

# The clinical perspective: How to personalise treatment in MS and how may biomarkers including imaging contribute to this?

Patrick Vermersch, Thomas Berger, Ralf Gold, Carsten Lukas, Alex Rovira, Bianca Meesen, Declan Chard, Manuel Comabella, Jacqueline Palace and Maria Trojano

## Abstract

**Background:** Multiple sclerosis (MS) is a highly heterogeneous disease, both in its course and in its response to treatments. Effective biomarkers may help predict disability progression and monitor patients' treatment responses.

**Objective:** The aim of this review was to focus on how biomarkers may contribute to treatment individualisation in MS patients.

**Methods:** This review reflects the content of presentations, polling results and discussions on the clinical perspective of MS during the first and second Pan-European MS Multi-stakeholder Colloquia in Brussels in May 2014 and 2015.

**Results:** In clinical practice, magnetic resonance imaging (MRI) measures play a significant role in the diagnosis and follow-up of MS patients. Together with clinical markers, the rate of MRI-visible lesion accrual once a patient has started treatment may also help to predict subsequent treatment responsiveness. In addition, several molecular (immunological, genetic) biomarkers have been established that may play a role in predictive models of MS relapses and progression. To reach personalised treatment decisions, estimates of disability progression and likely treatment response should be carefully considered alongside the risk of serious adverse events, together with the patient's treatment expectations.

**Conclusion:** Although biomarkers may be very useful for individualised decision making in MS, many are still research tools and need to be validated before implementation in clinical practice.

**Keywords:** Biological markers, disease progression, drug-related side effects and adverse reactions, magnetic resonance imaging, multiple sclerosis, treatment response

Date received: 15 April 2016; accepted: 23 April 2016

## Introduction

Multiple sclerosis (MS) is classically regarded as an idiopathic inflammatory demyelinating and neurodegenerative disease of the central nervous system (CNS). Most prevalence estimates of MS in western countries vary between 25 and 200 per 100,000, with incidences peaking around 30 years of age.<sup>1</sup> The disease is nowadays thrice as common in women than in men and is the leading cause of non-traumatic neurological disability in young adults in western countries.<sup>2</sup>

Because part of the disease process in MS is clinically silent over a long period of time, for example the number of new brain lesions seen on magnetic

resonance imaging (MRI) is substantially greater than the number of relapses occurring over the same period,<sup>3</sup> surrogate markers for disease activity and progression have been identified and have served as outcome measures in clinical trials. At present, the most important para-clinical measures predicting a patient's prognosis are changes in the CNS detected using MRI. In recent years, several prognostic molecular biomarkers have been evaluated as well. In addition, some biomarkers may assist in predicting a patient's treatment response. These markers may be useful to identify poor responders early on and to switch them to an alternative, more effective, therapy before substantial neurological damage has occurred. Alternatively, biomarkers may be used to predict a

Multiple Sclerosis Journal

2016, Vol. 22(2S) 18–33

DOI: 10.1177/  
1352458516650739

© The Author(s), 2016.  
Reprints and permissions:  
[http://www.sagepub.com.uk/  
journalsPermissions.nav](http://www.sagepub.com.uk/journalsPermissions.nav)

Correspondence to:

**P Vermersch**  
University of Lille, CHRU  
de Lille, Lille International  
Research Inflammation  
Center (LIRIC), INSRRM  
U995, FHU Imminent,  
F-59037 Lille, France.  
[patrick.vermersch@univ-  
lille2.fr](mailto:patrick.vermersch@univ-lille2.fr)

**Patrick Vermersch**  
University of Lille, CHRU  
de Lille, Lille International  
Research Inflammation  
Center (LIRIC), INSRRM  
U995, FHU Imminent, Lille,  
France

**Thomas Berger**  
Neuroimmunology and  
Multiple Sclerosis Clinic,  
Medical University of  
Innsbruck (MUI), Innsbruck,  
Austria

**Ralf Gold**  
Department of Neurology,  
St. Josef-Hospital, Ruhr  
University Bochum, Bochum,  
Germany

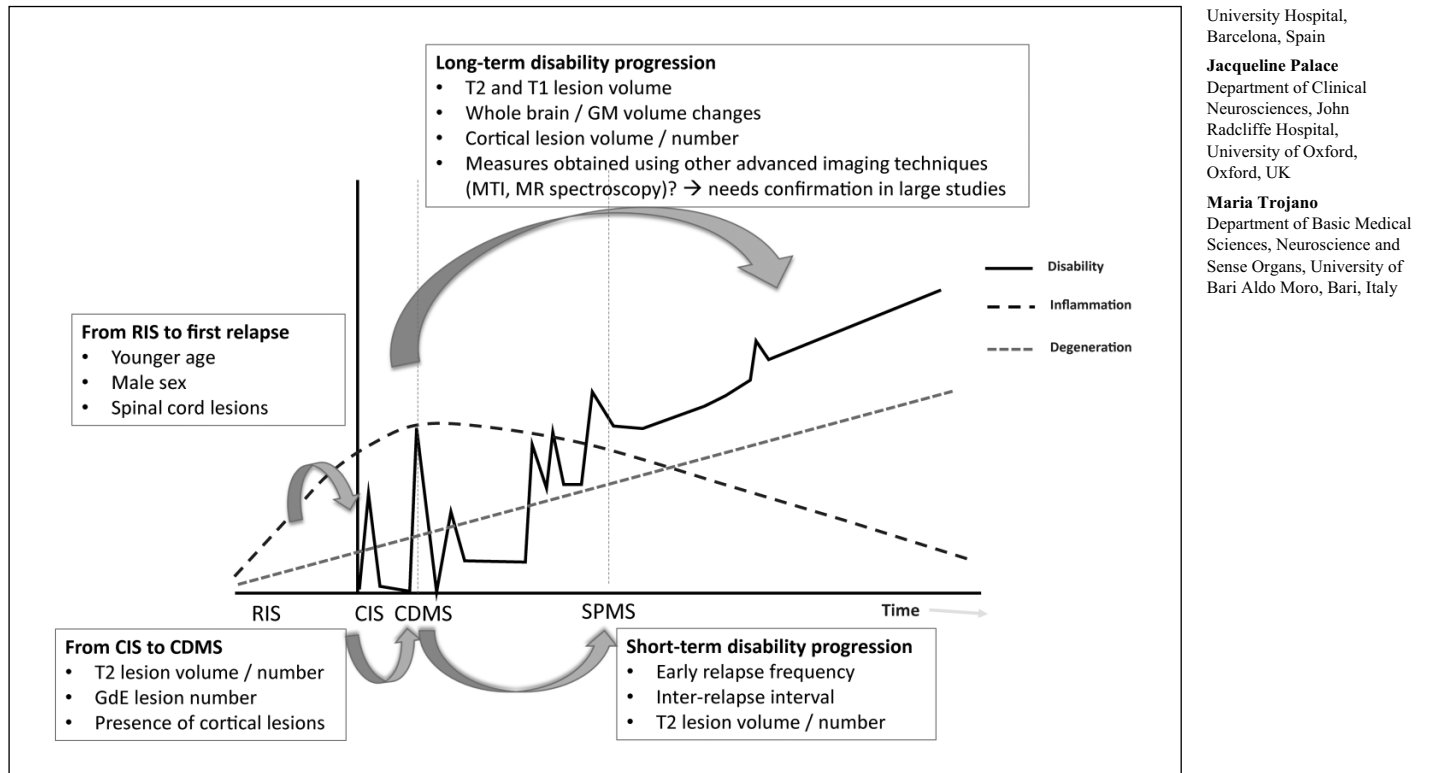
**Carsten Lukas**  
Department of Diagnostic  
and Interventional Radiology,  
St. Josef-Hospital, Ruhr  
University Bochum,  
Bochum, Germany

**Alex Rovira**  
Department of Radiology,  
Vall d'Hebron University  
Hospital, Barcelona, Spain

**Bianca Meesen**  
Managing Director at Ismar  
Healthcare, Lier, Belgium

**Declan Chard**  
NMR Research Unit, Queen  
Square Multiple Sclerosis  
Centre, UCL Institute of  
Neurology, University  
College London, London,  
UK/Biomedical Research  
Centre, University College  
London Hospitals (UCLH),  
National Institute for Health  
Research (NIHR), London,  
UK

**Manuel Comabella**  
Department of Clinical  
Neuroimmunology, Multiple  
Sclerosis Center of Catalonia  
(Cemcat), Vall d'Hebron



**Figure 1.** Schematic overview of risk factors and clinical and MRI biomarkers of progression in MS. GdE: gadolinium-enhancing; GM: grey matter; MR: magnetic resonance; MTI: magnetisation transfer imaging.

patient's risk of developing serious adverse events (SAEs) on treatment with a particular drug.<sup>4,5</sup> It should be noted that biomarkers that have proven valuable on a group level in clinical trials may not be suitable for the evaluation of individuals. In addition, the interval between measurements can (in part) have an impact on the sensitivity to clinically relevant changes.<sup>6</sup>

The current and future potential of biomarkers for predicting the disease course, treatment response and tolerability was discussed during the first and second Pan-European MS Multi-stakeholder Colloquium, which took place on 23–24 May 2014<sup>7</sup> and 15–16 May 2015<sup>8</sup> in Brussels. The goal of these colloquia was to enhance the communication and collaboration between the different stakeholders involved in MS care, including patients and their caregivers, health-care professionals, researchers, regulators and payers. The programmes developed by the chair and scientific committee aimed at prioritising actions needed to improve the quality of and access to care and treatment. At the first Colloquium, after introductory presentations on various subjects by the different stakeholders, the audience was asked to rank priorities from a list of potential action points. The outcome

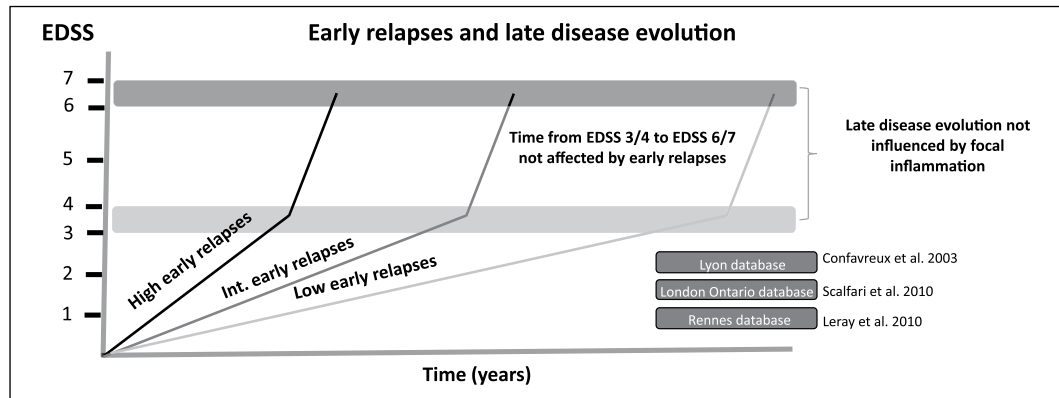
of this polling was used to stimulate further discussions among the speakers, a group of experts in the field and the audience during the first and second Colloquia.

This review summarises the content of the presentations, polling results and discussions related to the use of risk factors and clinical, MRI and other biomarkers in MS and their current and future utility for the individualisation of treatment.

## Predicting disease progression

### *Clinical markers of disease progression*

**Relapses.** Several studies have suggested an association between a higher relapse rate in the first 2–5 years after disease onset and a shorter first inter-relapse interval on the one hand and a more rapid disability progression on the other hand (Figure 1).<sup>9–13</sup> However, this predictive effect seems to disappear once the progressive course starts (e.g. when an Expanded Disability Status Scale (EDSS) score of 3–4 is reached) (Figure 2). Mean times from disease onset, progressive phase onset and an EDSS 3 to reaching EDSS 6, 8 and 10 are strikingly similar for patients



**Figure 2.** Early relapse rate predicts time to progressive MS, but not late disability progression.<sup>10</sup>

with different relapse numbers in the relapsing–remitting (RR) phase.<sup>10</sup> Also in patients with progressive onset MS, superimposed relapses do not appear to affect long-term outcomes.<sup>14</sup> However, an important point that we should consider in evaluating these results is that the main measure of disability (EDSS) used in these studies and in daily clinical practice has significant limitations: EDSS in the 4–7 range is insensitive to change in any functional system other than ambulation. Relapses affecting upper limb or brain stem function or cognition, more prevalent in later phases of the disease, might not affect the EDSS score. These limitations might explain the limited ability of EDSS to detect a delayed impact of relapses on disability progression.<sup>15</sup>

#### *MRI markers of disease progression*

*MRI lesions detected using conventional techniques.* Conventional MRI techniques, such as spin-echo and fluid-attenuated inversion recovery (FLAIR) T2-weighted and unenhanced and contrast-enhanced T1-weighted sequences mainly detect focal lesions or plaques in the brain and spinal cord of MS patients. Classical MRI measures in MS, which can be evaluated using these techniques, are the number and volume of gadolinium-enhancing (GdE) lesions, hyperintense lesions on T2-weighted scans and hypointense ‘black holes’ on T1-weighted scans. The number of GdE lesions has been found to be associated with the risk of future relapse and relapse rate.<sup>16</sup> However, subclinical activity detected using conventional MRI may occur at a 5–10 times higher rate than clinical observations would suggest.<sup>17</sup> Anomalies in the CNS suggestive of MS may also be identified by MRI before there is clinical evidence of the disease. This is referred to as ‘radiologically isolated syndrome’ (RIS). In a retrospective study including 451 subjects with RIS, about one-third of these subjects

had a first clinical event within 5 years of the first brain MRI study (at a mean age of 37.2 years).<sup>18</sup> Lesions within the cervical or thoracic spinal cord were identified as significant predictors for the development of a first clinical event (hazard ratio (HR): 3.08), together with younger age (HR: 0.98, i.e. an estimated risk of developing an event decreasing by 2% for every additional year of age) and male sex (HR: 1.93) (Figure 1).

The presence of GdE lesions and T2 hyperintense lesions is an important diagnostic criterion in MS, because of the established association between the number and volume of these lesions and conversion from clinically isolated syndrome (CIS) to clinically definite multiple sclerosis (CDMS) (Figure 1).<sup>19–22</sup> Despite their association with relapse rate, relatively weak correlations have been reported between conventional lesion metrics and disability progression, as measured using the EDSS.<sup>21,23,24</sup> In patients with relapsing–remitting multiple sclerosis (RRMS), early progression of T2 lesion load appears to predict progression to secondary progressive multiple sclerosis (SPMS) to some extent, but there appears to be little association between the burden of T2 lesions and future disability for EDSS values above 4.5 (Figure 2).<sup>24,25</sup> Chronic T1 hypointense lesions, detected on spin-echo sequences, have shown a better correlation with EDSS than GdE and T2 lesions,<sup>26</sup> and these lesions are believed to reflect severe and irreversible axonal damage.<sup>27</sup> Although potentially clinically relevant, T1-hypointense lesion assessment is still subjective and highly dependent on the type of T1-weighted sequence and field strength.<sup>28</sup> When considering the predictive role of MRI lesions it should be noted that the number of lesions accumulated over time may be a better predictor of future disability than the number of active (GdE) lesions at a single time point.<sup>29</sup> Moreover, the predictive value of active lesions may

be higher in the early RR phase than later in the disease evolution.<sup>25,30</sup>

**Brain atrophy.** Pathophysiological research over the past decades has shown that conventional MRI measures do not tell the full history of MS. Whereas focal inflammation and axonal demyelination in the white matter (WM) seem to be mainly associated with relapses, axonal/neuronal loss is currently believed to be the main driver of irreversible disability progression. These insights have triggered interest in measuring tissue volume loss (atrophy) in the CNS as a marker of neurodegeneration. Several MRI techniques to measure brain volume loss (and in case of sustained volume loss as per definition: atrophy) such as segmentation-based or registration-based methods have been introduced in the past. Segmentation-based methods measure global or regional brain volume (e.g. brain parenchymal fraction, WM fraction, grey matter (GM) fraction, normalised brain volume) at a single time point. Registration-based methods measure brain volume at two time points, in order to calculate the percentage brain volume change (PBVC), and are most suitable for evaluating global brain volume changes,<sup>31</sup> but are not usually designed to analyse regional volume changes over time.<sup>32,33</sup>

Studies have shown that brain atrophy affects the entire brain in MS, including WM and GM, and starts very early in the disease course.<sup>34,35</sup> Although some studies suggested that brain atrophy escalates with increasing disease stage, this was not confirmed in a large MAGNIMS (Magnetic Resonance Imaging in Multiple Sclerosis) study when data were corrected for baseline normalised brain volume.<sup>36,37</sup> An association between increasing early brain tissue volume loss and increasing long-term disability progression has been shown in several studies (Figure 1).<sup>38–41</sup> GM volume was found to be a stronger predictor of clinical disability than WM volume.<sup>39</sup>

Measuring brain atrophy is challenging because the change in volume over time is relatively small. In MS patients, brain atrophy occurs at a rate of 0.5%–1.3% per year, compared with 0.1%–0.3% per year reported for healthy subjects.<sup>34,42</sup> Although the estimation of brain atrophy seems to be an important prognostic marker, its implementation in the clinical workflow is limited by several factors.

As brain atrophy is not necessarily linear, progression in individuals is hard to predict. Also, no common agreement on a single measurement technique to be used in clinical research or clinical practice currently

exists. Differences between techniques limit direct comparison between results. Furthermore, several other confounding factors must be considered when evaluating disease progression based on brain volume loss/atrophy, including image acquisition and quality (e.g. imperfect skull extraction and outlining, imprecise registration, issues due to patient movement), the effect lesions can have on tissue segmentation (e.g. due to classification of T1 hypointense lesions as GM), pseudo-atrophy (reduction in inflammation due to disease-modifying treatment), change in brain water content due to hydration status or steroid use or even diurnal fluctuations, and other factors such as cardiovascular disease, smoking, high alcohol consumption and genetic factors.<sup>43,44</sup> Although brain atrophy measures are very valuable for group analysis, both biological and technical variability need to be improved to make them suitable for individual analysis.

**Advanced MRI measures.** The introduction of new advanced imaging techniques, including sensitive techniques to quantify diffuse damage or metabolic or functional changes in tissue appearing normal on conventional MRI scans, has considerably improved the detection of pathological changes in MS.<sup>17</sup>

A promising new sequence for diagnostic set-up and follow-up is double-inversion recovery (DIR). DIR has considerably improved the ability to detect cortical lesions in patients with MS. Studies have shown accumulation of cortical lesions over time and correlations with clinical and cognitive dysfunction.<sup>42</sup> The presence of at least one cortical lesion has been found to be associated with an increased risk of conversion from CIS to CDMS.<sup>45</sup> Cortical lesion volume and number have also been found to independently predict future disability accumulation in RRMS, SPMS and primary progressive multiple sclerosis (PPMS) patients (Figure 1).<sup>46,47</sup> However, about 80% of GM lesions still remain undetected with this technique.<sup>48</sup> DIR very rarely detects subpial cortical lesions,<sup>48</sup> which is the most abundant and specific cortical lesion type seen in histopathological work. There is no common sequence recommendation for cortical lesion detection, and DIR inter-rater reliability of cortical lesion scoring using consensus guidelines was found to be low, with a complete agreement on only ~20% of lesions between readers.<sup>49</sup>

Magnetisation transfer imaging (MTI) measures correlate with demyelination, remyelination and axonal loss.<sup>50</sup> MTI variables in GM and normal-appearing WM have been found to independently predict future disability (EDSS) progression in the long

term (3–8 years) in patients with RRMS, SPMS and PPMS (Figure 1).<sup>51–53</sup> However, MTI is time-consuming and its use in individuals is limited by the variability across sites (as the results depend on the method used) and the lack of normative values, which is a problem common to many current quantitative MRI techniques.

Proton magnetic resonance (MR) spectroscopy can provide information about metabolic changes in normal-appearing brain tissue and focal lesions. Reductions of *N*-acetyl-aspartate (NAA) levels (suggestive of neuroaxonal damage) are partly reversible and an association between greater increases in NAA levels after spinal relapse and greater recovery has been described.<sup>54</sup> However, MR spectroscopy is time-consuming and has a greater biological variability than other structural methods, and measures are method- and scanner-specific. Therefore, this technique is currently not considered suitable for use in multi-centre studies.

#### *Optical coherence tomography*

MS patients typically show thinning of particularly the innermost layers of the retina, even without a history of optic neuritis (ON).<sup>55</sup> Some studies have shown an inverse relationship between inner nuclear layer thickness and EDSS (progression).<sup>56,57</sup> Recently, a strong correlation between ganglion cell/inner plexiform atrophy and whole-brain, especially GM, atrophy, was established, particularly in patients with progressive MS, suggesting that it mirrors underlying disease progression.<sup>58</sup> Despite promising results, more large longitudinal studies are needed to evaluate the prognostic value of optical coherence tomography (OCT) in MS. A limitation of OCT is that measurements are affected by a history of ON, lesions elsewhere in the visual pathway and non-MS ocular conditions.<sup>55</sup> In addition, magnitudes of annual thinning of retinal layers are smaller than the variability between measurements.<sup>55</sup>

#### *Evoked potentials*

A number of studies have suggested that multimodal evoked potentials (EPs) (nerve latencies) may be valuable for monitoring and predicting disability in MS patients.<sup>59</sup> Although their diagnostic value is considered poor compared with MRI,<sup>59</sup> several studies have shown correlations between EP measures and (future) disability.<sup>59–61</sup> However, its use in clinical practice requires standardisation within and between laboratories.

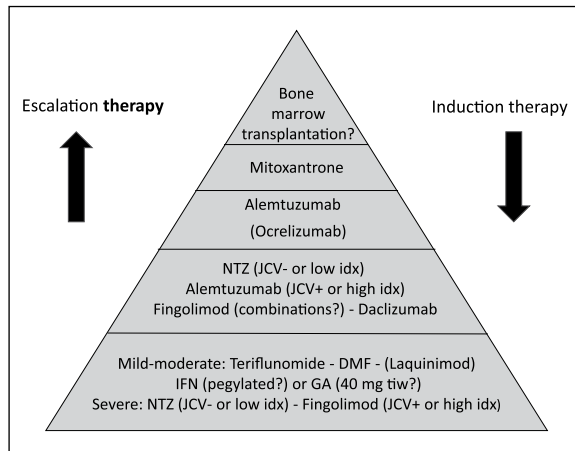
#### *Molecular biomarkers*

Apart from MRI, several molecular biomarkers have been identified for diagnosing MS and for monitoring and predicting disability progression.

The presence of immunoglobulin G (IgG) oligoclonal bands (OCB) and/or an elevated IgG index in the cerebrospinal fluid (CSF) support the *diagnosis of MS* in patients suspected to have demyelinating disease, but they do not contribute to proof of MS in the 2010 McDonald criteria. The best validated molecular biomarkers that predict *conversion to CDMS in patients with CIS* are the presence of IgG OCB<sup>62,63</sup> and the IgG index<sup>62</sup> in the CSF. Recently, serum auto-antibodies directed against the potassium channel KIR4.1 have been suggested as a candidate biomarker for the *diagnosis of MS*.<sup>64</sup> They are already detectable in the early stages of MS and have an excellent specificity but low sensitivity. However, a recent validation study could not replicate this finding in independent cohorts.<sup>65</sup>

Another CSF biomarker with strong evidence is Chitinase 3-like 1 (CHI3L1), which is expressed on monocytes and microglial cells and has been linked to astrocyte activation. CHI3L1 has not only been shown to predict conversion to CDMS<sup>66,67</sup> but also to predict more rapid disability progression.<sup>66,67</sup> Recently, the level of vitamin D in blood has been suggested as a candidate biomarker for *conversion to CDMS*<sup>62,68</sup> and *more rapid disability progression*.<sup>68,69</sup> In a study including patients with CIS, who were mainly treated with interferon (IFN) $\beta$ -1b, low serum levels of 25-hydroxyvitamin D (a marker of vitamin D status) predicted long-term clinical and MRI activity.<sup>68</sup> Furthermore, lower serum 25-hydroxyvitamin D levels were also associated with lower MRI activity in RRMS patients treated with IFN $\beta$ -1b in the prospective BEYOND (Betaferon Efficacy Yielding Outcomes of a New Dose) study.<sup>69</sup> However, these data should be further confirmed by other investigators before vitamin D can be used as a valid biomarker. In the future, biomarkers predicting conversion to CDMS may become useful in the decision which patients with CIS could benefit from early treatment initiation.

As mentioned above, a number of molecular biomarkers for *predicting future disease activity* have been suggested as well, including CHI3L1 and vitamin D levels. In addition, the neurofilament (NF) levels in the CSF and blood have been suggested as biomarker for predicting disability progression. NFs are major axonal cytoskeleton proteins, consisting of a light chain (NFL), an intermediate chain (NFM) and a heavy chain (NFH). NFL and NFH concentrations in



**Figure 3.** Treatment algorithm depicting the difference between escalation and induction therapy.

BG12: dimethyl fumarate; GA: glatiramer acetate; idx: JCV antibody index; IFN: interferon; JCV: John Cunningham virus; NTZ: natalizumab; tiw: three times weekly.

CSF are used in clinical practice as surrogate endpoints of neuroaxonal damage. Indeed, NFL levels seem to correlate with acute axonal damage,<sup>70</sup> while NFH levels may reflect chronic axonal damage and may be more strongly associated with disability progression.<sup>71,72</sup> Furthermore, serum NFL levels appear to correlate with MRI activity and disability scores and may present an easily accessible biomarker predicting disability progression.<sup>73</sup>

Although these molecular biomarkers may gain importance in the future, their integration into clinical practice requires further evaluation. Longitudinal studies in large cohorts of patients are needed to better assess the natural history of MS in relation to baseline levels of these biomarkers.

### Predicting treatment response and tolerability

Initiation of the right drug at the right time is a crucial goal in MS in order to minimise further inflammation, neurodegeneration and resulting irreversible disability progression.<sup>74</sup> The majority of patients with RRMS will start with a first-line disease-modifying drug (DMD) with a moderate efficacy but a good safety profile (e.g. IFN $\beta$ , glatiramer acetate (GA), teriflunomide, dimethyl fumarate (DMF)).<sup>75</sup> In those who fail to respond to these agents, this is followed by second-, third-, fourth- or even fifth-line treatments (Figure 3). For each treatment step, drug efficacy may increase along with the associated risk of SAEs. However, for patients with a high disease activity, starting therapy with highly effective but aggressive therapies such as natalizumab (NTZ) or alemtuzumab (ATZ) (also

referred to as induction therapy) may be appropriate to rapidly reduce disease activity. Once disease control has been achieved, therapy can be scaled back to better tolerated – but potentially less efficacious – drugs for long-term maintenance.<sup>75</sup> Current experience with induction therapy is limited, particularly its immunogenic effect in the long term, and not tested against escalation strategies in randomised controlled trials.

As not all patients will sufficiently respond to first-line treatment, and conversely not all patients will develop SAEs upon treatment with highly aggressive drugs, it would be very useful to be able to predict a patient's likely treatment response and risk of SAEs. In this way, poor responders can be switched to an alternative therapy early on, before substantial neurological damage has occurred, and patients can be spared from potential SAEs associated with a particular drug, such as progressive multifocal leukoencephalopathy (PML) upon treatment with NTZ or autoimmune disorders upon treatment with ATZ.<sup>4,5</sup> Therefore, current research is focusing on the identification of clinical, imaging, immunological and genetic biomarkers that may help individualise treatment.

### Response to first-line treatments

**Response to IFN $\beta$  and GA.** Regarding the injectable first-line DMDs IFN $\beta$  and GA, direct comparative studies have not shown superiority of one drug over another in terms of efficacy (BEYOND,<sup>76</sup> REGARD<sup>77</sup> (REbif vs Glatiramer Acetate in Relapsing Multiple Sclerosis Disease)). Moreover, these trials do not give any indication on the most appropriate first-line treatment choice at the patient-specific level. Additionally, surrogate markers are needed to predict which patients will respond to first-line DMDs. Several studies have shown that, in patients with RRMS, high disease activity, that is, high frequency of relapse, high rate of disability progression and/or high number of MRI lesions at baseline or in the first year of treatment, may predict (mid- and long-term) failure to IFN $\beta$  and GA.<sup>5,78–83</sup>

Next to these clinical and MRI measures, the titre of neutralising antibodies (NABs) against IFN $\beta$  has been established as a clinically useful predictor of poor treatment response.<sup>84</sup> Indeed, NABs reduce the therapeutic effect of IFN $\beta$  on relapse rate and MRI lesion activity.<sup>84,85</sup> Therefore, for patients with sustained high-titre NABs consideration should be given to DMDs other than IFN $\beta$ .<sup>84</sup> In addition, other immunological biomarkers have been suggested to predict response to IFN $\beta$  in patients with RRMS, including

several chemokines and cytokines.<sup>4,72</sup> However, for most of them, for example, serum interleukin-17F (IL-17F), the predictive value is still highly debated.<sup>86,87</sup> Moreover, for those markers that have already been validated, usefulness in clinical practice still needs to be demonstrated.<sup>72</sup>

Furthermore, many pharmacogenomics studies have tried to identify genetic variants that may predict response to IFN $\beta$  or GA. So far, two genome-wide association studies have suggested a role for *GPC5*, glutamate receptors and *ADAR* in response to IFN $\beta$ .<sup>88</sup> Very few studies have evaluated the pharmacogenomics of response to GA.<sup>88–90</sup> To bring pharmacogenomics from academic research to clinical practice, a joint effort between academy and industry is necessary.<sup>91</sup> A large-scale pharmacogenomics study in GA-treated RRMS patients, including consenting patients from the FORTE (Feasibility of Retinoids in the Treatment of Emphysema;  $N=604$ ) and the GALA (Glatiramer Acetate Lowfrequency Administration) studies ( $N=1158$ ), is currently ongoing.<sup>92</sup> An 11-single nucleotide polymorphisms (SNP) signature for GA response was identified in the GALA study and validated in the FORTE study. This multi-SNP signature may be able to predict which GA-naive RRMS patients will be high responders, exhibiting annualised relapse rate (ARR) reductions significantly higher than the average response ( $\approx 33\%$ ) reported in clinical trials.<sup>92</sup> The predictive value of this multi-SNP signature is being validated in an independent cohort.

**Response to oral first-line DMDs.** Due to the development of several oral DMDs, the therapeutic landscape of MS has considerably changed over the last decade; first-line treatment choice has even become more complex. So far, there are no efficacy data showing superiority of the new oral drugs (e.g. teriflunomide, DMF) over IFN $\beta$  or GA. Indeed, the CONFIRM (Comparator and an Oral Fumarate in Relapsing–Remitting Multiple Sclerosis) study was designed to show superiority or non-inferiority of DMF (twice or three times daily) versus placebo, and not versus GA, which was only added as a reference comparator.<sup>93</sup> Similarly, the TENERE (TERifluNo-midE and REbif) study did not show a statistical difference in time to treatment failure between teriflunomide (7 or 14 mg) and IFN $\beta$ -1a.<sup>94</sup> Thus, again, these data do not facilitate the personalised treatment decision between oral and injectable first-line DMDs and biomarkers are needed to predict the patient's treatment response to DMF or teriflunomide. However, to our knowledge, no such biomarkers have been described yet.<sup>72,95</sup> It remains to be investigated whether new imaging techniques such as MTI and diffusion tensor

MRI, OCT or positron emission tomography may be useful for this purpose.<sup>95</sup>

#### Response to second-line treatments

A promising biomarker which objectively reflects response to second-line treatments is the level of NFLs in the CSF.<sup>72,95</sup> NFL concentration in CSF can serve as surrogate endpoint of neuroaxonal damage<sup>71,72</sup> and thus as surrogate endpoint for treatment efficacy. Although NFL levels in CSF were shown to be reduced upon treatment with NTZ,<sup>96,97</sup> fingolimod,<sup>98</sup> mitoxantrone or rituximab,<sup>99</sup> the predictive value on individual patients is very modest. A potential disadvantage of this biomarker is the need for a lumbar puncture to collect CSF. However, a recent study has shown reduced serum levels of NFL antibodies in NTZ-treated RRMS patients, suggesting that they may serve as a biomarker of treatment efficacy as well.<sup>100</sup> However, their usefulness still needs to be confirmed in clinical practice.

Biomarkers that may predict response to second-line drugs, such as SNPs in the ABC transporter genes for mitoxantrone,<sup>101</sup> are still in the exploratory or validation phase.<sup>72</sup>

#### Tolerability

An established immunological biomarker to predict a patient's risk of SAEs is the presence of anti-John Cunningham virus (JCV) antibodies (anti-JCV-Abs) in serum.<sup>4,72</sup> In patients treated with NTZ, positive anti-JCV-Ab status, longer duration of treatment with NTZ and prior immunosuppressive treatment were shown to be associated with an increased risk of PML.<sup>102</sup> Based on these three parameters, a risk stratification algorithm was developed to counsel patients treated with or considering treatment with NTZ on their risk of PML. Anti-JCV-Ab positive patients with no prior use of immunosuppressants may even be further stratified according to their anti-JCV-Ab index, which is a corollary to anti-JCV-Ab titre. Patients whose anti-JCV-Ab index is more than 1.5 and whose treatment duration is longer than 24 months, have been shown to have a substantially greater risk of PML.<sup>103</sup> Thus, they should be encouraged to switch to an alternative drug or undergo strict monitoring (including frequent MRI scanning) to detect PML if NTZ is not discontinued.<sup>104</sup>

Similarly, patients developing secondary autoimmunity (autoimmune thyroid disease, idiopathic thrombocytopenic purpura) following treatment with ATZ were shown to have twofold greater pre-treatment

**Table 1.** Standardised brain MRI acquisition protocol for an optimised follow-up of MS patients developed by MAGNIMS.<sup>106</sup>

<p>Baseline evaluation</p> <ul style="list-style-type: none"> <li>• Mandatory sequences <ul style="list-style-type: none"> <li>◦ Axial proton density or T2-FLAIR/T2-weighted</li> <li>◦ Sagittal 2D or 3D T2-FLAIR</li> <li>◦ 2D or 3D contrast-enhanced T1-weighted</li> </ul> </li> <li>• Optional sequences <ul style="list-style-type: none"> <li>◦ Unenhanced high-resolution isotropic 3D T1-weighted</li> <li>◦ 2D and/or 3D double-inversion recovery (DIR)</li> <li>◦ Axial diffusion-weighted imaging</li> </ul> </li> </ul> <p>Follow-up examinations</p> <ul style="list-style-type: none"> <li>• Mandatory sequences <ul style="list-style-type: none"> <li>◦ Axial proton density or T2-FLAIR/T2-weighted</li> </ul> </li> <li>• Highly recommended sequence <ul style="list-style-type: none"> <li>◦ 2D or 3D contrast-enhanced T1-weighted</li> </ul> </li> <li>• Optional sequences <ul style="list-style-type: none"> <li>◦ Unenhanced high-resolution isotropic 3D T1-weighted</li> <li>◦ 2D and/or 3D DIR</li> <li>◦ Axial diffusion-weighted imaging</li> </ul> </li> </ul> <p>MRI: magnetic resonance imaging; MS: multiple sclerosis; MAGNIMS: Magnetic Resonance Imaging in Multiple Sclerosis; FLAIR: fluid-attenuated inversion recovery; 2D: two-dimensional; 3D: three-dimensional.</p>
--

serum levels of IL-21. The original IL-21ELISA kit, containing antibodies from ascites, has been recently withdrawn in order to switch to the more ethical cell culture-based antibody enzyme linked immunosorbent assay (ELISA) kits.<sup>105,106</sup> However, a recent study has shown that the currently available IL-21 kits have little or no predictive value for the risk to develop secondary autoimmunity on ATZ treatment.<sup>106</sup>

Finally, increased risk of developing secondary acute promyelocytic leukaemia (sAPL) after treatment with mitoxantrone was suggested to be linked to genetic variants in DNA repair and drug-metabolising enzymes (*BRCA2*, *XRCC5*, *CYP3A4*), resulting in impaired detoxification of chemotherapy or inefficient repair of drug-induced genetic damage.<sup>107,108</sup> More research efforts are needed to identify other biomarkers that may predict drug tolerability at the patient-specific level.

#### *Implementation of biomarkers in clinical routine*

MRI measures remain the most important biomarkers for diagnosis and routine follow-up of patients with MS in clinical practice. Brain WM and GM lesion volumes and brain atrophy measures all correlate with disability scores, and can be undertaken using images that can be acquired with all conventional clinical MRI scanners. Beyond the role of WM lesions in the diagnosis of MS,

it has yet to be determined how MRI and molecular biomarkers can be usefully integrated into patient-specific measures for routine use in clinical practice.

An important issue that hampers implementation of MRI in multifactorial decision models and their integration into the routine clinical workflow is the lack of standardised MRI protocols for monitoring disease evolution, particularly for patients receiving DMDs. A standardised basic MRI acquisition protocol should be simple and feasible, robust, fast (around 30 minutes), scanner vendor-independent, field strength-independent, and supported by national and international scientific societies, payers and pharmaceutical companies. A standardised MRI acquisition protocol for the diagnosis and follow-up of MS patients has been recently developed by MAGNIMS network (Table 1).<sup>109</sup> European and national MS as well as neurological and (neuro)radiological societies can play an important role in the implementation of this protocol by supporting its use in clinical practice. As differences in MRI outcomes between centres may be in the same range or even exceed yearly changes due to disease progression or differences between placebo and treatment groups observed in clinical trials, ideally the same MRI machine and protocol should be used in the same patient for as many years as is feasible.

Integration of MRI into routine clinical practice also requires further automation of measurements and



evaluation.<sup>110</sup> There is need for fully automated pipelines to perform high-quality cross-sectional/longitudinal volumetric analysis, with automated detection and filling of lesions (to lessen their confounding effect on atrophy measures). These tools should be integrated in all major MR vendors' post-processing software and allow transfer of information into Picture Archiving and Communication Systems (PACS). Although outsourcing of specialised MRI analysis to dedicated companies may be a good solution for the short term, the pipelines should eventually become integrated into clinical routine allowing fast interpretation of images by (neuro)radiologists.

Not only the MRI protocols but also the way MRI results are reported by radiologists should be standardised and combine conventional (written) and structured reports. Dedicated teaching courses for radiologists on MRI standards in MS and interpretation of images could be conducted on a regular basis in order to deepen skills and knowledge of MRI in MS. This will facilitate and improve the communication between radiologists and clinicians and support analysis for research and decision making. In the future, certification by, for example, the European Committee for Treatment and Research in Multiple Sclerosis (ECTRIMS) or the European Society of Neuroradiology (ESNR) of centres and radiologists/neuroradiologists that fulfil the minimum technical requirements, have adequate quality control programmes and use standardised protocols, may help to accelerate harmonisation.

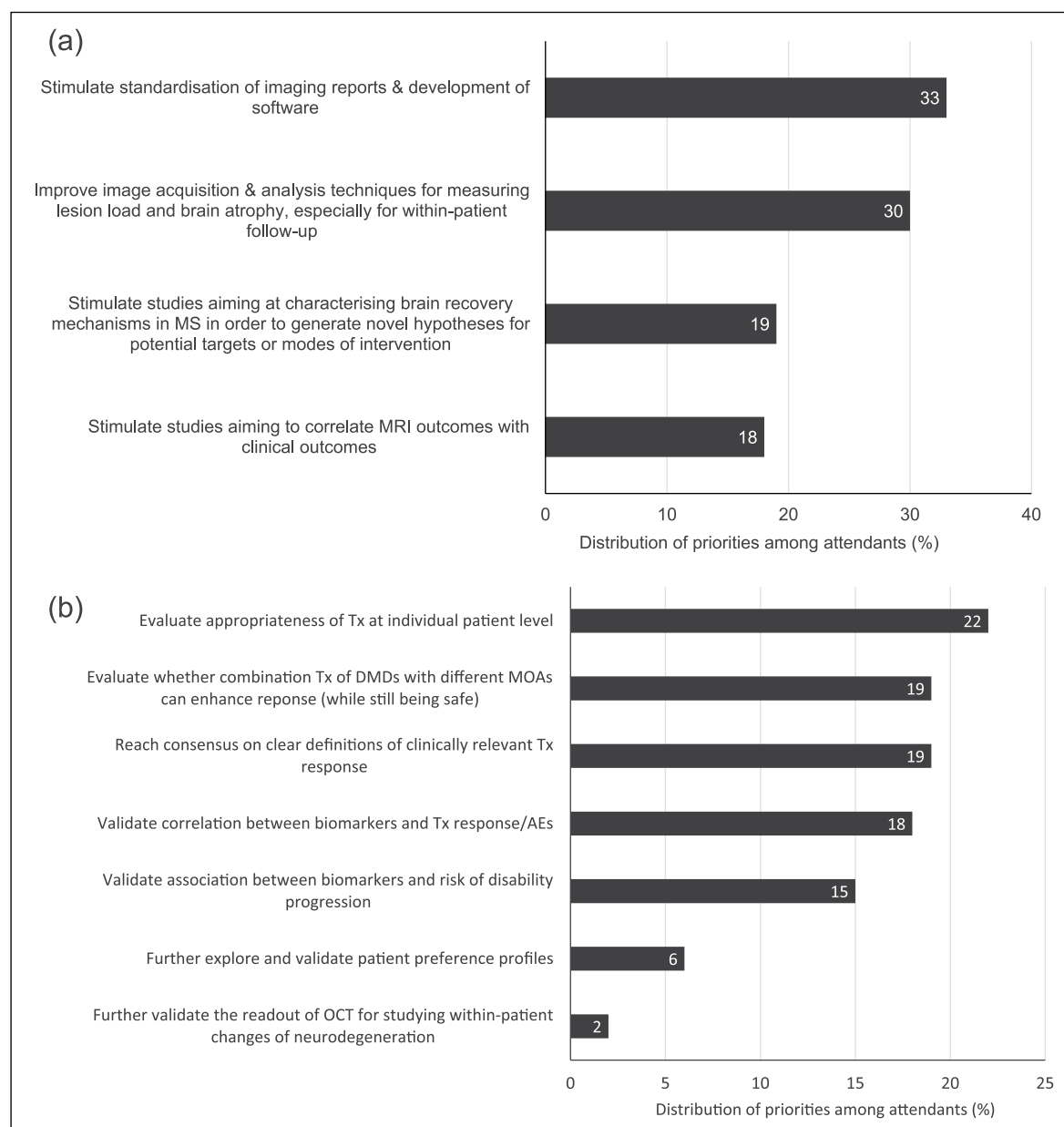
One of the most important prerequisites for the successful implementation of routine MRI evaluations (e.g. including atrophy quantification) into clinical practice is that payers (patients, drug companies, private insurance) and health authorities recognise its clinical value. To achieve this, the identification of the most robust MRI biomarkers for disease evolution and treatment response in the individual patient, definitions and thresholds for MRI activity, and their integration into patient risk stratification algorithms are crucial. Today, most existing data regarding markers of treatment response are based on studies with IFN $\beta$  and may not apply to other DMDs that have different modes of action.<sup>5,78–80</sup> Therefore, more research is warranted. As some DMDs may take up to 6 months to become effective (e.g. GA), an additional baseline scan at 3–6 months after initiation of treatment is recommended to adequately analyse changes over time and to minimise the pseudo-atrophy effect.

The current limitations of existing MRI techniques for use in clinical practice and the need for

standardised protocols were confirmed by the polling results at the first Pan-European MS Multi-stakeholder Colloquium. Indeed, when attendants were asked to rank priorities in MRI research, stimulating standardisation of imaging reports and development of software was considered highest priority, followed by improvement of image acquisition and analysis techniques for patient follow-up (Figure 4). The opinions of clinicians about these issues were further explored in an online questionnaire about the optimisation of imaging/MRI for use in clinical practice undertaken in preparation for the second MS Multi-stakeholder Colloquium. Among the 143 respondents of this questionnaire, mostly neuroradiologists (70%), 77.8% indicated that they already used a standardised MRI protocol for MS patients in their practice. The majority (74.1%) indicated that incorporation of measures of brain volume loss in clinical practice would be valuable. Over 80% partly or fully agreed that the development of simplified but robust techniques should be accelerated in order to allow radiologists on site to perform measurements of brain volume loss themselves and to report directly to the neurologist. In addition, 57.3% of the respondents indicated that reimbursement of MRI analysis in MS should in the first place be obtained from insurance companies and public health organisations.

#### *How to integrate the patient's treatment expectations into individualised decision making?*

In order to take a patient-tailored treatment decision, it is not only important to predict the patient's treatment response and his or her risk of developing SAEs, but also to consider the patient's treatment expectations. Physicians' concerns and their willingness to accept SAEs in return for improved drug efficacy may differ substantially from patients' preferences.<sup>111</sup> In general, physicians may be more concerned about the physical manifestations of MS, while patients may be more worried about less tangible domains such as mental health, role limitations due to emotional problems and vitality. However, delaying disability progression remains the most important treatment expectation for patients, being more important than preventing SAEs and decreasing relapse rate (Figure 5).<sup>112,113</sup> Hence, it is not surprising that patients might be more willing to accept SAEs in return for a reduced risk of disability progression than physicians.<sup>114</sup> Given these differences in treatment perspectives between patients and physicians, neurologists should strongly encourage their patients to formulate their own values and preferences regarding their medical care.<sup>115</sup> In addition, healthcare providers have a duty to ensure patients



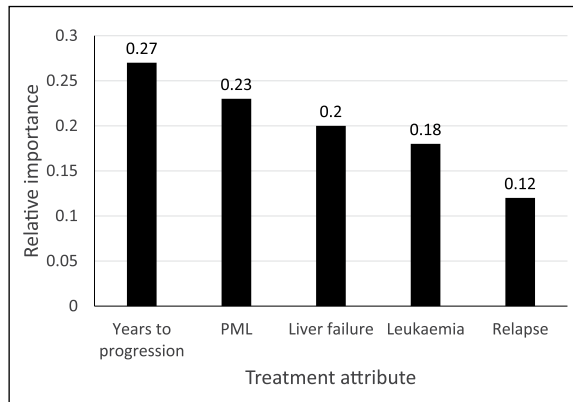
**Figure 4.** Polling results from the first Multi-stakeholder MS Colloquium showing priorities in (a) MRI research and implementation in clinical practice and (b) individualised treatment of MS. The x-axis shows percentages of points for each option (3 points for the first priority, 2 points for the second priority and 1 point for the third priority) versus the total number of points of all options together.

AEs: adverse events; DMDs: disease-modifying drugs; MOAs: mechanisms of action; OCT: optical coherence tomography; Tx: treatment.

understand the complex information given to them. Patient preferences should be carefully considered, together with the available evidence on efficacy and tolerability for each treatment option, the patient's predicted chance of treatment response and his or her predicted risk of SAEs. In this way, we can progress to personalised or patient-tailored decision making, choosing the treatment option that best matches the patient's treatment expectations, with a

good balance between desired efficacy, tolerability and quality of life improvement.

The importance of tailoring treatment for MS to each individual patient also became clear from the polling among the attendants of the first Pan-European MS Multi-stakeholder Colloquium. When participants were asked to rank key issues regarding personalised treatment, evaluating the appropriateness of treatment



**Figure 5.** Delaying disability progression is the most important treatment attribute for patients with multiple sclerosis (MS). A total of 651 patients with MS (mean age: 47 years) with a broad range of disability levels were asked to choose between hypothetical pairs of treatment alternatives with varying levels of clinical efficacy and associated risks. From these trade-off tasks, the relative importance of each attribute was determined. Reproduced from Johnson et al.<sup>112</sup> with kind permission from Springer Science and Business Media.

at individual patient level turned out to be the key priority (Figure 4). In contrast, further exploring and validating patient preferences was not frequently ranked among the top three priorities. This finding suggests that across stakeholders' awareness integrating of individual patient's treatment expectations into decision making in MS still needs to be improved.

### Conclusion

Biomarkers may be very useful tools for individualised decision making in MS. They may assist in diagnosing MS, predicting and monitoring disability progression, and in predicting a patient's treatment response and risk of SAEs. In current clinical practice, MRI markers are still the most important biomarkers for diagnosis and routine follow-up of patients with MS, while they may also help to predict response to IFN $\beta$  or GA. In addition to clinical and MRI markers, several molecular biomarkers have been identified as well, with the level of NFs in CSF being among the most promising ones, both to predict disease progression and to monitor treatment response.

Although biomarkers can theoretically be used to individualise treatment of patients with MS, their implementation in the clinical decision model currently remains very limited. The validation of biomarkers is a long (it can take 5–15 years before a potential biomarker has been validated for use in

clinical practice) and complicated process, requiring replicated evidence of correlation with clinical measures and evaluation of effectiveness, cost-effectiveness, and predictive accuracy in clinical trials and real-life clinical practice.<sup>116</sup> In addition, implementation of biomarkers into the decision model requires regulatory approval, reimbursement agreements, ability to interpret and use the results to take decisions, acceptance in clinical guidelines and patient and clinician acceptance. The implementation of a standardised MRI protocol for monitoring disease evolution would be an important first step towards a better evaluation of MS patients in the near future. Such efforts should be accompanied by dedicated training courses on this subject to maintain a high level of competence. In addition, further development and evaluation of automated measurements that could easily be integrated into the clinical workflow should be fostered to improve practicability of such measurements and facilitate serial analysis and comparison across centres. In addition, more research is needed to discover new clinical, imaging, genetic and immunological biomarkers, to validate new and existing biomarkers and to implement them in clinical practice. Finally, patients' preferences should be actively integrated into the decision-making process. In order to come to a patient-tailored treatment decision, the patient's predicted risks of disability progression, treatment response and SAEs with a particular treatment should be carefully considered together with his/her treatment expectations. This will ultimately help neurologists to optimise drug choices for each individual patient at the right moment during their disease course.

### Acknowledgements

The authors thank Ismar Healthcare, Lier, Belgium for providing assistance with the writing of the article based on the content of the presentations given at the Colloquia and the outcome of the related discussions and voting polls.

### Declaration of Conflicting Interests

The author(s) declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: Patrick Vermersch received honoraria for advisory boards and consulting from Biogen Idec, Teva, Genzyme-Sanofi, Bayer Schering, Merck Serono, Novartis and Almirall and has received grant for research from Bayer Schering, Merck Serono and Biogen Idec. Thomas Berger has received honoraria for lecturing and consulting from Almirall, Bayer Schering, Biogen, Genzyme-Sanofi, Merck, Novartis, Ratiopharm, UCB and Teva. His institution has

received unrestricted research grants from Biogen, Novartis and Teva, and payments for antibody assays (AQP4 and anti-neuronal antibodies) and for AQP4 antibody validation experiments organised by Euroimmun. Ralf Gold has received honoraria, consultant fees or other support from Baxter, Bayer Schering, Biogen Idec, CLB, Behring, Genzyme, Merck Serono, Novartis, Talecris, Teva and Wyeth. Carsten Lukas has received consulting and speaker's honoraria from Biogen Idec, Bayer Schering, Novartis, Sanofi, Genzyme and Teva, and has received research scientific grant support from Bayer Schering, Teva and Merck Serono. He holds an endowed professorship supported by the Novartis Foundation. Alex Rovira has received honoraria or consultation fees from Biogen Idec, Novartis, Genzyme, and OLEA Medical and fees for participating in company sponsored speaker's bureaus for Bayer, Genzyme, Sanofi-Aventis, Bracco, Merck Serono, Teva, OLEA Medical, Stendhal, Novartis and Biogen Idec. Bianca Meesen is the Managing Director of Ismar Healthcare who received support from Teva Pharmaceuticals Europe for facilitating the organisation of the colloquium and providing writing assistance. Declan Chard has received grants/research supports from the Multiple Sclerosis (MS) Society of Great Britain and Northern Ireland and from the University College London Hospitals (UCLH)/University College London (UCL) National Institute for Health Research (NIHR) Biomedical Research Centre; has received honoraria (paid to his employer) from Ismar Healthcare NV, Swiss MS Society, Serono Symposia International Foundation (now Excemed), Merck, Bayer and Teva for faculty-led education work; Teva for advisory board work; meeting expenses from Merck, Teva, Novartis and the National MS Society; and has previously held stock in GlaxoSmithKline. Manuel Comabella has received compensation for consulting services and speaking honoraria from Bayer Schering Pharma, Merck Serono, Biogen Idec, Teva Pharmaceuticals, Sanofi-Aventis, Genzyme and Novartis. Jacqueline Palace is partly funded by highly specialised services to run a National congenital myasthenia service and a neuromyelitis service. She has received support for scientific meetings and honorariums for advisory work from Merck Serono, Biogen Idec, Novartis, Teva, Chugai Pharma, Alexion and Bayer Schering, and unrestricted grants from Merck Serono, Novartis, Biogen Idec, Teva, Genzyme and MS Society. Her hospital trust receives funds for her role as clinical lead for the RSS, and she has received grants from the MS society and Guthrie Jackson Foundation for unrelated research studies. She is a board member for the charitable European MS

foundation 'The Charcot Foundation' and on the steering committee for a European collaborative MS imaging group 'MAGNIMS' (Magnetic Resonance Imaging in Multiple Sclerosis). Maria Trojano has served on scientific Advisory Boards for Biogen, Novartis, Almirall, Roche and Genzyme; has received speaker honoraria from Biogen Idec, Bayer Schering, Sanofi-Aventis, Merck Serono, Teva, Genzyme, Almirall and Novartis; and has received research grants for her Institution from Biogen Idec, Merck Serono and Novartis.

### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by the Teva Pharmaceuticals Europe.

### References

1. Koch-Henriksen N and Sørensen PS. The changing demographic pattern of multiple sclerosis epidemiology. *Lancet Neurol* 2010; 9: 520–532.
2. Pugliatti M, Rosati G, Carton H, et al. The epidemiology of multiple sclerosis in Europe. *Eur J Neurol* 2006; 13: 700–722.
3. Sormani MP, Stubinski B, Cornelisse P, et al. Magnetic resonance active lesions as individual-level surrogate for relapses in multiple sclerosis. *Mult Scler* 2011; 17: 541–549.
4. Derfuss T. Personalized medicine in multiple sclerosis: Hope or reality? *BMC Med* 2012; 10: 116.
5. Río J, Comabella M and Montalban X. Predicting responders to therapies for multiple sclerosis. *Nat Rev Neurol* 2009; 5: 553–560.
6. Miller DH, Altmann DR and Chard DT. Advances in imaging to support the development of novel therapies for multiple sclerosis. *Clin Pharmacol Ther* 2012; 91: 621–634.
7. The first Pan-European multiple sclerosis multi-stakeholder colloquium: Exploring opportunities and challenges for improving multiple sclerosis management (agenda), <http://2014.ms-colloquium.org/programme> (2014, accessed 3 December 2015).
8. The second Pan-European multiple sclerosis multi-stakeholder colloquium: Accelerating adoption of innovation for better care (agenda), <http://ms-colloquium.org/programme> (2015, accessed 3 December 2015).
9. Confavreux C, Vukusic S and Adeleine P. Early clinical predictors and progression of irreversible disability in multiple sclerosis: An amnesic process. *Brain* 2003; 126: 770–782.

10. Scafari A, Neuhaus A, Degenhardt A, et al. The natural history of multiple sclerosis: A geographically based study. 10. Relapses and long-term disability. *Brain* 2010; 133: 1914–1929.
11. Leray E, Yaouanq J, Le Page E, et al. Evidence for a two-stage disability progression in multiple sclerosis. *Brain* 2010; 133: 1900–1913.
12. Weinshenker BG, Bass B, Rice GPA, et al. The natural history of multiple sclerosis: A geographically based study. I. Clinical course and disability. *Brain* 1989; 112: 133–146.
13. Tremlett H, Yousefi M, Devonshire V, et al. Impact of multiple sclerosis relapses on progression diminishes with time. *Neurology* 2009; 73: 1616–1623.
14. Kremenchutzky M, Cottrell D, Rice G, et al. The natural history of multiple sclerosis: A geographically based study. 7. Progressive-relapsing and relapsing-progressive multiple sclerosis: A re-evaluation. *Brain* 1999; 122: 1941–1949.
15. Lublin FD. Relapses do not matter in relation to long-term disability: No (they do). *Mult Scler* 2011; 17: 1415–1416.
16. Rovaris M, Comi G, Ladkani D, et al. Short-term correlations between clinical and MR imaging findings in relapsing-remitting multiple sclerosis. *AJNR Am J Neuroradiol* 2003; 24: 75–81.
17. Filippi M, Rocca MA, DeStefano N, et al. Magnetic resonance techniques in multiple sclerosis: The present and the future. *Arch Neurol* 2011; 68: 1514–1520.
18. Okuda DT, Siva A, Kantarci O, et al. Radiologically isolated syndrome: 5-year risk for an initial clinical event. *PLoS ONE* 2014; 9: e90509.
19. Polman CH, Reingold SC, Edan G, et al. Diagnostic criteria for multiple sclerosis: 2005 revisions to the ‘McDonald Criteria’. *Ann Neurol* 2005; 58: 840–846.
20. Swanton JK, Rovira A, Tintore M, et al. MRI criteria for multiple sclerosis in patients presenting with clinically isolated syndromes: A multicentre retrospective study. *Lancet Neurol* 2007; 6: 677–686.
21. Brex PA, Ciccarelli O, O’Riordan JI, et al. A longitudinal study of abnormalities on MRI and disability from multiple sclerosis. *N Engl J Med* 2002; 346: 158–164.
22. Tintore M, Rovira À, Río J, et al. Defining high, medium and low impact prognostic factors for developing multiple sclerosis. *Brain*. Epub ahead of print 21 April 2015. DOI: 10.1093/brain/awv105.
23. Barkhof F. MRI in multiple sclerosis: Correlation with expanded disability status scale (EDSS). *Mult Scler* 1999; 5: 283–286.
24. Fisniku LK, Brex PA, Altmann DR, et al. Disability and T2 MRI lesions: A 20-year follow-up of patients with relapse onset of multiple sclerosis. *Brain* 2008; 131: 808–817.
25. Li DKB, Held U, Petkau J, et al. MRI T2 lesion burden in multiple sclerosis: A plateauing relationship with clinical disability. *Neurology* 2006; 66: 1384–1389.
26. Barkhof F, Karas GB and van Walderveen MAA. T1 hypointensities and axonal loss. *Neuroimaging Clin N Am* 2000; 10: 739–752.
27. Van Walderveen MAA, Kamphorst W, Scheltens P, et al. Histopathologic correlate of hypointense lesions on T1-weighted spin-echo MRI in multiple sclerosis. *Neurology* 1998; 50: 1282–1288.
28. Sahraian MA, Radue EW, Haller S, et al. Black holes in multiple sclerosis: Definition, evolution, and clinical correlations. *Acta Neurol Scand* 2010; 122: 1–8.
29. Swanton JK, Fernando KT, Dalton CM, et al. Early MRI in optic neuritis: The risk for disability. *Neurology* 2009; 72: 542–550.
30. Sormani MP, Rovaris M, Comi G, et al. A reassessment of the plateauing relationship between T2 lesion load and disability in MS. *Neurology* 2009; 73: 1538–1542.
31. Sastre-Garriga J, Ingle GT, Chard DT, et al. Grey and white matter volume changes in early primary progressive multiple sclerosis: A longitudinal study. *Brain* 2005; 128: 1454–1460.
32. Vrenken H, Jenkinson M, Horsfield MA, et al. Recommendations to improve imaging and analysis of brain lesion load and atrophy in longitudinal studies of multiple sclerosis. *J Neurol* 2013; 260: 2458–2471.
33. De Stefano N, Airas L, Grigoriadis N, et al. Clinical relevance of brain volume measures in multiple sclerosis. *CNS Drugs* 2014; 28: 147–156.
34. Chard DT, Griffin CM, Parker GJM, et al. Brain atrophy in clinically early relapsing-remitting multiple sclerosis. *Brain* 2002; 125: 327–337.
35. Pérez-Miralles F, Sastre-Garriga J, Tintoré M, et al. Clinical impact of early brain atrophy in clinically isolated syndromes. *Mult Scler* 2013; 19: 1878–1886.
36. Fisher E, Lee JC, Nakamura K, et al. Gray matter atrophy in multiple sclerosis: A longitudinal study. *Ann Neurol* 2008; 64: 255–265.
37. De Stefano N, Giorgio A, Battaglini M, et al. Assessing brain atrophy rates in a large population of untreated multiple sclerosis subtypes. *Neurology* 2010; 74: 1868–1876.
38. Fisher E, Rudick RA, Simon JH, et al. Eight-year follow-up study of brain atrophy in patients with MS. *Neurology* 2002; 59: 1412–1420.

39. Roosendaal SD, Bendfeldt K, Vrenken H, et al. Grey matter volume in a large cohort of MS patients: Relation to MRI parameters and disability. *Mult Scler* 2011; 17: 1098–1106.
40. Lukas C, Minneboo A, de Groot V, et al. Early central atrophy rate predicts 5 year clinical outcome in multiple sclerosis. *J Neurol Neurosurg Psychiatry* 2010; 81: 1351–1356.
41. Popescu V, Agosta F, Hulst HE, et al. Brain atrophy and lesion load predict long term disability in multiple sclerosis. *J Neurol Neurosurg Psychiatry* 2013; 84: 1082–1091.
42. Filippi M and Agosta F. Imaging biomarkers in multiple sclerosis. *J Magn Reson Imaging* 2010; 31: 770–788.
43. De Stefano N, Airas L, Grigoriadis N, et al. Clinical relevance of brain volume measures in multiple sclerosis. *CNS Drugs* 2014; 28: 147–156.
44. Nakamura K, Brown RA, Narayanan S, et al. Diurnal fluctuations in brain volume: Statistical analyses of MRI from large populations. *Neuroimage* 2015; 118: 126–132.
45. Filippi M, Rocca MA, Calabrese M, et al. Intracortical lesions: Relevance for new MRI diagnostic criteria for multiple sclerosis. *Neurology* 2010; 75: 1988–1994.
46. Calabrese M, Rocca MA, Atzori M, et al. Cortical lesions in primary progressive multiple sclerosis: A 2-year longitudinal MR study. *Neurology* 2009; 72: 1330–1336.
47. Calabrese M, Rocca MA, Atzori M, et al. A 3-year magnetic resonance imaging study of cortical lesions in relapse-onset multiple sclerosis. *Ann Neurol* 2010; 67: 376–383.
48. Geurts JGG, Calabrese M, Fisher E, et al. Measurement and clinical effect of grey matter pathology in multiple sclerosis. *Lancet Neurol* 2012; 11: 1082–1092.
49. Geurts JGG, Roosendaal SD, Calabrese M, et al. Consensus recommendations for MS cortical lesion scoring using double inversion recovery MRI. *Neurology* 2011; 76: 418–424.
50. Schmierer K, Scaravilli F, Altmann DR, et al. Magnetization transfer ratio and myelin in postmortem multiple sclerosis brain. *Ann Neurol* 2004; 56: 407–415.
51. Agosta F, Rovaris M, Pagani E, et al. Magnetization transfer MRI metrics predict the accumulation of disability 8 years later in patients with multiple sclerosis. *Brain* 2006; 129: 2620–2627.
52. Khaleeli Z, Altmann DR, Cercignani M, et al. Magnetization transfer ratio in gray matter: A potential surrogate marker for progression in early primary progressive multiple sclerosis. *Arch Neurol* 2008; 65: 1454–1459.
53. Santos AC, Narayanan S, De Stefano N, et al. Magnetization transfer can predict clinical evolution in patients with multiple sclerosis. *J Neurol* 2002; 249: 662–668.
54. Ciccarelli O, Altmann DR, McLean MA, et al. Spinal cord repair in MS: Does mitochondrial metabolism play a role? *Neurology* 2010; 74: 721–727.
55. Balk LJ and Petzold A. Current and future potential of retinal optical coherence tomography in multiple sclerosis with and without optic neuritis. *Neurodegener Dis Manag* 2014; 4: 165–176.
56. Saidha S, Sotirchos ES, Ibrahim MA, et al. Relationships of the inner nuclear layer of the retina with clinicoradiologic disease characteristics in multiple sclerosis; A retrospective study. *Lancet Neurol* 2012; 11: 963–972.
57. Tátrai E, Simó M, Iljicsov A, et al. In vivo evaluation of retinal neurodegeneration in patients with multiple sclerosis. *PLoS ONE* 2012; 7: e30922.
58. Saidha S, Al-Louzi O, Ratchford JN, et al. Optical coherence tomography reflects brain atrophy in multiple sclerosis: A four-year study. *Ann Neurol* 2015; 78: 801–813.
59. Leocani L and Comi G. Clinical neurophysiology of multiple sclerosis. *Handb Clin Neurol* 2014; 122: 671–679.
60. Ramanathan S, Lenton K, Burke T, et al. The utility of multimodal evoked potentials in multiple sclerosis prognostication. *J Clin Neurosci* 2013; 20: 1576–1581.
61. Schlaeger R, D'Souza M, Schindler C, et al. Electrophysiological markers and predictors of the disease course in primary progressive multiple sclerosis. *Mult Scler* 2014; 20: 51–56.
62. Kuhle J, Disanto G, Dobson R, et al. Conversion from clinically isolated syndrome to multiple sclerosis: A large multicentre study. *Mult Scler* 2015; 21: 1013–1024.
63. Dobson R, Ramagopalan S, Davis A, et al. Cerebrospinal fluid oligoclonal bands in multiple sclerosis and clinically isolated syndromes: A meta-analysis of prevalence, prognosis and effect of latitude. *J Neurol Neurosurg Psychiatry* 2013; 84: 909–914.
64. Srivastava R, Aslam M, Kalluri SR, et al. Potassium channel KIR4.1 as an immune target in multiple sclerosis. *N Engl J Med* 2012; 367: 115–123.
65. Brickshawana A, Hinson SR, Romero MF, et al. Investigation of Kir4.1 potassium channel as

- putative antigen of multiple sclerosis: A cohort study. *Lancet Neurol* 2014; 13: 795–806.
66. Comabella M, Fernández M, Martin R, et al. Cerebrospinal fluid chitinase 3-like 1 levels are associated with conversion to multiple sclerosis. *Brain* 2010; 133: 1082–1093.
67. Cantó E, Tintoré M, Villar LM, et al. Chitinase 3-like 1: Prognostic biomarker in clinically isolated syndromes. *Brain* 2015; 138: 918–931.
68. Ascherio A, Munger KL, White R, et al. Vitamin D as an early predictor of multiple sclerosis activity and progression. *JAMA Neurol* 2014; 71: 306–314.
69. Fitzgerald KC, Munger KL, Köchert K, et al. Association of vitamin D levels with multiple sclerosis activity and progression in patients receiving interferon beta-1b. *JAMA Neurol* 72: 1458–1465.
70. Eikelenboom MJ, Petzold A, Lazeron RHC, et al. Multiple sclerosis: Neurofilament light chain antibodies are correlated to cerebral atrophy. *Neurology* 2003; 60: 219–223.
71. Kuhle J, Leppert D, Petzold A, et al. Neurofilament heavy chain in CSF correlates with relapses and disability in multiple sclerosis. *Neurology* 2011; 76: 1206–1213.
72. Comabella M and Montalban X. Body fluid biomarkers in multiple sclerosis. *Lancet Neurol* 2014; 13: 113–126.
73. Disanto G, Adutori R, Dobson R, et al. Serum neurofilament light chain levels are increased in patients with a clinically isolated syndrome. *J Neurol Neurosurg Psychiatry*. Epub ahead of print 25 February 2015. DOI: 10.1136/jnnp-2014-309690.
74. Gold R, Wolinsky JS, Amato MP, et al. Evolving expectations around early management of multiple sclerosis. *Ther Adv Neurol Disord* 2010; 3: 351–367.
75. Edan G and Le Page E. Induction therapy for patients with multiple sclerosis: Why? When? How? *CNS Drugs* 2013; 27: 403–409.
76. O'Connor P, Filippi M, Arnason B, et al. 250µg or 500µg interferon beta-1b versus 20 mg glatiramer acetate in relapsing-remitting multiple sclerosis: A prospective, randomised, multicentre study. *Lancet Neurol* 2009; 8: 889–897.
77. Mikol DD, Barkhof F, Chang P, et al. Comparison of subcutaneous interferon beta-1a with glatiramer acetate in patients with relapsing multiple sclerosis (the REBif vs Glatiramer Acetate in Relapsing MS Disease [REGARD] study): A multicentre, randomised, parallel, open-label trial. *Lancet Neurol* 2008; 7: 903–914.
78. Romeo M, Martinelli-Boneschi F, Rodegher M, et al. Clinical and MRI predictors of response to interferon-beta and glatiramer acetate in relapsing-remitting multiple sclerosis patients. *Eur J Neurol* 2013; 20: 1060–1067.
79. Prosperini L, Gallo V, Petsas N, et al. One-year MRI scan predicts clinical response to interferon beta in multiple sclerosis. *Eur J Neurol* 2009; 16: 1202–1209.
80. Río J, Rovira À, Tintoré M, et al. Relationship between MRI lesion activity and response to IFN-beta in relapsing-remitting multiple sclerosis patients. *Mult Scler* 2008; 14: 479–484.
81. Bermel RA, You X, Foulds P, et al. Predictors of long-term outcome in multiple sclerosis patients treated with interferon beta. *Ann Neurol* 2013; 73: 95–103.
82. Rudick RA, Lee JC, Simon J, et al. Defining interferon beta response status in multiple sclerosis patients. *Ann Neurol* 2004; 56: 548–555.
83. Río J, Rovira A, Tintoré M, et al. Evaluating the response to glatiramer acetate in relapsing-remitting multiple sclerosis (RRMS) patients. *Mult Scler* 2014; 20: 1602–1608.
84. Polman CH, Bertolotto A, Deisenhammer F, et al. Recommendations for clinical use of data on neutralising antibodies to interferon-beta therapy in multiple sclerosis. *Lancet Neurol* 2010; 9: 740–750.
85. Paolicelli D, D'Onghia M, Pellegrini F, et al. The impact of neutralizing antibodies on the risk of disease worsening in interferon beta-treated relapsing multiple sclerosis: A 5 year post-marketing study. *J Neurol* 2013; 260: 1562–1568.
86. Axtell RC, de Jong BA, Boniface K, et al. T helper type 1 and 17 cells determine efficacy of interferon-beta in multiple sclerosis and experimental encephalomyelitis. *Nat Med* 2010; 16: 406–412.
87. Bushnell SE, Zhao Z, Stebbins CC, et al. Serum IL-17F does not predict poor response to IM IFNβ-1a in relapsing-remitting MS. *Neurology* 2012; 79: 531–537.
88. Mahurkar S, Suppiah V and O'Doherty C. Pharmacogenomics of interferon beta and glatiramer acetate response: A review of the literature. *Autoimmun Rev* 2014; 13: 178–186.
89. Macciardi F, Cohen J, Comabella Lopez M, et al. A genetic model predicts the response to glatiramer acetate in two different cohorts. *Neurology* 2012; 78: P05.129.
90. Macciardi F, Cohen J, Comabella Lopez M, et al. A genetic model to predict response to glatiramer acetate developed from a genome wide association study (GWAS). *Neurology* 2012; 78: S20.003.

91. Ritchie MD. The success of pharmacogenomics in moving genetic association studies from bench to bedside: Study design and implementation of precision medicine in the post-GWAS era. *Hum Genet* 2012; 131: 1615–1626.
92. Ross C, Towfic F, Laifenfeld D, et al. A multi-SNP signature predicts high response to Copaxone (Glatiramer Acetate) in RRMS patients. *Mult Scler* 2014; 20(Suppl. 1): 509–510.
93. Fox RJ, Miller DH, Phillips JT, et al. Placebo-controlled phase 3 study of oral BG-12 or glatiramer in multiple sclerosis. *N Engl J Med* 2012; 367: 1087–1097.
94. Vermersch P, Czlonkowska A, Grimaldi LME, et al. Teriflunomide versus subcutaneous interferon beta-1a in patients with relapsing multiple sclerosis: A randomised, controlled phase 3 trial. *Mult Scler* 2014; 20: 705–716.
95. Katsavos S and Anagnostouli M. Biomarkers in multiple sclerosis: An up-to-date overview. *Mult Scler Int* 2013; 2013: 340508.
96. Gunnarsson M, Malmeström C, Axelsson M, et al. Axonal damage in relapsing multiple sclerosis is markedly reduced by natalizumab. *Ann Neurol* 2011; 69: 83–89.
97. Kuhle J, Malmeström C, Axelsson M, et al. Neurofilament light and heavy subunits compared as therapeutic biomarkers in multiple sclerosis. *Acta Neurol Scand* 2013; 128: e33–e36.
98. Kuhle J, Disanto G, Lorscheider J, et al. Fingolimod and CSF neurofilament light chain levels in relapsing-remitting multiple sclerosis. *Neurology* 2015; 84: 1639–1643.
99. Axelsson M, Malmeström C, Gunnarsson M, et al. Immunosuppressive therapy reduces axonal damage in progressive multiple sclerosis. *Mult Scler* 2014; 20: 43–50.
100. Amor S, van der Star BJ, Bosca I, et al. Neurofilament light antibodies in serum reflect response to natalizumab treatment in multiple sclerosis. *Mult Scler* 2014; 20: 1355–1362.
101. Cotte S, von Ahsen N, Kruse N, et al. ABC-transporter gene-polymorphisms are potential pharmacogenetic markers for mitoxantrone response in multiple sclerosis. *Brain* 2009; 132: 2517–2530.
102. Bloomgren G, Richman S, Hotermans C, et al. Risk of natalizumab-associated progressive multifocal leukoencephalopathy. *N Engl J Med* 2012; 366: 1870–1880.
103. Plavina T, Subramanyam M, Bloomgren G, et al. Anti-JC virus antibody levels in serum or plasma further define risk of natalizumab-associated progressive multifocal leukoencephalopathy. *Ann Neurol* 2014; 76: 802–812.
104. Wattjes MP, Rovira À, Miller D, et al. Evidence-based guidelines. MAGNIMS consensus guidelines on the use of MRI in multiple sclerosis-establishing disease prognosis and monitoring patients. *Nat Rev Neurol* 2015; 11: 597–606.
105. Jones JL, Phuah CL, Cox AL, et al. IL-21 drives secondary autoimmunity in patients with multiple sclerosis, following therapeutic lymphocyte depletion with alemtuzumab (Campath-1H). *J Clin Invest* 2009; 119: 2052–2061.
106. Azzopardi L, Thompson SA, Harding KE, et al. Predicting autoimmunity after alemtuzumab treatment of multiple sclerosis. *J Neurol Neurosurg Psychiatry* 2014; 85: 795–798.
107. Hasan SK, Buttari F, Ottone T, et al. Risk of acute promyelocytic leukemia in multiple sclerosis: Coding variants of DNA repair genes. *Neurology* 2011; 76: 1059–1065.
108. Chan A and Lo-Coco F. Mitoxantrone-related acute leukemia in MS: An open or closed book? *Neurology* 2013; 80: 1529–1533.
109. Rovira À, Wattjes MP, Tintoré M, et al. MAGNIMS consensus guidelines on the use of MRI in multiple sclerosis-clinical implementation in the diagnostic process. *Nat Rev Neurol* 2015; 11: 471–480.
110. Shiee N, Bazin PL, Ozturk A, et al. A topology-preserving approach to the segmentation of brain images with multiple sclerosis lesions. *Neuroimage* 2010; 49: 1524–1535.
111. Rothwell PM, McDowell Z, Wong CK, et al. Doctors and patients don't agree: Cross sectional study of patients' and doctors' perceptions and assessments of disability in multiple sclerosis. *BMJ* 1997; 314: 1580–1583.
112. Johnson FR, Van Houtven G, Özdemir S, et al. Multiple sclerosis patients' benefit-risk preferences: Serious adverse event risks versus treatment efficacy. *J Neurol* 2009; 256: 554–562.
113. Wilson LS, Loucks A, Bui CT, et al. Patient preferences for attributes of disease modifying therapies: Results of a choice based conjoint analysis. *Value Health* 2013; 16: A107.
114. Heesen C, Kleiter I, Nguyen F, et al. Risk perception in natalizumab-treated multiple sclerosis patients and their neurologists. *Mult Scler* 2010; 16: 1507–1512.
115. Pietrolongo E, Giordano A, Kleinefeld M, et al. Decision-making in multiple sclerosis consultations in Italy: Third observer and patient assessments. *PLoS ONE* 2013; 8: e60721.
116. Chan A, Pirmohamed M and Comabella M. Pharmacogenomics in neurology: Current state and future steps. *Ann Neurol* 2011; 70: 684–697.