the consortium are listed before the references.

Received 21 October 2015; revised 22 January 2016; accepted 2 February 2016; published online 30 March 2016

genetic drift. Several strategies have been developed in order to maintain the genetic diversity of a single population or breed. However, when a group of breeds is considered, new problems arise because in order to maintain the global genetic diversity some breeds may have to be prioritized relative to the others. In this context, the choice of a breed/s for conservation depends on its contribution to overall genetic diversity, and the importance of both between- and within-breed genetic variation must be considered and appropriately weighted in conservation decisions, even though the estimation of such contributions is not immediately obvious (Barker, 2001; Toro *et al.*, 2002; Bruford, 2004; Ginja *et al.*, 2013). Weitzman approach (Weitzman, 1992) is often used as a measure of the between-breed genetic diversity to calculate the partial contribution (PC) of each breed to total genetic diversity. This methodology is ultimately based on the pairwise genetic distances between operational taxonomical units. However, this approach, which prioritizes breeds that are more differentiated, has been criticized for not considering within-breed information (Ruane, 1999; Toro *et al.*, 2009). On the other hand, the contribution of within-breed genetic diversity to total diversity could be estimated by the loss or gain of the global heterozygosity when each breed is removed from the metapopulation. Taking into account that both within- and between-components of genetic diversity should be considered to maximize the retention of genetic diversity, various weights applied to each component have been developed (Caballero and Toro, 2002; Piyasatianand Kinghorn, 2003; Olivier *et al.*, 2005). For example, Olivier and Foulley (2005) combined the two components by weighing Weitzman approach by the degree of breed differentiation (Wright's fixation index F_{ST} (Fisher, 1930) and the within-breed diversity component, measured as the global loss or gain in heterozygosity when a breed is removed, weighted by $1-F_{ST}$. However, a general consensus regarding the appropriate weight and the methodology that better combines both sources of information has not been reached. Notwithstanding the limitations of the above methodologies, different alternatives have been proposed that combine both sources of information based on kinship coefficients. Although initially these methodologies considered genealogical information in the absence of pedigree connections, as is often the case when various breeds are considered, molecular markers have proved efficient to estimate between-breed kinships and, as an extension, they have been used to weigh the contribution of each breed to total genetic diversity (Cañón *et al.*, 2011; Ginja *et al.*, 2013). Eding and Meuwissen (2001) proposed a Core Set methodology with the aim of minimizing the global coancestry of the metapopulation based on molecular marker kinship estimations (MEK). The same authors later proposed two extensions of MEK based on log-linear regressions, obtained with the weighted log-linear model (WLM) or the weighted log-linear mixed model (WLMM) (Eding and Meuwissen, 2001, 2003), using average molecular coancestries (f_m) based on allele frequencies (Caballero and Toro, 2002).

Another methodology for combining within- and between-breed information that weights the within-contribution (based on average coancestries of each breed) and between-contribution (based on genetic distances) equally was developed by Caballero and Toro (2002) and Faubel *et al.* (2004). In spite of the different methodologies proposed, there is no definite recommendation related to the relative importance of the between- and within-components of genetic diversity, and how they should be weighed in establishing conservation priorities.

Several studies have estimated the genetic diversity of pig breeds worldwide (Faubel *et al.*, 2004; Ollivier and Foulley, 2005; Vicente *et al.*, 2008; Burgos-Paz *et al.*, 2013; Gama *et al.*, 2013; Herrero-

Medrano *et al.*, 2013; Ramírez *et al.*, 2014; Revidatti *et al.*, 2014). Among those studies, specific results have been published based on the same set of breeds and microsatellites assessed here, reporting the genetic relationships among Iberian Peninsula pig breeds (Gama *et al.*, 2013) and Criollo pig breeds (Revidatti *et al.*, 2014). In addition, Gama *et al.* (2013) briefly evaluated conservation priorities among Iberian pig breeds considering the current levels of within- and between-breed genetic diversity. However, a comprehensive analysis in terms of conservation priorities of Criollo breeds is lacking, in spite of their importance in the social economy, where they play a major role as a source of food and income. In the present study, a comprehensive sampling of Criollo pig breeds and their ancestors (Iberian and Commercial pig breeds) were jointly analysed in order to evaluate conservation priorities based on different methodologies that considered either within-breed, or between-breed or both genetic diversity components.

MATERIALS AND METHODS

Sampling and genotyping

A total of 1670 samples belonging to 45 populations/breeds from the Iberian Peninsula (Spain and Portugal), Europe (local and commercial breeds) and America were analysed (Supplementary Figure 1 and Table 1). Individual genotypes have been previously reported by Gama *et al.* (2013) for Iberian Peninsula breeds and by Revidatti *et al.* (2014) for Criollo pig breeds for a panel of 24 microsatellites recommended for genetic diversity studies by the International Society for Animal Genetics/Food and Agriculture Organization of the United Nations. In addition, wild boar samples from four different European geographic locations (Spain, Portugal, Italy and Poland) that covered the two major mitochondrial DNA haplogroups observed in the continental European wild boars (E1a and E1c) (Scandura *et al.*, 2011) were included in the analysis.

Commercial (Duroc, Pietrain, Large White and Landrace) and the remaining European local pig breeds (Berkshire, Tamworth and Large Black) were genotyped for the same panel of 24 microsatellites used in Gama *et al.* (2013) and Revidatti *et al.* (2014): IGF1, S0002, S0005, S0026, S0068, S0090, S0101, S0155, S0178, S0215, S0225, S0226, S0227, S0228, S0355, S0386, SW024, SW072, SW240, SW632, SW857, SW911, SW936 and SW951. Laboratory protocols have been described in detail in Gama *et al.* (2013) and Revidatti *et al.* (2014) and allele nomenclature was standardized according to the ISAG (International Society for Animal Genetics) Pig Comparison Test.

All the European pig breeds included in the analysis are officially recognized and are registered in herdbooks except for Negro de Formentera (NFO) and Negro de los Pedroches (NPE). However, the Criollo pig populations analysed are bred under quite different management systems, and most of them are known by their geographical location rather than receiving an official breed recognition. Therefore, the Criollo animals included in our analyses were not enrolled in official herdbooks, but they belong to what is considered the traditional type of the Criollo population that they belong to.

Two different analyses were carried out: (1) each breed was considered as a unit without any grouping; (2) breeds were grouped based on a previous analysis of their genetic relationships (Gama *et al.*, 2013; Revidatti *et al.*, 2014). In this case, Iberian breeds were divided in two groups (Mediterranean and Celtic) based on the neighbour-net drawn from Nei's DA genetic distance and STRUCTURE results previously reported by Gama *et al.* (2013). According to their geographic origin, local European pig breeds (Large Black, Berkshire and Tamworth) were included in the Celtic group. Criollo and commercial pig breeds were grouped according to Revidatti *et al.* (2014) and Martínez (data not shown). Therefore, the breeds analysed were grouped as follows: *Celtic group* (CT): Chato Murciano (CHM), Berkshire (BRK), Large Black (LBL), Tamworth (TWR), Malhado de Alcobaça (MAL), Negro Canario (NCA), Celta (CEL) and Bisaro (BIS); *Mediterranean group* (MD): Manchado de Jabugo (MJA), Euskal Txerria (ETX), Negro de Formentera (NFO), Mangaliça (MAN), Torbiscal (TOR), Retinto (RET), Entrepelado (ENT), Lampiño (LAM), Alentejano (ALE), Negro de los Pedroches (NPE) and Negro Mallorquin (NMA); *Commercial 1* (C1): Duroc (DUR); and *Commercial 2* (C2): Landrace

Table 1 Breed groups, breed names, acronyms, number of samples genotyped per breed (*N*), country of origin, mean observed (H_o) and expected heterozygosities (H_e) and mean number of alleles per locus (MNA) estimated with 24 microsatellites in 45 pig breeds

Group	Breed	Acronym	<i>N</i>	Country	H_e	H_o	MNA
C1	DUROC	DUR	50	UK	0.56	0.55	4.0
C2	PIETRAIN	PIE	46	UK	0.60	0.60	5.2
C2	LARGE WHITE	LWH	29	UK	0.56	0.55	4.6
C2	LANDRACE	LDR	26	UK	0.60	0.57	4.4
C2	LARGE WHITEXLANDRACE	LXW	19	UK	0.61	0.67	4.5
CA	MULEFOOT	MFT	36	USA	0.43	0.41	3.3
CA	CRIOLLO DE EL SALVADOR	SAL	21	El Salvador	0.67	0.57	6.3
CA	CRIOLLO DEL PACIFICO	CCP	42	Colombia	0.69	0.64	8.0
CA	CRIOLLO BOLIVIANO	BOL	34	Bolivia	0.66	0.63	7.2
CA	PAMPA ROCHA	PRC	32	Uruguay	0.58	0.57	5.1
CA	CARACOLERO DRY	NED	50	Argentina	0.70	0.60	8.2
CA	CARACOLERO WET	NEW	40	Argentina	0.63	0.55	6.8
CB	RED WATTLE HOG	RWH	35	USA	0.49	0.48	4.3
CB	NEW MEXICO	BCS	20	USA	0.63	0.55	5.4
CB	PELON MEXICANO	MEX	49	Mexico	0.63	0.45	6.5
CB	CRIOLLO CUBANO	CUB	50	Cuba	0.65	0.65	7.6
CB	CRIOLLO VENEZOLANO	VEN	30	Venezuela	0.64	0.59	6.3
CB	ZUNGO	ZUN	33	Colombia	0.60	0.58	5.2
CB	CRIOLLO ECUATORIANO	ECU	50	Ecuador	0.70	0.61	8.7
CC	GUINEA HOG	GHO	34	USA	0.56	0.48	5.2
CC	CRIOLLO DE GUADALUPE	GUA	35	Guadeloupe	0.69	0.64	7.1
CC	SANPEDREÑO	SPD	14	Colombia	0.63	0.64	5.0
CT	CHATO MURCIANO	CHM	50	Spain	0.46	0.39	4.5
CT	NEGRO CANARIO	NCA	50	Spain	0.55	0.49	5.8
CT	CELTA	CEL	27	Spain	0.60	0.50	4.5
CT	BISARO	BIS	49	Portugal	0.63	0.55	5.6
CT	MALHADO DE ALCOBAÇA	MAL	36	Portugal	0.52	0.51	3.7
CT	BERKSHIRE	BRK	47	UK	0.44	0.45	3.2
CT	TAMWORTH	TWR	41	UK	0.52	0.52	3.3
CT	LARGE BLACK	LBL	47	UK	0.54	0.51	4.5
MD	RETINTO	RET	50	Spain	0.55	0.52	5.5
MD	ENTREPELADO	ENT	50	Spain	0.56	0.56	5.5
MD	TORBISCAL	TOR	50	Spain	0.53	0.47	4.8
MD	NEGRO DE LOS PEDROCHES	NPE	29	Spain	0.56	0.55	4.2
MD	LAMPIÑO	LAM	50	Spain	0.57	0.53	5.8
MD	MANCHADO DE JABUGO	MJA	41	Spain	0.39	0.40	3.4
MD	NEGRO DE FORMENTERA	NFO	20	Spain	0.42	0.35	3.8
MD	NEGRO MALLORQUIN	NMA	20	Spain	0.56	0.51	4.6
MD	EUSKAL TXERRIA	ETX	50	Spain	0.39	0.39	3.8
MD	ALENTEJANO	ALE	50	Portugal	0.59	0.52	6.5
MD	MANGALIÇA	MAN	25	Hungary	0.42	0.35	3.2
WB	WILD-BOAR PORTUGAL	PWB	39	Portugal	0.59	0.56	6.5
WB	WILD-BOAR SPAIN	SWB	50	Spain	0.60	0.55	6.3
WB	WILD-BOAR POLAND	EWB	12	Poland	0.58	0.58	4.5
WB	WILD-BOAR ITALY	IWB	12	Italy	0.46	0.49	3.0
Total		45	1670	18	0.57 ± 0.04	0.53 ± 0.02	5.23 ± 2

(LAN), Large White (LWH), Pietrain (PIE) and Large White × Landrace crosses (LXW). Criollo breeds were grouped in three groups: *Criollo A* (CA): Mulefoot (MFT), Pampa Rocha (PRC), Caracolero Wet (NEW), Criollo Boliviano (BOL), Criollo El Salvador (SAL), Criollo Pacifico (CCP) and Caracolero Dry (NED); *Criollo B* (CB): Red Wattle Hog (RWH), Criollo Cubano (CUB), Pelon Mexicano (MEX), Zungo (ZUN), New Mexico (BCS), Criollo Venezolano (VEN) and Criollo Ecuatoriano (ECU); and *Criollo C* (CC): Guinea Hog (GHO), Criollo Guadalupe (GUA) and Sanpedreño (SPD). Finally, all wild boar samples were included in a single group comprising the following populations: wild boar from Italy (IWB), Spain (SWB), Portugal (PWB) and Poland (EWB).

For a better understanding of the statistical methodologies used, the different approaches were classified as follows: methods that considered (1) between-breed (*Weitzman*) or (2) within-breed (*Heterozygosity*) genetic diversity; methods that combined (3) Weitzman and Heterozygosity procedures; methods that (4) minimize the overall kinship coefficients (*Core Set*) and finally a combined approach that considered (5) coancestry (within) and Nei's genetic distance (between) sources of information.

Weitzman procedure

The Reynold genetic distance (Reynolds *et al.*, 1983) was used in the algorithm described by Weitzman (1992) to calculate the partial contribution (PC_{WE}) of

each breed to total genetic diversity. Because of the large number of breeds analysed, the application of Weitzman formula is too computationally intensive and not practical, and hence an approximation described by García *et al.* (2005) was used when each breed was considered as a unit. Thresholds ranging from 0.5 to 0.25 were used to test the coherence of the results. The threshold corresponds to the coefficient of variation of the diversity matrix. In this procedure, in each iteration, the coefficient of variation is assessed, and if it is lower than the threshold, the approximation is launched; otherwise the exact algorithm is applied. Therefore, the lower the threshold, the more accurate the results will be, at the expense of computation time. A threshold of 0.25 was finally used to assess breed contributions to the overall diversity.

Heterozygosity

Observed and unbiased expected heterozygosities and average number of alleles per locus were obtained with microsatellite toolkit (Park, 2001). The partial contribution (PC_{He}) of each breed to the total genetic diversity was estimated as the proportion of heterozygosity lost when each breed or group is removed from the data set.

Weitzman and heterozygosity combined approaches

In a first set of analyses, PC_{WE} and PC_{He} were analysed together in order to assess the contribution of within- and between-breed information to total genetic diversity. Ollivier *et al.* (2005) proposed an aggregate diversity equation that uses Wright's F_{ST} to weight between- and within-breed genetic diversities as: $PC_{FST} = PC_{WE} \times F_{ST} + PC_{He} \times (1 - F_{ST})$. F -statistics were estimated according to Weir and Cockerham (1984) using GENETIX 4.05.2 software (Belkhir *et al.*, 1996–2004). In addition, Piyasatian and Kinghorn (2003) suggested an alternative aggregate approximation that gives greater weight to the between-breed component: $PC_{5:1} = 0.833 \times PC_{WE} + (1 - 0.833) \times PC_{He}$, and this procedure was also assessed in our data set.

Core Set methodologies

The Core Set methodology of Eding *et al.* (2002) combines within- and between-breed information with the goal of minimizing the kinship coefficient of the metapopulation. Because of the absence of genealogical data, MEK (Eding and Meuwissen, 2001) and their extensions, that is, the WLM and WLMM, were used (Eding and Meuwissen, 2003). A matrix M containing the average MEK between and within populations is constructed, and the relative contributions of each population are estimated in such a way that the average MEK in the Core Set is minimal. However, because of sampling errors in the relationships matrix, it is possible to find negative values. In this scenario, an iterative process assigns a zero to the more negative value and recalculates the contributions after removal of that population, until all contributions are equal to or greater than zero. Eding and Meuwissen (2003) described two variations of MEK based on log-linear regression obtained with the WLM, which is designed to further reduce the number of zero contributions, and WLMM, which allows for only one null contribution, and hence it is easier to establish a ranking of all populations for conservation priorities.

Finally, average molecular coancestries based on allele frequencies (f_m), estimated as described by Caballero and Toro (2002), were also used. The MEK, WLM and WLMM were estimated following the methodology described in Cañón *et al.* (2011) that is based on Eding and Meuwissen (2001) for MEK and Eding and Meuwissen (2003) for WLM and WLMM. Partial contributions to the overall genetic diversity for each methodology (PC_{MEK} , PC_{WLM} , PC_{WLMM} , PC_{f_m}) were estimated with a FORTRAN programme developed by Eding and Meuwissen (2003).

Coancestry and Nei's genetic distance combined approach

This methodology combined both sources of information but considered other estimations for between- and within-breed genetic diversity than Weitzman and Heterozygosity approaches. Caballero and Toro (2002) and Fabuel *et al.* (2004) proposed this methodology using the same rationale behind the Core Set method. Based on the definition of diversity as the opposite of average coancestry, they partitioned the coancestry in two components. The within-breed genetic diversity estimated as the average coancestry for each breed and the between-breed component estimated as the average minimum Nei's

distance for each breed to the others. This methodology considers different weights for each population proportionally to the effective or the actual population size, and in the absence of this information, sample sizes can be used. However, under this scenario, the accuracy of the contributions estimated depends on the similarity among the ratio of breed sample size to total sample size and the ratio of breed population size to total population size. These estimates were also calculated ignoring sample sizes, as suggested by Fabuel *et al.* (2004).

RESULTS

A total of 348 alleles were detected across the 24 autosomal microsatellite markers analysed. The mean number of allele/locus per breed (Table 1) was 5.2 ± 2 , ranging from 3.2 (Mangalica and Berkshire) to 8.7 (Cr. Ecuatoriano). Expected and observed heterozygosities by breed (Table 1) ranged from 0.39 to 0.7 and from 0.35 to 0.67, respectively. Generally, Mediterranean and Celtic pig breeds had lower expected and observed heterozygosities than Criollo pig breeds. The mixed origin of the Landrace \times Large White population probably explains its high observed heterozygosity. When the pig breeds were grouped in Mediterranean, Celtuc, Criollo, Commercial and Wild Pig categories, the expected heterozygosity values increased because of the reproductive isolation among the breeds grouped; as could be anticipated, the differences among H_e and H_o were also greater than in individual pig breeds because of substructuring (Table 2).

Weitzman and heterozygosity procedures

The partial contributions of each pig breed to total genetic diversity according to the different thresholds following Weitzman procedure (PC_{WE}) are shown in Table 3. The results show that a total of 17 pig breeds were prioritized for conservation (contributions greater than 3), mainly consisting of exotic, some Mediterranean and some Celtic pig breeds (represented in bold in Table 3). The second group prioritized for conservation (contributions between 2 and 3) and was formed by seven pig breeds (Landrace, Negro Mallorquin, Wild Boar-Poland, Wild Boar-Portugal, Pietrain, Large White \times Landrace, Pampa Rocha and Negro de los Pedroches). The remaining pig breeds had lower PC_{WE} and were of the Criollo type. As expected, the PC_{He} showed opposite results to PC_{WE} (Table 3) and, with a few exceptions, Criollo pig breeds had the highest PC_{He} . The majority of the Mediterranean and Celtic pig breeds were not prioritized for conservation under this criterion, and recent well-documented bottlenecks (Gama *et al.*, 2013) could explain their negative estimated PC_{He} values.

Table 2 Breed groups, number of samples genotyped per group (N), mean observed (H_o) and expected heterozygosities (H_e) and mean number of alleles per locus (MNA) estimated with 24 microsatellites in 8 pig groups

	Group	N	H_e	H_o	MNA
Iberian	MD	435	0.63	0.48	10.5
	CT	347	0.69	0.49	9.9
Criollo	CA	255	0.70	0.57	12.1
	CB	267	0.68	0.56	11.7
	CC	83	0.69	0.58	9.3
Commercial	C1	50	0.56	0.55	4.0
	C2	120	0.69	0.59	7.4
Wild Boar	WB	113	0.65	0.55	9.0
Total	8	1670	0.66 ± 0.05	0.55 ± 0.04	9.2 ± 3.5

Table 3 Contributions of pig breeds to overall genetic diversity according to: Weitzman method (PC_{WE}), proportional variation of expected heterozygosity (PC_{He}), aggregate diversity (PC_{Fst}), the Piyasatian and Kinghorn equation ($PC_{5.1}$), marker estimated kinships (MEKs), weighted log-linear model (WLM), weighted log-linear mixed model (WLMM) and average coancestries (fm)

Group	Acronym	PC_{WE}	PC_{He}	$PC_{WE-PC_{He}}$ combined approaches			Core Set			
				PC_{Fst}	$PC_{5.1}$	PC_{MEK}	PC_{WLM}	PC_{WLMM}	fm	
C1	DUR	3.7	-0.03	0.74	3.11	0	0.00433	0.027	0.019	
C2	LWH	4.5	0.09	1.00	3.77	0.105	0.08	0.035	0.158	
C2	PIE	2.2	0.17	0.59	1.88	0	0.01	0.028	0	
C2	LDR	2.5	0.28	0.73	2.11	0.055	0.06	0.028	0.093	
C2	LXW	2.2	0.32	0.71	1.88	0.063	0	0.030	0.029	
CA	MFT	4.8	-0.47	0.61	3.92	0	0	0.010	0	
CA	PRC	2.1	0.13	0.53	1.74	0	0	0.016	0	
CA	NED	0.8	0.22	0.34	0.72	0	0	0.021	0	
CA	BOL	0.9	0.33	0.44	0.78	0	0.20	0.036	0.138	
CA	SAL	0.3	0.43	0.40	0.32	0	0	0.023	0	
CA	CCP	0.8	0.59	0.63	0.75	0.206	0.03	0.027	0.074	
CA	NEW	0.4	0.60	0.55	0.41	0.056	0.01	0.027	0	
CB	RWH	4.1	-0.23	0.65	3.34	0	0	0.014	0	
CB	CUB	0.6	-0.10	0.03	0.45	0	0.32	0.036	0	
CB	MEX	0.5	0.18	0.25	0.45	0	0	0.022	0	
CB	ZUN	1.2	0.18	0.38	1.00	0	0	0.023	0	
CB	BCS	0.9	0.22	0.37	0.82	0	0.01	0.027	0.086	
CB	VEN	1.1	0.31	0.46	0.93	0	0.02	0.026	0	
CB	ECU	0.3	0.56	0.51	0.34	0	0	0.026	0.029	
CC	GHO	3.7	-0.02	0.75	3.08	0.102	0.04	0.026	0	
CC	SPD	1.0	0.27	0.41	0.86	0	0	0.023	0	
CC	GUA	0.9	0.52	0.59	0.80	0.383	0.10	0.028	0.727	
CT	CHM	4.5	-0.49	0.53	3.64	0	0	0.011	0	
CT	BRK	4.9	-0.41	0.69	4.05	0	0	0.014	0	
CT	LBL	4.2	-0.12	0.76	3.46	0	0.23	0.031	0.01	
CT	TWR	4.6	-0.06	0.89	3.81	0.070	0	0.024	0.084	
CT	MAL	4.0	-0.04	0.79	3.31	0	0	0.016	0	
CT	NCA	4.3	0.05	0.91	3.55	0.100	0.00047	0.025	0.100	
CT	CEL	1.1	0.09	0.31	0.96	0	0	0.021	0	
CT	BIS	1.0	0.26	0.41	0.86	0	0	0.023	0	
MD	MJA	4.7	-0.81	0.32	3.77	0	0	0	0	
MD	ETX	4.5	-0.75	0.32	3.59	0	0	0.021	0	
MD	NFO	4.8	-0.64	0.47	3.86	0	0	0.011	0	
MD	MAN	4.6	-0.61	0.45	3.71	0	0	0.018	0	
MD	TOR	3.8	-0.21	0.62	3.16	0	0	0.017	0	
MD	RET	0.4	-0.19	-0.06	0.32	0	0	0.015	0	
MD	ENT	0.3	-0.15	-0.05	0.25	0	0	0.015	0	
MD	LAM	1.0	-0.09	0.13	0.78	0	0	0.018	0	
MD	ALE	0.8	-0.08	0.10	0.65	0	0	0.017	0	
MD	NPE	1.6	-0.06	0.28	1.30	0	0	0.016	0	
MD	NMA	2.4	0.04	0.52	2.00	0	0	0.016	0	
WB	IWB	4.9	-0.40	0.69	4.01	0.205	0.03	0.027	0.096	
WB	EWB	2.4	-0.01	0.48	1.98	0	0.02	0.029	0.007	
WB	PWB	2.0	0.03	0.42	1.63	0	0.05	0.031	0	
WB	SWB	1.1	0.10	0.31	0.95	0	0	0.023	0	

Highly prioritized breeds are in bold and shaded dark.

When pig breeds were analysed by group (Table 4), commercial groups (C1 and C2) showed greater PC_{WE} , whereas Criollo groups, CA and CB had lower contributions to overall diversity. In addition, MD and CT considerably reduced their PC_{WE} as a consequence of their higher genetic heterogeneity that reduced differences with respect to the other breed groups. On the other hand, CA and C2 had the

highest PC_{He} (> 1), whereas a total of three groups had negative PC_{He} (MD, C1 and WB).

Overall, the C2 group had both high PC_{HE} and PC_{WE} values, even though these estimates are considered to represent distinct sources of information regarding contributions to global genetic diversity. Therefore, the results indicate that the breeds in this group not only have a

Table 4 Contributions of pig groups to overall genetic diversity according to: Weitzman method (PC_{WE}), proportional variation of expected heterozygosity (PC_{He}), aggregate diversity (PC_{Fst}), the Piyasatian and Kinghorn proposition ($PC_{5:1}$), average coancestries (fm), marker estimated kinships (MEKs), weighted log-linear model (WLM) and weighted log-linear mixed model (WLMM)

Acronym	PC_{WE}	PC_{He}	$PC_{WE-PC_{He}}$ combined approaches		Core Set				
			PC_{Fst}	$PC_{5:1}$	PC_{MEK}	PC_{WLM}	PC_{WLMM}	fm	
MD	9.3	-1.3	-0.6	7.5	0	0	0	0	0
CT	6.6	0.87	1.2	5.6	0	0	0.05	0.02	
CA	3.2	1.15	1.3	2.8	0.28	0.23	0.15	0.18	
CB	3.5	0.45	0.6	2.9	0	0.05	0.1	0	
CC	11.9	0.53	1.2	10.0	0.11	0.19	0.16	0.1	
C1	33.7	-2.3	-0.1	27.7	0.09	0.05	0.08	0.09	
C2	24.0	1.08	2.5	20.2	0.4	0.26	0.24	0.42	
WB	14.1	-0.5	0.4	11.7	0.13	0.22	0.22	0.2	

Highly prioritized breeds are in bold and shaded dark.

high level of genetic differentiation but also high levels of heterozygosity.

Weitzman and heterozygosity combined approaches

The between-breed component was weighted by the overall F_{ST} value of 0.21 when individual pig breeds were analysed or 0.06 when breed groups were considered.

The linear combinations of between- and within-breed genetic contributions to overall diversity gave conservation priority to the breeds in groups C1 and C2 and some in CT when the PC_{Fst} criterion was used, but the highest priority was given to Large White, Mulefoot, several CT breeds and IWB when the $PC_{5:1}$ perspective was adopted (Table 3). Among Criollo breeds, only Mulefoot and Guinea Hog were located in the upper linear combination range of the values estimated. The Negro Canario breed showed high values for both linear combinations, as a consequence of its high PC_{WE} and positive PC_{He} .

When breed groups are considered (Table 4), the C2 and C1 groups showed the highest values for PC_{Fst} and $PC_{5:1}$, respectively. In spite of the negative value of C1 group for PC_{He} , its high PC_{WE} and the weight given to this component explained the highest value for this group in $PC_{5:1}$.

Core Set methodologies

The large number of breeds with null contributions to overall genetic diversity when kinship-based methods are used together (PC_{MEK} , PC_{WLM} , and PC_{fm}) allows that highly prioritized populations can be easily identified (Table 3). Positive values for the three methods were found in most of the commercial pig breeds. However, the highest individual values for MEK, WLM and fm criteria were obtained for the Criollo breeds Guadalupe, Cubano and Guadalupe, respectively. It is remarkable that all pig breeds in the MD group showed null contributions with the MEK, WLM and fm methodologies. Except for the Mediterranean types, at least one pig breed belonging to each group had positive values for both PC_{MEK} , PC_{WLM} and PC_{fm} . This happened specifically for Large White and Landrace in commercial breeds, Pacifico, Caracolero Wet and Guinea Hog in Criollo, Negro Canario in Celtic type pigs and the Italian group of wild boar. Figure 1 shows contour plots of MEKs estimated from individual breeds

(Figure 1a) or breed-group genotypes (Figure 1b). Individual breeds and groups of breeds were sorted according to the average within-marker estimated kinships, with hot colour areas (red) representing higher inbreeding coefficients. In Figure 1a, except for one Criollo breed (Mulefoot), the breeds with higher levels of within-breed kinships belonged to Mediterranean or Celtic types: Berkshire, Chato Murciano, Manchado de Jabugo, Euskal Txerria, Negro de Formentera, Mangalica, Lampiño, Entrepelado and Retinto. The contour plot also revealed between-breed kinships and, as expected, Mediterranean pig breeds were the more closely related with each other (green areas in Figure 1a). On the other hand, Lampiño, Entrepelado, Retinto, Manchado de Jabugo and Negro de Formentera were the Mediterranean breeds more closely related to their counterparts (light blue). In Figure 1b, the Duroc, Iberian and wild boar groups showed higher within-group kinship levels and, as previously shown (Gama *et al.*, 2013), the Mediterranean and wild boar populations had the highest breed relationships.

The PC_{WLMM} methodology allows for only one null contribution, and hence it is easier to establish a ranking of all pig breeds for conservation priorities. Although some individual Criollo breeds, such as Criollo Boliviano and Criollo Cubano, showed high PC_{WLMM} estimates (0.36), on average the commercial pig breeds had the highest PC_{WLMM} values (Table 3). Overall, three pig breeds (Large White, Guadalupe and Negro Canario) and the Italian wild boar population showed estimates greater than 0 for the four kinship methodologies.

When breed groups were considered, the C2 group had the highest contribution to overall genetic diversity in the four Core Set methodologies (Table 4).

Coancestry and Nei's genetic distance combined approach

The results of the calculations of breed contributions following the methodology of Caballero and Toro (2002) are shown in Table 5, both considering and ignoring sample sizes (Fabel *et al.*, 2004).

The Caracolero Wet and Cr. Ecuador had the highest $PC_{WEIGHTED}$ and $PC_{UNWEIGHTED}$, respectively, and the five breeds having the highest contributions with both methodologies were Criollo pig breeds. However, if the sample sizes assumed in the calculations do not reflect the effective population size of each breed in relation to the size of the metapopulation, then those breeds with larger samples could be favoured. In order to avoid the possible misinterpretation that would result in this case, the contribution of each breed was also estimated ignoring sample size, but only minor differences were detected among breeds. It is also arguable whether ignoring sample sizes is much better than considering these to obtain biased contributions (see Fabel *et al.*, 2004 for an additional discussion).

Surprisingly, when pig breeds were grouped, the MD and CT had the highest contribution to overall genetic diversity (Table 6), most probably because of their larger sample sizes. On the other hand, when sample sizes were ignored, MD had the lowest contribution and C2 the highest.

DISCUSSION

The development of livestock species in the American continent had particular characteristics because of the introduction of some domesticated farm animals, including pigs, since 1492. Hence, the current pig breeds located in the American continent originated initially from animals introduced by the first settlers that over time have adapted to different local environments. In the past few years, the genetic improvement of pig breeds, particularly in Europe and the United States, allowed for a second wave of genetic introgression into

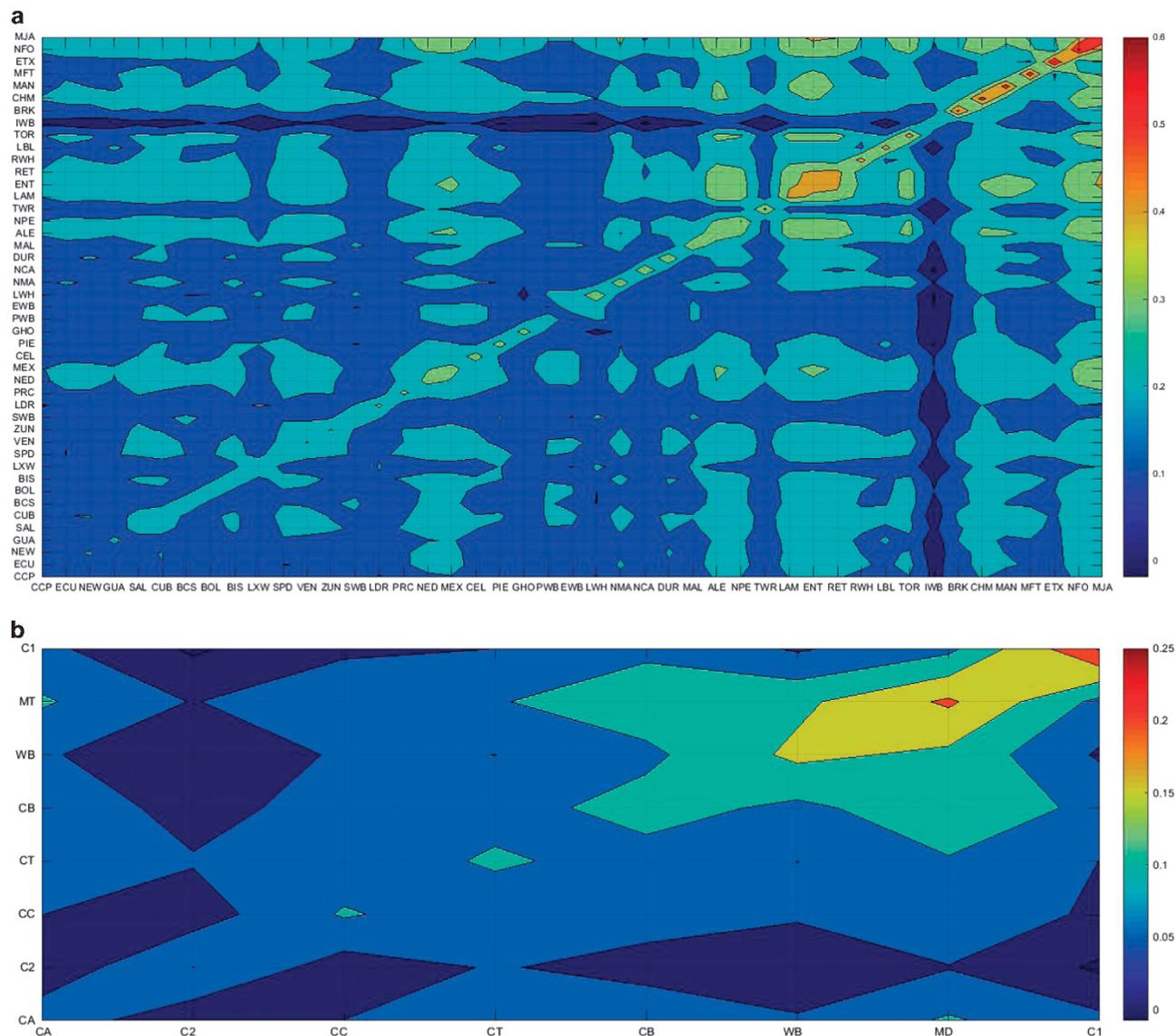


Figure 1 (a) Contour plots of MEKs: Chato Murciano (CHM), Berkshire (BRK), Large Black (LBL), Tamworth (TWR), Malhado de Alcobaça (MAL), Negro Canario (NCA), Celta (CEL) and Bisaro (BIS), Manchado de Jabugo (MJA), Euskal Txerria (ETX), Negro de Formentera (NFO), Mangaliça (MAN), Torbiscal (TOR), Retinto (RET), Entrepelado (ENT), Lampiño (LAM), Alentejano (ALE), Negro de los Pedroches (NPE), Negro Mallorquin (NMA), Duroc (DUR), Landrace (LAN), Large White (LWH), Pietrain (PIE), Large White × Landrace crosses (LXW), Mulefoot (MFT), Pampa Rocha (PRC), Caracolero Wet (NEW), Criollo Boliviano (BOL), Criollo El Salvador (SAL), Criollo Pacifico (CCP), Caracolero Dry (NED), Red Wattle Hog (RWH), Criollo Cubano (CUB), Pelon Mexicano (MEX), Zungo (ZUN), New Mexico (BCS), Criollo Venezolano (VEN) and Criollo Ecuatoriano (ECU), Guinea Hog (GHO), Criollo Guadalupe (GUA) and Sanpedreño (SPD). Wild boar populations were also included: Wild Boar Italy (IWB), Wild Boar Spain (SWB), Wild Boar Portugal (PWB) and Wild Boar Europe (EWB). Hot colour areas (red) representing higher inbreeding coefficients. (b) Contour plots of MEKs of group pigs breeds: Celtic type (CT), Mediterranean type (MT), Commercial group 1 (C1), Commercial group 2 (C2), Criollo group A (CA), Criollo group B (CB) and Criollo group C (CC). Hot colour areas (red) representing higher inbreeding coefficients.

America's pig populations of commercial and highly selected pig breeds (Food and Agriculture Organization (FAO), 2007). Although the term breed emerged some 200 years ago in Europe, corresponding to animals with uniform characteristics, in America the term 'Criollo' has a very broad meaning, referring to direct descendants of the animals brought to the New World by the Spaniards and Portuguese, such that, in many cases, animals with a high degree of heterogeneity are grouped under the classification of Criollo. Nevertheless, the actual ancestry of current Criollo pig breeds is not clear, given the influences that pig breeds of several

origins may have had and/or the differences among extant Iberian pig breeds and those that existed centuries ago.

Currently, the conservation of animal genetic resources is universally accepted as a key factor in sustainable development and to respond to changing conditions, including climate change, new or resurgent disease threats, new requirements on human nutrition and changing market demands or societal needs (Hoffman, 2010). In addition, human food demand in developing countries is predicted to rise at a faster rate than the increase in food production. Therefore, the greater the efforts for conservation of animal genetic resources, the

Table 5 Contributions of pig breeds to overall diversity according to Caballero and Toro (2002) and Fabuel *et al.* (2004)

Group	Acronym	<i>Fii</i>	<i>Dnei</i>	Contribution to <i>f</i>	Contribution to <i>D</i>	<i>PC</i> _{WEIGHTED} (%)	<i>PC</i> _{UNWEIGHTED} (%)
C1	DUR	0.44	0.16	0.0085	0.021	3.0	2.2
C2	PIE	0.40	0.15	0.0070	0.021	2.9	2.3
C2	LWH	0.44	0.21	0.0040	0.013	1.9	2.4
C2	LDR	0.40	0.17	0.0036	0.012	1.7	2.4
C2	LXW	0.39	0.16	0.0026	0.009	1.2	2.4
CA	MFT	0.57	0.20	0.0079	0.014	1.9	2.0
CA	SAL	0.33	0.10	0.0029	0.010	1.4	2.4
CA	CCP	0.31	0.11	0.0051	0.020	2.8	2.4
CA	BOL	0.34	0.13	0.0043	0.016	2.3	2.4
CA	PRC	0.42	0.14	0.0054	0.014	1.9	2.2
CA	NEW	0.30	0.10	0.0061	0.024	3.3	2.4
CA	NED	0.37	0.11	0.0064	0.018	2.5	2.3
CB	RWH	0.51	0.17	0.0071	0.014	1.9	2.1
CB	BCS	0.37	0.13	0.0029	0.009	1.3	2.4
CB	MEX	0.37	0.10	0.0078	0.022	3.0	2.3
CB	CUB	0.35	0.11	0.0072	0.023	3.2	2.4
CB	VEN	0.35	0.11	0.0043	0.014	1.9	2.4
CB	ZUN	0.40	0.13	0.0053	0.014	2.0	2.3
CB	ECU	0.30	0.09	0.0064	0.024	3.3	2.4
CC	GHO	0.44	0.17	0.0055	0.015	2.1	2.3
CC	GUA	0.31	0.10	0.0043	0.017	2.3	2.5
CC	SPD	0.37	0.12	0.0021	0.006	0.9	2.3
CT	CHM	0.54	0.18	0.0109	0.019	2.7	2.0
CT	NCA	0.45	0.19	0.0076	0.022	3.1	2.3
CT	CEL	0.40	0.13	0.0044	0.012	1.6	2.3
CT	BIS	0.37	0.11	0.0075	0.022	3.1	2.3
CT	MAL	0.48	0.17	0.0066	0.015	2.1	2.2
CT	BRK	0.56	0.20	0.0099	0.018	2.6	2.0
CT	TWR	0.48	0.20	0.0071	0.017	2.5	2.2
CT	LBL	0.46	0.18	0.0080	0.020	2.8	2.2
MD	RET	0.45	0.12	0.0098	0.020	2.8	2.1
MD	ENT	0.44	0.11	0.0097	0.020	2.8	2.1
MD	TOR	0.47	0.16	0.0094	0.021	2.9	2.1
MD	NPE	0.44	0.13	0.0054	0.012	1.7	2.1
MD	LAM	0.42	0.12	0.0091	0.021	2.9	2.2
MD	MJA	0.61	0.18	0.0106	0.014	2.0	1.8
MD	NFO	0.58	0.19	0.0047	0.007	1.0	1.9
MD	NMA	0.44	0.15	0.0035	0.008	1.2	2.2
MD	ETX	0.61	0.25	0.0108	0.019	2.7	2.0
MD	ALE	0.41	0.10	0.0092	0.021	2.9	2.1
MD	MAN	0.57	0.20	0.0055	0.009	1.3	2.0
WB	PWB	0.41	0.14	0.0063	0.017	2.4	2.3
WB	SWB	0.40	0.13	0.0081	0.022	3.0	2.3
WB	EWB	0.42	0.15	0.0020	0.005	0.7	2.3
WB	IWB	0.55	0.24	0.0022	0.005	0.7	2.2

Average coancestries (*fii*) and Nei's genetic distances (*DNei*), contributions to global coancestry (*f*) and to average Nei's distance (*D*), and proportional contributions (*PC*) to a pool of maximum genetic diversity weighted and unweighted by sample sizes. Highly prioritized breeds are in bold and shaded dark.

greater the probability of satisfying the predictable increase in human food demands (Hoffman, 2010).

In spite of the fact that the term breed is well accepted as a unit for conservation, funds available for conservation of animal genetic resources are limited, and the criteria used to invest public capital to maintain them may imply the definition of breed conservation priorities. Factors that need to be considered in the definition of such criteria include, in addition to genetic diversity and environmental sustainability aspects, those related to the social, cultural and historical role of breeds (Ruane, 1999; Gandini *et al.*, 2004). Concerning the factors associated with the contribution of a breed to genetic diversity,

there is no consensus on how to weight the between- and within-population components of diversity (Caballero and Toro, 2002; Eding *et al.*, 2002). In principle, maintenance of between-breed genetic diversity should be prioritized if the aim of the breeding programme in a medium-term scenario is to focus on crossbreeding plans. However, in anticipation of possible environmental changes, within-breed genetic diversity could be more important to face unforeseen challenges. In our analyses, the Weitzman and heterozygosity approaches were in accordance with the history of the breeds analysed. As the Weitzman approach favours the between-population diversity component, the more distant breeds were prioritized and, in

Table 6 Contributions of group pig breeds to diversity according to Caballero and Toro (2002) and Fabuel *et al.* (2004)

Group	N	<i>fii</i>	<i>Dnei</i>	Contribution to <i>f</i>	Contribution to <i>D</i>	<i>PC</i> _{WEIGHTED} (%)	<i>PC</i> _{UNWEIGHTED} (%)
MD	435	0.37	0.17	0.053	0.21	24.8	11.6
CT	347	0.31	0.15	0.033	0.17	20.9	12.3
CA	255	0.30	0.13	0.026	0.13	15.2	12.1
CB	267	0.32	0.12	0.031	0.13	15.5	11.8
CC	83	0.31	0.17	0.007	0.04	5.1	12.5
C1	50	0.44	0.32	0.004	0.03	3.1	12.8
C2	120	0.31	0.27	0.003	0.07	8.2	14.0
WB	113	0.35	0.23	0.008	0.06	7.1	12.8

Average coancestries (*fii*) and Nei's genetic distances (*Dnei*), contributions to global coancestry (*f*) and to average Nei's distance (*D*), and proportional contributions (*PC*) to a pool of maximum genetic diversity weighted and unweighted by sample sizes.

consequence, breeds belonging to the Celtic (CT) and Mediterranean (MD) groups showed the highest *PC*_{WE} values. However, although seven of the nine CT breeds had a high *PC*_{WE}, the MD breeds showed a high variation in *PC*_{WE} among breeds, such that MD breeds with a well-known influence in Criollo breeds, including Retinto, Entrepelado or Lampiño, clearly had a lower *PC*_{WE} than those breeds with a minor influence, such as Euskal Txerria, Negro de Formentera or Mangalica (Burgos-Paz *et al.*, 2013). On the other hand, commercial breeds had an intermediate *PC*_{WE} whereas the heterogeneous genetic origins of Criollo breeds explain their low *PC*_{WE}. Nonetheless, Mulefoot and Red Wattle Hog were the only Criollo breeds with high *PC*_{WE}, probably as a consequence of their strong degree of between-breed differentiation among Criollo pig breeds, as well as within-breed homogeneity (Revidatti *et al.*, 2014). In contrast, the heterozygosity-based approach prioritized within-population diversity, and hence Criollo breeds had the highest values, except for a few exceptions such as Mulefoot and Red Wattle Hog, in these cases probably as a result of population bottlenecks given their reduced census (<http://www.albc-usa.org/cpl/wtchlist.html#pigs>). On the other hand, the MD and CT groups had the lowest *PC*_{HE}, indicating that their levels of genetic diversity are generally lower than in Commercial and Criollo breeds. These results indicate that, in general, genetically unhealthy breeds (that is, highly inbred or with low heterozygosity) are prioritized under a between-breed genetic diversity criterion, whereas breeds holding higher genetic variability receive the priority under the within-breed criterion. Therefore, depending on the perspective taken, conservation priorities can be argued for or against each option. However, the differences in management of the breeds analysed, where the majority of the Criollo breeds are open to a certain degree of genetic introgression and gene flow, whereas MD, CT and Commercial breeds are closed, may lead to different solutions, by taking into account the breed considered. Thus, although commercial breeds do not require funding for their preservation, because their maintenance is not under threat, the maintenance of highly differentiated breeds within the MD and CT groups is needed to prevent the permanent loss of the genetic diversity that they represent. In Criollo breeds, which have been selected for different environments for several centuries, the preservation of within-breed genetic diversity ensures their ability to cope with environmental changes. Furthermore, the use of systematic cross-breeding programmes with more distant breeds (including MD, CT or Commercial breeds) could probably allow taking advantage of complementarity among breeds, thus achieving high levels of heterosis. However, most criteria used for the establishment of conservation priorities will ignore Criollo breeds with low levels of heterozygosity, such as Mulefoot, Red Wattle Hog, Guinea Hog or Cr. Cubano that are characterized by a low census or recent signals of strong

bottlenecks (<http://www.albc-usa.org/cpl/wtchlist.html#pigs>; Velázquez *et al.*, 1998), but can nevertheless retain important adaptive traits.

Approaches based on minimizing molecular coancestry, which consider both between- and within-breed information, resulted in a very low conservation priority given to all MD breeds and the majority of CT breeds, except Large Black. On the other hand, most commercial breeds had positive contributions to overall diversity, even though their preservation requires less funding support. Still, Criollo breeds generally had the highest contribution values, even though many Criollo breeds had null contributions to overall genetic diversity. Among the Criollo breeds, the Cr. Boliviano, Cr. Pacífico, Cr. Cubano and Cr. Guadalupe were the breeds with higher contributions to overall diversity, but these are also believed to be the ones that may have suffered a greater influence from pigs of Chinese origin (Burgos-Paz *et al.*, 2013).

Under the kinship methodologies, however, breed conservation priorities clearly reflect the genetic structure of the pig breeds analysed. The genetic influence observed in most Criollo pig breeds is expected to derive from breeds of the Iberian Peninsula (Ramírez *et al.*, 2009), and hence the conservation of some Criollo populations should also imply the underlying conservation of the Iberian Peninsula genetic contribution. For this reason, the majority of breeds from the Iberian Peninsula had low contributions, and only three pig breeds belonging to MD or CT (Negro de Formentera, Large Black and Tamworth) had notable contributions to overall diversity, probably because of their island origin (that is, isolated) and reduced genetic influence on Criollo pig breeds. On the other hand, Cr. Guadalupe, Cr. Pacífico and Cr. Cubano had higher contribution values with the kinship methodology, probably reflecting a greater influence received from Chinese pigs (Burgos-Paz *et al.*, 2013).

The methodologies presented here to establish conservation priorities are based upon the use of neutral genetic markers, but the current availability of high-density single-nucleotide polymorphism (SNP) arrays provides extensive information on polymorphic variation across coding and noncoding genome regions, and increase the precision and accuracy of kinship estimates within and among populations. Their use is also becoming feasible because of the lower genotyping costs, for example, of the Porcine SNP60 Bead-array (Ramos *et al.*, 2009) that has recently been used for comparative studies between European pig breeds (Manunza *et al.*, 2013; Burgos-Paz *et al.*, 2013; Herrero-Medrano *et al.*, 2014). However, there is a well-known ascertainment bias implicit when SNP chips are used in cattle and other livestock species (Albrechtsen *et al.*, 2010; Lachance and Tishkoff, 2013). Furthermore, this bias is even greater when studies involve highly differentiated local populations, for instance, the Criollo pig breeds that are usually not considered in the development of high-density SNP panels.

In addition to genetic information and breed contributions to the conservation of genetic resources, other aspects such as the regional importance of the breed and its social, cultural, environmental and economic impact must also be considered in the final decision for conservation. Pig production is an important source of high-quality protein food and provides income from the sale of animals that contribute directly to support local economies, but also, in many cases, plays an important social and cultural role, especially in traditions that reinforce local identity. These strong interactions among the human community, the breed and the surrounding environment are an integral part of the environmental and social equilibrium of a given region, which is passed through generations, and is a part of local cultural heritage (Ruane, 1999). Moreover, the community will often maintain a wealth of indigenous knowledge on how to sustainably co-manage the animals and the local environment, an example of which is the maintenance of the so-called 'dehesa' or 'montado' for several centuries in the Iberian Peninsula. Therefore, in addition to genetic considerations, the cultural and environmental importance of a breed should also be taken in consideration when defining conservation priorities.

DATA ARCHIVING

Data available from the Dryad Digital Repository: <http://dx.doi.org/10.5061/dryad.7cq0k>.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We thank the different breeders associations and research groups who kindly provided biological samples used in this study. Members of the CYTED XII-H and CONBIAND networks are thanked for valuable cooperation over the years. Catarina Ginja received funding from the Fundação para a Ciência e a Tecnologia, Portugal, through an Invited Scientist fellowship (Ref. SFRH/BCC/52687/2014).

THE BIOPIG CONSORTIUM

Águeda Pons¹¹, AP Ponce Alvarado¹², A Sierra Vasquez¹³, Begoña Peinado Ramón¹⁴, Cecilio Barba Capote¹⁵, E Pérez Pineda¹⁶, Inmaculada Martín Burriel¹⁷, José Antonio Carril Gonzalez-Barros¹⁸, José Luis Vega-Pla¹⁹, JJ Montes-Sánchez²⁰, LA Alvarez²¹, María Inés Carolino²², Mariano Gómez²³, S Llambi²⁴, M Montenegro²⁴, MA Revidatti²⁵ and R Galíndez²⁶

¹¹Serveis de Millora Agrària i Pesquera, Islas Baleares, Spain; ¹²Universidad Autónoma Juan Misael Saracho de Tarija, Tarija, Bolivia; ¹³ITA Conkal, Yucatán, México; ¹⁴Centro de Investigación y Desarrollo Agroalimentario, La Alberca, Murcia, Spain; ¹⁵Departamento de Producción Animal, Universidad de Córdoba, Campus de Excelencia Internacional Agroalimentario ceiA3, Córdoba, Spain; ¹⁶Universidad de Granma, Bayamo, Cuba; ¹⁷Clementina Rodellar: Laboratorio de Genética Bioquímica, Facultad de Veterinaria, Universidad de Zaragoza, Zaragoza, Spain; ¹⁸ASOPORCEL, Palacio de Ferias y Congresos, Lugo, Spain; ¹⁹Laboratorio de Investigación Aplicada, Cría Caballar de las Fuerzas Armadas, Córdoba, Spain; ²⁰Centro de Investigaciones Biológicas del Noroeste, La Paz, Baja California Sur, México; ²¹Universidad Nacional de Colombia, Sede Palmira, Palmira, Valle del Cauca, Colombia; ²²Conceição O Sousa, Fátima Santos Silva: INIAV, Vale de Santarém, Portugal; ²³Servicio de Ganadería, Diputación Foral de Bizkaia, Bilbao, Spain; ²⁴Departamento de Genética y Mejora Animal, Instituto de Producción Animal, UDELAR, Montevideo, Uruguay; ²⁵Departamento de Producción Animal, Facultad de Ciencias Veterinarias, Universidad Nacional del Nordeste, Corrientes, Argentina; ²⁶Universidad Central de Venezuela, Maracay, Venezuela

- Albrechtsen A, Nielsen FC, Nielsen R (2010). Ascertainment biases in SNP chips affect measures of population divergence. *Mol Biol Evol* **27**: 2534–2547.
- Barker JSF (2001). Conservation and management of genetic diversity: a domestic animal perspective. *Can J Forest Res* **31**: 588–595.
- Belkhir K, Borsa P, Chikhi L, Raufaste N., Bonhomme F (1996–2004). *GENETIX 4.05, logiciel sous Windows TM pour la génétique des populations*. Laboratoire Génome, Populations, Interactions, CNRS UMR 5171. Université de Montpellier II: Montpellier (France).
- Benítez W (2000). Los cerdos criollos en America Latina. En: *Los cerdos locales en los sistemas de producción tradicionales*. Estudio FAO producción y sanidad animal, pp 148–191.
- Bruford MW (2004). Conservation genetics of UK livestock: from molecules to management. In: Simm G, Villanueva B, Townsend S (eds), *Farm Animal Genetic Resources*. Nottingham University Press: Nottingham, UK, pp 151–169.
- Burgos-Paz W, Souza CA, Megens HJ, Ramayo-Caldas Y, Melo M, Lemús-Flores C et al. (2013). Porcine colonization of the Americas: a 60k SNP history. *Heredity* **110**: 321–330.
- Caballero A, Toro MA (2002). Analysis of genetic diversity for the management of conserved subdivided populations. *Conserv Genet* **3**: 289–299.
- Cañón J, García D, Delgado JV, Dunner S, Gama LT, Landi V et al. (2011). Relative breed contributions to neutral genetic diversity of a comprehensive representation of Iberian native cattle. *Animal* **5**: 1323–1334.
- Eding H, Crooijmans RP, Groenen MA, Meuwissen TH (2002). Assessing the contribution of breeds to genetic diversity in conservation schemes. *Genet Sel Evol* **34**: 613–633.
- Eding H, Meuwissen TH (2001). Marker-assisted estimates of between and within population kinships from genetic marker data for the construction of core sets in genetic conservation schemes. *J Anim Breed Genet* **118**: 141–159.
- Eding H, Meuwissen TH (2003). Linear methods to estimate kinships from genetic marker data for construction of core sets in genetic conservation schemes. *J Anim Breed Genet* **120**: 289–302.
- Fabuel E, Barragán C, Silió L, Rodríguez MC, Toro MA (2004). Analysis of genetic diversity and conservation priorities in Iberian pigs based on microsatellite markers. *Heredity* **93**: 104–113.
- Fisher RA (1930). *The Genetical Theory of Natural Selection*. Oxford University Press: Oxford.
- Food and Agriculture Organization (FAO) (2004). *Measurement of Domestic Animal Diversity – A Review of Recent Diversity Studies*. FAO of the United Nations, Commission on Genetic Resources, Working Group on Animal Genetic Resources: Rome, Italy.
- Food and Agriculture Organization (FAO) (2007). *The State of the World's Animal Genetic Resources for Food and Agriculture*. Commission on the genetic resources for Food and Agriculture Organization of the United Nations: Rome, Italy.
- Gama LT, Martínez AM, Carolino I, Landi V, Delgado JV, Vicente AA et al. (2013). Genetic structure, relationships and admixture with wild relatives in native pig breeds from Iberia and its islands. *Genet Sel Evol* **45**: 18.
- Gandini GC, Ollivier L, Danell B, Distl O, Georgoudis A, Groeneveld E et al. (2004). Criteria to assess the degree of endangerment of livestock breeds in Europe. *Livest Prod Sci* **91**: 173–182.
- García D, Corral N, Canon J (2005). Combining inter- and intrapopulation information with the Weitzman approach to diversity conservation. *J Hered* **96**: 704–712.
- Ginja C, Gama LT, Cortés O, Delgado JV, Dunner S, García D et al. (2013). Analysis of conservation priorities of Iberoamerican cattle based on autosomal microsatellite markers. *Genet Sel Evol* **45**: 1–18.
- Herrero-Medrano JM, Megens HJ, Groenen AM, Bosse M, Pérez-Enciso M, Crooijmans R (2014). Whole-genome sequence analysis reveals differences in population management and selection of European low-input pig breeds. *BMC Genomics* **15**: 601.
- Herrero-Medrano JM, Megens HJ, Groenen AM, Ramis G, Bosse M, Pérez-Enciso M et al. (2013). Conservation genomic analysis of domestic and wild pig populations from the Iberian Peninsula. *BMC Genet* **14**: 106.
- Hoffman I (2010). Climate change and the characterization, breeding and conservation of animal genetic resources. *Anim Genet* **41**: 32–46.
- Lachance J, Tishkoff SA (2013). SNP ascertainment bias in population genetic analyses: why it is important, and how to correct it. *Bioessays* **35**: 780–786.
- Linares V, Linares G, Mendoza G (2011). Caracterización etnozotécnica y potencial carnívero de Sus scrofa 'cerdo criollo' en Latinoamérica. *Sci Agropecu* **2**: 97–110.
- Manunza A, Zidi A, Yeghoyan S, Balteanu VA, Carsai TC, Scherbakov O et al. (2013). A high throughput genotyping approach reveals distinctive autosomal genetic signatures for European and Near Eastern wild boar. *PLoS One* **8**: e55891.
- McGlone J, Pond WG (2003). *Pig Production: Biological Principles and Applications*. Thomson Delmar Learning: New York.
- Oldenbroek JK (1999). *Genebanks and the Conservation of Farm Animal Genetic Resources*. DLO Institute for Animal Sciences and Health: Lelystad, The Netherlands.
- Ollivier L, Alderson L, Gandini GC, Foulley JL, Haley CS, Joosten R et al. (2005). An assessment of European pig diversity using molecular markers: partitioning of diversity among breeds. *Conserv Genet* **6**: 729–741.
- Ollivier L, Foulley JL (2005). Aggregate diversity: new approach combining within- and between-breed genetic diversity. *Livest Prod Sci* **95**: 247–254.
- Park SDE (2001). Trypanotolerance in West African cattle and the population genetic effects of selection. PhD thesis, University of Dublin.
- Piyasatian N, Kinghorn BP (2003). Balancing genetic diversity, genetic merit and population viability in conservation programmes. *J Anim Breed Genet* **120**: 137–149.

- Ponzoni RW (1997). Genetic resources and conservation. In: Piper LR, Ruvinsky A (eds), *The Genetics of Sheep*. CAB International: New York, NY, USA, pp 433–466.
- Ramos AM, Crooijmans RP, Affara NA, Amaral AJ, Archibald AL, Beever JE *et al.* (2009). Design of a high density SNP genotyping assay in the pig using SNPs identified and characterized by next generation sequencing technology. *Plos One* **5**: 4.
- Ramírez O, Burgos-Paz W, Casas E, Ballester M, Bianco E, Olalde I *et al.* (2014). Genome data from a sixteenth century pig illuminate modern breed relationships. *Heredity* **114**: 175–184.
- Ramírez O, Ojeda A, Tomàs A, Gallardo D, Huang LS, Folch JM *et al.* (2009). Integrating Y-chromosome, mitochondrial and autosomal data to analyze the origin of pig breeds. *Mol Biol Evol* **26**: 2061–2072.
- Revidatti MA, Delgado Bermejo JV, Gama LT, Landi Perati V, Ginja C, Alvarez LA *et al.* (2014). Genetic characterization of local Criollo pig breeds from the Americas using microsatellite markers. *J Anim Sci* **92**: 4823–4832.
- Reynolds J, Weir BS, Cockerham CC (1983). Estimation of the coancestry coefficient: basis for a short-term genetic distance. *Genetics* **105**: 767–779.
- Royo LJ, Alvarez I, Fernández L, Pérez-Pardal A, Alvarez-Sevilla A, Godinho R *et al.* (2007). In: Proceedings of the 6th International Symposium on the Mediterranean Pig: 11–13 October 2007; Messina, Italy Nanni Costa L, Zambonelli P, Russo V (eds): *Genetic Characterisation of Celtic-Iberian Pig Breeds Using Microsatellites*. Università di Bologna, Italy, pp 31–34.
- Ruane J (1999). A critical review of the values of genetic distance studies in conservation of animal genetic resources. *J Anim Breed Genet* **116**: 317–323.
- Scandura M, Iacolina L, Apollonio M (2011). Genetic diversity in the European wild boar *Sus scrofa*: phylogeography, population structure, and wild x domestic hybridization. *Mammal Rev* **41**: 125–137.
- Toro M, Barragán C, Óvilo C, Rodríguez J, Rodríguez C, Silió L (2002). Estimation of coancestry in Iberian pigs using molecular markers. *Conserv Genet* **3**: 309–320.
- Toro MA, Fernandez J, Caballero A (2009). Molecular characterization of breeds and its use in conservation. *Livest Sci* **120**: 174–195.
- Velázquez F, Barba C, Pérez-Pineda E, Delgado JV (1998). El cerdo negro criollo cubano: origen, evolución y situación actual. *Arch Zootec* **47**: 561–564.
- Vicente A, Carolino M, Sousa M, Ginja C, Silva F, Martinez A *et al.* (2008). Genetic diversity in native and commercial breeds of pigs in Portugal assessed by microsatellites. *J Anim Sci* **86**: 2496–2507.
- Weir BS, Cockerham CC (1984). Estimating F-statistics for the analysis of population structure. *Evolution* **38**: 1358–1370.
- Weitzman ML (1992). On diversity. *Q J Econ* **107**: 363–405.

Supplementary Information accompanies this paper on Heredity website (<http://www.nature.com/hdy>)