



The Open Biotechnology Journal

Content list available at: www.benthamopen.com/TOBIOTJ/

DOI: 10.2174/1874070701610010173



Phytodecontamination of Water Systems from Phenolic Endocrine Disruptors and the Regulation Role of Natural Organic Matter

Elisabetta Loffredo* and Andreina Traversa

Dipartimento di Scienze del Suolo, della Pianta e Degli Alimenti, Università Degli Studi di Bari Aldo Moro, Via Amendola 165/A, 70126 Bari, Italia

Abstract: In the last decades an increasing number of natural and synthetic compounds have been recognized as endocrine disruptors (EDs) because of their hormone-like activity and capacity to alter the normal hormonal functions of animals and humans. Among EDs, there are phenolic compounds widely present in terrestrial and aquatic systems, such as bisphenol A, 4-nonylphenol (NP), 4-tert-octylphenol, estrone, ethynilestradiol and so on. Estrogenic effects of these molecules have been ascertained on mollusks, crustaceans, fishes, amphibians and mammals starting from concentrations of $1 \mu\text{gL}^{-1}$. Thus, the removal of EDs from polluted media is a priority goal in order to avoid risks for the ecosystem health. Nowadays, several physico-chemical methods are mainly used for the removal of EDs from liquid and solid matrices. Nevertheless, these methods are expensive, difficult to apply and may produce a negative impact on the environment. Recently, most of studies on soil and water remediation from EDs address more sustainable techniques using bacteria, fungi, microbial enzymes and plants. Phytoremediation uses photoautotrophic organisms to uptake, transform, volatilize or stabilize pollutants present in waters, sediments, soils and atmosphere. As this technology is solar driven and exploits natural sources, it is consequently environmentally safe and cost-effective. A fundamental role in the phytoremediation process is played by natural organic molecules, mainly dissolved organic matter and humic substances. These compounds are ubiquitous in all terrestrial and aquatic environments and they interact at various extent with all contaminants deriving from agricultural, industrial and urban activities. Natural organic matter has a relevant biological activity and may also regulate the decontamination capacity of plants and other organisms, such as algae and fungi. In this review, some results of phytodecontamination studies conducted using herbaceous plant species which are presented and discussed. Further, the modulation role of natural organic matter on the phytodecontamination process is highlighted.

Keywords: Endocrine disruptor, humic substances, natural organic matter, organic contaminant, phytoremediation, plant.

PHYTOREMEDIATION

In the last decades, the increasing human activities have contributed markedly to the environmental pollution, particularly regarding natural waters. The huge number of organic and inorganic contaminants present in natural systems impose the need to intervene with specific technologies to restore the original status of these sites accomplishing environmental standards set by laws. Series of national legislative decrees along with European directives regulate the remediation and environmental restoration of contaminated sites, defining the procedures and criteria for the elimination of pollution sources and the reduction of concentrations of pollutants according to the principles and rules of the European Community.

Among the numerous methodologies explored and tested in recent years for remediation, phytoremediation has conquered increasing attractiveness - as a viable alternative towards traditional engineering-type techniques, mainly physico-chemical - owing to its efficiency and sustainability in terms of eco-compatibility and low costs [1]. In fact, both *in situ* and *ex situ* phytoremediation techniques use plant organisms with reduced needs, easy to grow, monitor and adaptable to different edaphic and climatic conditions. Further, plants give a valuable contribution to soil fertility both

* Address correspondence to this author at the Dipartimento di Scienze del Suolo, della Pianta e degli Alimenti, Università degli Studi di Bari Aldo Moro, Via Amendola 165/A, 70126 Bari, Italia; Tel: +39-0805442282; Fax: +39-0805442850; Email: elisabetta.loffredo@uniba.it

directly, through root expansion and exudation, and indirectly promoting the growth and activity of microorganisms. Finally, an added-value of phytoremediation originates from the aesthetic value of plants. All these aspects make this technology very competitive and attractive with a broad consensus by public opinion.

Despite the great potential of phytoremediation, in open field applications the following aspects may limit its success: (i) the confinement of the roots to the surface layer of the contaminated matrix; (ii) the longer time needed in comparison with conventional decontamination techniques; (iii) the susceptibility of plants to abiotic stresses and biotic attacks; (iv) the possible low tolerance of plants towards the pollutant concentration; (v) the possibility to remove only hydrophilic and moderately hydrophobic compounds.

The various phytotechnologies developed are based on the basic physiological processes occurring in higher plants and associated microorganisms, such as transpiration, photosynthesis, metabolism and mineral nutrition [2]. Phytoextraction (also known as phytoaccumulation, phytoabsorption, or phytosequestration) consists in absorbing pollutants from soil through the roots and then translocate them to the stem or leaves. After an adequate period, the removal of aerial plant organs ensure the permanent decontamination of the site. That is particularly important for the removal of metals. Rhizofiltration is another process that plants adopt to uptake metals in aquatic environments. Several aquatic species are very effective in removing heavy metals from water and wastewaters [3]. However, some of these plants, such as pennywort (*Hydrocotyle umbellata* L.) [4] and duckweed (*Lemna minor* L.) [5], have small and slow-growing roots that represent a limit to their use for remediation. Terrestrial plants, such as sunflower (*Helianthus annuus* L.) and Indian mustard (*Brassica juncea* Czern.) [6], are more suitable for rhizofiltration, since they produce longer, thicker, fibrous root systems with large surface areas for the sorption of pollutants. Phytostabilization (or phytoimmobilization) relies on the ability of plants to reduce the mobility or bioavailability of pollutants and thereby avoiding their dispersion by leaching or erosion phenomena and possible entrance into groundwater or food chain. However, this mechanism is only a temporary solution, because the pollutants remain in the contaminated matrix. Plants can immobilize heavy metals in soils by means of sorption by roots, precipitation, complexation or valence reduction in the rhizosphere [7 - 9]. Phytostabilization results more efficient in fine-textured soils with high organic matter content. Phytopumping is a technique used for both organic and inorganic pollutants that uses plants as organic "pumps" to uptake large volumes of contaminated water as part of the transpiration process [10]. The plants that are capable to remove large amounts of water from the soil, such as willow (*Salix* spp.), are most suitable for this technique. Gatliff [11] has shown that *Salix* spp. are capable of taking 200 liters of water per day, thus being able to decontaminate large amounts of groundwater in shallow aquifers. A relevant role for organic pollutants has played by phytodegradation or phytotransformation. This process regards the capacity of some plants to degrade or transform organic contaminants by means of their enzymes. Xenobiotics absorbed by plants can follow different pathways: (i) absorption and relocation in tissues; (ii) complete degradation; and (iii) partial degradation and volatilization. Some plants, named as "Green Liver" for the biosphere, are able to accumulate and detoxify organic xenobiotics collected from contaminated sites through their metabolism. Degradation of phenols was achieved by horseradish, potato and white radish [12, 13], and trichloroethylene in soil and groundwater was efficiently transformed by poplar trees [14, 15]. Another plant process is phytovolatilization and occurs when, under particular conditions, some contaminants absorbed by plants are converted into volatile forms and transferred into the atmosphere through the transpiration process. As a consequence of a partial degradation, some organic xenobiotics, such as trichloroethylene, may become volatile and be transpired by plants through the leaves and stem. However, this phenomenon could generate volatile catabolites of greater toxicity, which would invalidate the decontamination efforts and relocate the contaminant in soil. Rhizodegradation concerns an indirect action of plants which produce root exudates – containing carbohydrates, organic acids, amino acids, flavonoids etc. - that stimulate the degradative metabolism of microorganisms in the rhizosphere. In addition, plants themselves can release enzymes capable of degrading the organic contaminants in the soil. Hedge and Fletcher [16] have shown that phenolic compounds released by red mulberry (*Morus rubra* L.) roots may stimulate the degradation of recalcitrant pollutants by microorganisms. Finally, phytodesalination is an emerging technique that uses halophytes to remediate salt-affected soils restoring the normal plant growth [17, 18].

The choice of the proper phytoremediation technique *in vivo* and *in vitro* applications can guarantee the success of the depollution process. A technique of *in vivo* phytoremediation could be adopted where the contaminant is not in contact with the plant, like in the case of deep aquifers. In such case, the contaminant may be extracted with various technologies and then transferred in suitable areas for the phytoremediation. After treatment, the water or the soil decontaminated can return to their original place, while plants can be harvested and the biomass recycled. As expected, this technology is quite expensive. In a process of *in vitro* phytoremediation, plant extracts, such as enzymes, are used

in decontaminating polluted matrices. Apparently, this technology is the most expensive, because of the high costs of extraction and preparation of enzymes, but some plants, such as *Artemisia dracunculata* var *sativa*, are very efficient in secreting enzymes under stressed conditions, thus lowering the cost of the treatment [10]. Unfortunately, these enzymes are easily degradable in soil conditions that might be a limit of this application. An *in vitro* technique has the advantage of applicability in different environmental conditions and for sites not accessible to plants.

Numerous plant species have shown an appreciable phytoremediative ability towards different classes of organic contaminants. However, a number of these plants cannot be safely used because of their tendency to be invasive or potentially invasive. Some plant species used in phytoremediation from phenolic endocrine disruptors are referred in Table 1.

ENDOCRINE DISRUPTORS

Endocrine disruptor compounds (EDs) are a wide group of persistent organic pollutants known or suspected to interfere either directly or indirectly with the normal functioning of the endocrine system of animals and humans by acting as hormone-like substances [19 - 21]. Exposure to EDs may severely compromise human reproductive functions and success [22], and can lead, especially during the stages of pregnancy and lactation, to serious and long-lasting disturbances of endocrine functions [23]. EDs may enter fresh and sea water due to agricultural practices and by application, discharge and disposal of urban and industrial effluents, sludges, and other wastes. The environmental fate of some endocrine disruptors has been extensively investigated [21, 24 - 26].

Table 1. Some plant species used in phytoremediation from phenolic endocrine disruptors.

Plant Species	Compound	Reference
<i>Chlorella fusca</i>	Bisphenol A	[58]
<i>Cyndodon dactylon</i> , <i>Festuca arundinacea</i> , <i>Lolium perenne</i> , <i>Agropyron fragile</i> , <i>Trifolium repens</i> , <i>Cucumis sativus</i> , <i>Cucurbita pepo</i> , <i>Raphanus sativus</i>	Bisphenol A	[39 - 41]
<i>Dracaena sanderiana</i> , <i>D. fragrans</i>	Bisphenol A	[61]
<i>Eucalyptus perriniana</i> <i>Glycine max</i> , <i>Triticum aestivum</i>	Bisphenol A	[62]
<i>Digitalis purpurea</i> , <i>Datura stramonium</i>	Bisphenol A	[42]
<i>Monoraphidium braunii</i>	Bisphenol A	[56]
<i>Phragmites australis</i>	Bisphenol A	[63]
<i>Portulaca oleracea</i> <i>Pseudokirchneriella subcapitata</i> , <i>Scenedesmus acutus</i> , <i>S. quadricauda</i>	Bisphenol A	[53]
<i>Coelastrum reticulatum</i>	Bisphenol A	[60]
<i>Salvia</i> spp.	Bisphenol A	[64]
<i>Stephanodiscus hantzschii</i>	Bisphenol A	[57]
<i>Vicia faba</i> , <i>Lycopersicon esculentum</i> , <i>Triticum durum</i> , <i>Lactuca sativa</i>		
	Bisphenol A	[43]
<i>Raphanus sativus</i> , <i>Lolium perenne</i>	17 α ethynilestradiol	[40]
<i>Portulaca oleracea</i>	17 β estradiol	[53]
<i>Portulaca oleracea</i>	octylphenol	[53]
<i>Crested wheatgrass</i>	4-Nonylphenol	[44]
<i>Lolium perenne</i> , <i>Brassica rapa</i>	4-Nonylphenol	[45]
<i>Lupinus</i> spp.	4-Nonylphenol	[49]
<i>Portulaca oleracea</i>	4-Nonylphenol	[53]
<i>Raphanus sativus</i> , <i>Lolium perenne</i>	4-Nonylphenol	[39]

Among EDs there are a number of phenolic compounds commonly used in industrial processes and anthropogenic activities, such as bisphenol A, nonylphenol, octylphenol, estrone, estradiol, ethinilestradiol and so on. Bisphenol A (BPA) is used as an intermediate compound in the preparation of epoxy resins and polycarbonates, and as a stabilizer for plastics such as polyvinyl chloride. BPA is present in a wide range of manufactured goods, such as adhesives, food and drink packaging, electrical and electronic parts [24, 27]. 4-nonylphenol (NP) is a component of nonylphenol ethoxylates which are a group of surfactants used as emulsifiers for pesticides, antioxidants for rubbers and plastics,

additives for lubricant oils, and in many other applications [28]. The 17α -ethynylestradiol (EE2) is a synthetic steroid hormone used as component of contraceptive pills. A number of works have been conducted to remove these compounds from natural and anthropogenic water systems using plants. In this review we discuss results of phytoremediation from BPA, NP and EE2 during the germination and growth of a number of herbaceous plant species. Some mechanisms involved in the phytoremediation process are also considered.

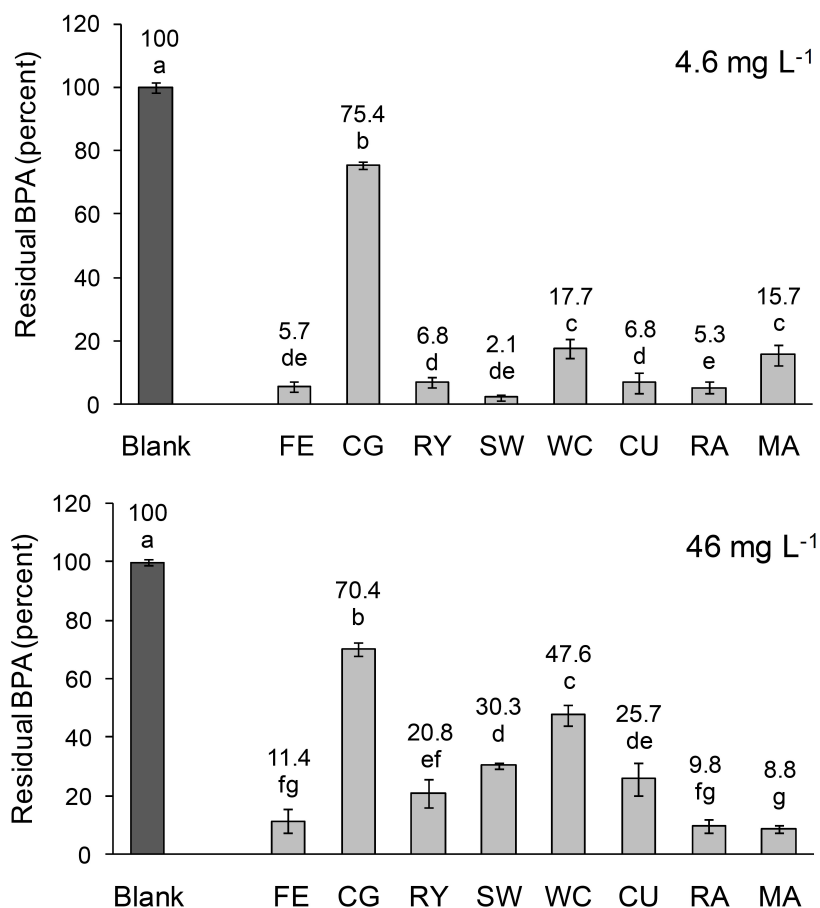


Fig. (1). Residual BPA measured in the blank (without seeds) and in the aqueous medium at the end of germination in the presence of BPA at two concentrations for fescue (FE), couch grass (CG), perennial ryegrass (RY), Siberian wheatgrass (SW), white clover (WC), cucumber (CU), radish (RA), and marrow plant (MA). Values with the same letter are not significantly different according to the least significant differences (LSD) test at probability ≤ 0.05 . The vertical line on each bar indicates the standard error for five replicates. [From Loffredo *et al.* [41] with permission].

THE INFLUENCE OF NATURAL ORGANIC MATTER ON PHYTOREMEDIATION

The distribution and availability of EDs in soil and water systems depend on several factors, including the physical and chemical properties of the compounds and the type and extent of their interaction with the inorganic, organic and biological components of the environmental system. Generally, the affinity of EDs for organic colloids is greater than that for mineral colloids. Thus, the content and nature of suspended and dissolved organic matter, and especially its humified fractions, play a major role in determining the fate of EDs in the natural system.

Approximately 40-60 % of the total organic carbon occurring in natural waters is represented by humic substances (HS) that are ubiquitous natural non-living organic materials. HS consist of relatively high-molecular-mass, yellow-to-black colored organic compounds of mixed aliphatic and aromatic nature, formed by secondary synthesis reactions (humification) of products of microbial and chemical decay and transformation with recalcitrant residues of biomolecules originated from organisms during life and after death [29]. On the basis of their solubility in water at

different pH, HS are divided into two main fractions, humic acids (HAs), which are soluble in dilute alkaline solution, and fulvic acids (FAs), which are soluble at any pH values *e.g.*, [29]. HAs and FAs cannot be regarded as single chemical entities described by unique, chemically defined molecular formulas, but they can be operationally described by model structures constructed on the basis of available chemical and physicochemical data *e.g.*, [29, 30]. A “typical” model macromolecule of soil HA basically consists of aromatic, phenolic, quinonic and heterocyclic “building blocks” that are randomly condensed or linked by aliphatic, oxygen, nitrogen, or sulphur bridges, and bear aliphatic, glucidic, aminoacidic and lipidic surface chains and chemically reactive functional groups (mainly acidic, *e.g.*, carboxylic and phenolic, but also alcoholic hydroxyls, carbonyls, *etc.*) *e.g.*, [30]. FAs generally feature structure and composition less complex than those of HAs, have a lower molecular mass and aromaticity and higher solubility, aliphatic character and content of O-containing functional groups. HA and FA are rich in hydrophilic and hydrophobic sites, exhibit a polydispersed and polyelectrolytic character, possess surface activity, present a relatively open, flexible, sponge-like structure rich of holes [31, 32], and contain a variable amount of highly reactive organic free radical moieties of prevalent semiquinonic nature [30]. All these properties qualify HS, and especially HAs, as privileged natural organic compounds in the interaction with organic contaminants.

The strength and stability of the interaction between organic contaminants and HS influence their persistence, immobilization and accumulation, mobility and transport, bioavailability and biotoxicity, degradability, volatilization and so on. In particular, HS are shown to be able to modify water solubility of organic contaminants, exert catalytic activity on some of them, act as photosensitizers in promoting their photodegradation, and, especially, adsorb them *e.g.*, [33 - 37]. Adsorption of contaminants onto HAs occurs through specific physical and chemical binding mechanisms, including ionic, hydrogen and covalent bonding, charge-transfer or electron donor-acceptor mechanisms, dipole-dipole and Van der Waals forces, ligand exchange, and cation and water bridging [33 - 37]. However, adsorption of nonpolar (hydrophobic) organic contaminants can be better described in terms of non-specific, hydrophobic or partitioning processes between the aqueous phase and the solid organic phase.

The role of HS in the phytoremediation of contaminated natural systems is well accepted but poorly investigated. Ke *et al.* [38] have shown a reduced ability of mangrove *Kandelia candel* (L.) Druce to remove pyrene with the addition of HA to contaminated sediments, as pyrene was more tightly bound to the organic matter. Recently published works demonstrated the fundamental control of natural organic matter on phytoremediation processes [39, 40].

PHYTOREMEDIATION OF AQUEOUS SYSTEMS FROM BISPHENOL A

In a recent study, Loffredo *et al.* [41] assessed the removal of BPA, at the concentrations of 4.6 mg L⁻¹ and 46 mg L⁻¹, from aqueous solutions during the germination and growth of eight herbaceous plants, the forage grasses couch grass *Cynodon dactylon* (L.) Pers., fescue *Schedonorus arundinaceus* (Schreb.), perennial ryegrass (*Lolium perenne* L.), Siberian wheatgrass *Agropyron fragile* (Roth) P. Candargy and white clover (*Trifolium repens* L.) and the horticultural species cucumber (*Cucumis sativus* L.), marrow plant (*Cucurbita pepo* L.) and radish (*Raphanus sativus* L.). Results obtained by these authors are shown in Fig. (1). The plant efficiency in BPA removal followed the order: radish > Siberian wheatgrass > fescue > perennial ryegrass = cucumber (more than 93 % of BPA removed) > marrow plant > white clover > couch grass at the lesser concentration of 4.6 mg L⁻¹, and marrow plant > fescue > radish (more than 84 % of BPA removed) > perennial ryegrass > cucumber > Siberian wheatgrass > white clover > couch grass at the concentration of 46 mg L⁻¹ (Fig. 1). Further, in the range of concentrations tested, BPA removal was proportional to the amount of compound present in the medium. Among the removal mechanisms, uptake by seedlings and degradation by seedling exudates seemed to play the most relevant role. Similarly, in a study with plant cell cultures of soybean *Glycine max* (L.) Merr., wheat (*Triticum aestivum* L.), foxglove (*Digitalis purpurea* L.), and thorn apple (*Datura stramonium* L.), Schmidt and Schuphan [42] attributed the rapid decrease of BPA concentration in the liquid medium to uptake by plant cells.

In other trials, BPA removal was assessed during plant growth in hydroponic culture [41]. In these experiments, the two plants perennial ryegrass and radish were grown for 8 days or 16 days in a nutrient solution added with the compound at concentrations of 4.6 mg L⁻¹ and 46 mg L⁻¹. After 8 days, only a reduced amount of BPA disappeared in the medium, with a maximum of 29% for radish at the lower BPA dose. Conversely, after 16 days perennial ryegrass was able to remove about 97% and 90% of BPA and radish removed 82% and 100%, respectively at the lower and higher concentration (Fig. 2). In these experiments, BPA removal was attributed to both plant uptake and degradation by co-present microorganisms stimulated by released plant exudates [41]. In a previous study conducted in hydroponic conditions, Ferrara *et al.* [43] found significant reductions of the compound in the BPA-contaminated growth medium

of tomato and broad bean after a period of 21 days.

THE EFFECTS OF NATURAL ORGANIC MATTER ON PHYTOREMEDIATION OF ENDOCRINE DISRUPTORS BY HERBACEOUS SPECIES

In a very recent study, Gattullo *et al.* [39] assessed the removal of the endocrine disruptor 4-nonylphenol (NP) at a concentration of 1 mg L^{-1} by ryegrass (*Lolium perenne* L.) and radish (*Raphanus sativus* L.) during their germination. The decontamination process was evaluated in water only or in water containing two organic fractions simulating the organic content of real aqueous systems: a soil humic acid (HA) and a river natural organic matter (NOM) at concentrations of 10 and 200 mg L^{-1} . At the end of germination experiments, NP in water only was not toxic for ryegrass and radish which removed, respectively, 37 and 51 % of the initial NP added. Similar results were obtained by Dettenmaier and Doucette [44] with crested wheatgrass and Domene *et al.* [45] with *Lolium perenne* and *Brassica rapa*. In water added with HA at the two doses, the removal of NP measured by ryegrass and radish was significantly higher than that found in water only, and this positive effect could be attributed to its stimulation of plant enzymatic activity, as previously shown by Nardi *et al.* [46] and Vaughan *et al.* [47]. When water was added with lower dose of NOM, the residual NP measured in the germination media of both ryegrass and radish was similar to that measured in water only. On the contrary, the addition of NOM at the higher dose caused significant and differentiated effects on NP removal by ryegrass (30 % increase) and radish (23 % decrease), with respect to water alone. The different compositional and functional properties of the HA and NOM fractions, in particular the phenolic groups content, might have had an important role in the biological activity. In the case of ryegrass, the stimulation could be ascribed to the same effects reported above for HA, whereas the reduced capacity of radish could depend on the ascertained phytotoxicity of NOM. According to a previous study, the simultaneous presence in the medium of an organic molecule and a humic fraction might cause synergistic negative effect on plant growth [48]. The disappearance of NP in the germination media could be ascribed to seedling activity along with a possible contribution by their associated microorganisms. In fact, Bokern *et al.* [49], in a study on the capability of root cultures of two *Lupinus* species to remove NP, found that most of the initial NP was absorbed by plant and partially transformed in bound residues, while a smaller amount of NP was absorbed and partially transformed or mineralized by microorganisms.

In another study, radish and ryegrass were tested for their capacity to tolerate and remove the combination of bisphenol A (BPA), 17 α -ethynylestradiol (EE2), and linuron at the concentrations of, respectively, 1, 0.1 and 1 mg L^{-1} (first combination) or 10, 1, and 10 mg L^{-1} (second combination) from the following aqueous media: distilled water, a solution of natural organic matter (NOM) at a concentration of 20 mg L^{-1} , a lake water and a river water [40]. Biometric measurements of seedlings at the end of germination experiments revealed that the phytotoxicity of the two combinations of EDs depended on the medium used. In particular, radish showed a discrete tolerance to the EDs in distilled water and lake water but it was inhibited in the NOM solution and river water. Ryegrass appeared negatively affected mainly in river water [40].

In the absence of seedlings, the three compounds did not significantly degrade in the four media with a maximum degradation of 5.8% for EE2 in both combinations. A marked removal of each compound was measured at the end of germination of both plant species except for linuron removal by ryegrass. In the experiments with radish, EDs removal followed the order: EE2 > BPA > linuron. In general, the medium did not significantly influence the removal capacity by this plant, even if the decontamination was less efficient in river water. With both ED combinations, ryegrass removed the three compounds in the order of BPA > EE2 > linuron. The amounts of BPA and EE2 removed by ryegrass were large and dependent on the type of aqueous medium, whereas the removal of linuron was the lowest and similar for the different media. Ryegrass showed generally a better decontamination capacity in lake water and distilled water rather than in NOM solution and river water. The residual amounts of BPA, EE2 and linuron measured in the different media at the end of phytoremediation experiments with radish or ryegrass were significantly lower than the initial amounts, but likely dangerous for aquatic organisms. It is reasonable to expect that an increase of plant biomass used in the phytoremediation could have improved phytoremediation. In this study, when the percentages of ED removed by ryegrass and radish were compared, apparently radish showed a better activity. However, when the comparison was made on the basis of μg of ED removed per g of fresh biomass, in general ryegrass was more efficient than radish in removing BPA and EE2 in both combinations. The monocotyledon ryegrass demonstrated a better decontamination performance than the dicotyledon radish. This is in agreement with other studies demonstrating that, among herbaceous plants, monocotyledonous species possess a higher phytoremediation potential for EDs than dicotyledonous ones [39, 41]. The principal mechanism of removal of BPA, EE2 and linuron was absorption and

transformation by plants, whereas adsorption on root surface and temporary accumulation in plant tissues had a minor role [41, 50 - 52].

The influence of HA or NOM on phytoremediation of EDs has been also assessed in experiments using seedlings grown in hydroponics. Gattullo *et al.* [39] tested the capability of ryegrass and radish seedlings grown for 15 days in nutrient solution to remove NP at a concentration of 1 mg L^{-1} . After 1 and 2 days of growth, their removal capacity was much higher than that observed in germination experiments, indicating that the decontamination efficiency increased with plant age. Imai *et al.* [53] also reported a complete removal of NP by *Portulaca oleracea* L. after 1 day of growth. The addition of 10 mg L^{-1} HA did not affect the removal efficiency of both species, whereas 200 mg L^{-1} HA reduced the removal efficiency of both species after 2 days. The addition of 10 mg L^{-1} NOM improved the ryegrass efficiency after 1 day, whereas 200 mg L^{-1} NOM reduced NP removal by radish after 2 days. Interactions of NP with the two organic fractions at the highest dose, may have resulted in a lower product bioavailability for plants and, therefore, a lesser removal by the same. At the end of experiments, only a little amount of NP was accumulated in plants, and even less in the presence of the two organic fractions, this may be due to a reduced plant uptake. Soares *et al.*, [28] have suggested that the scarce accumulation of NP in plants could be related to its hydrophobic nature. In addition, according to González *et al.* [54] and Matsui *et al.* [55], NP similarly to other phenolic compounds could be bound to polysaccharides and proteins of cell walls on the root surface.

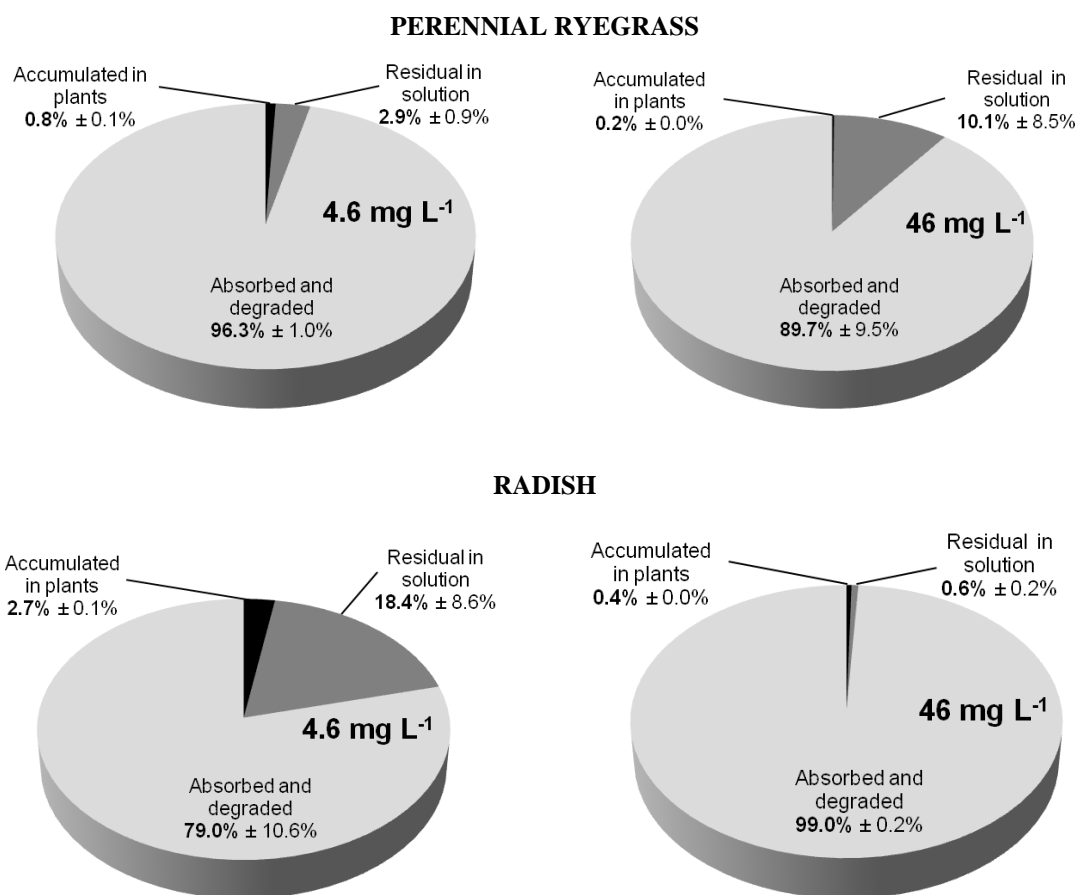


Fig. (2). Distribution of BPA in plants and media after 16 days of growth of perennial ryegrass and radish in the presence of BPA at 4.6 mg L^{-1} and 46 mg L^{-1} . [From Loffredo *et al.* [41] with permission].

In another study, Gattullo *et al.* [56] have tested the capability of the freshwater green alga *Monoraphidium braunii* (Nägeli) to remove BPA at concentrations of 2, 4, and 10 mg L^{-1} , also in presence of NOM at concentrations of 2, 5 and 20 mg L^{-1} . Generally, after 2 and 4 days BPA at lower concentrations was not toxic for the alga, whereas at the highest concentration it reduced algal growth and photosynthetic efficiency. A similar growth response was shown by the marine diatom *Stephanodiscus hantzschii* [57] and the green alga *Chlorella fusca* [58] which showed an evident toxicity

at BPA concentrations equal or higher than 5 mg L⁻¹ and 9 mg L⁻¹, respectively. The sole NOM and its combinations with bisphenol A at the lower concentrations increased the cell number and the chlorophyll a content of algae, whereas all combinations between BPA at highest concentration and NOM resulted very toxic for the algae. After 4-day growth, good removal efficiency was exerted by *M. braunii* at concentrations of 2, 4 and 10 mg L⁻¹ removing, respectively, 39%, 48% and 35% of the initial bisphenol A. NOM at any concentration scarcely influenced the BPA removal. In this study, *M. braunii* was more efficient than numerous different green algal species tested by Hirooka *et al.* [59] and Nakajima *et al.* [60] but less than *Chlorella fusca* [60]. It can be assumed that BPA was taken up by algal cells and successively accumulated and/or metabolized by these.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

This work was funded by the University of Bari, Italy.

REFERENCES

- [1] Jones KC. Organic contaminants in the environments. New York: Elsevier Applied Science 1991.
- [2] Marmiroli N, Marmiroli M, Maestri E. Phytoremediation and phytotechnologies: a review for the present and the future Soil and Water Pollution Monitoring, Protection and Remediation, NATO Science Series. Dordrecht: Springer 2006; Vol. 69: pp. 403-16. [http://dx.doi.org/10.1007/978-1-4020-4728-2_26]
- [3] Tel-Or E, Forni C. Phytoremediation of hazardous toxic metals and organics by photosynthetic aquatic systems. Plant Biosyst 2011; 145: 224-35. [<http://dx.doi.org/10.1080/11263504.2010.509944>]
- [4] Dierberg FE, DeBusk TA, Goulet NA. Aquatic plants for water treatment and resource recovery. In: Reddy KR, Smith WH, Eds. Orlando, FL: Magnolia Publishing 1987; pp. 497-504.
- [5] Mo SC, Choi DS, Robinson JW. Uptake of mercury from aqueous solution by duckweed: the effect of pH, copper, and humic acid. J Environ Health Sci 1989; A24: 135-46.
- [6] Dushenkov V, Kumar PB, Motto H, Raskin I. Rhizofiltration: the use of plants to remove heavy metals from aqueous streams. Environ Sci Technol 1995; 29: 1239-45. [<http://dx.doi.org/10.1021/es00005a015>]
- [7] Barceló J, Poschenrieder C. Phytoremediation: principles and perspectives. Contrib Sci 2003; 2: 333-44.
- [8] Yoon J, Cao X, Zhou Q, Ma LQ. Accumulation of Pb, Cu, and Zn in native plants growing on a contaminated Florida site. Sci Total Environ 2006; 368: 456-64. [<http://dx.doi.org/10.1016/j.scitotenv.2006.01.016>]
- [9] Wuana RA, Okieimen FE. Heavy metals in contaminated soils: a review of sources, chemistry, risks and best available strategies for remediation. ISRN Ecol 2011; 4: 1-20. [<http://dx.doi.org/10.5402/2011/402647>]
- [10] Susarla S, Medina VF, McCutcheon SC. Phytoremediation: an ecological solution to organic chemical contamination. Ecol Eng 2002; 18: 647-58. [[http://dx.doi.org/10.1016/S0925-8574\(02\)00026-5](http://dx.doi.org/10.1016/S0925-8574(02)00026-5)]
- [11] Gatliff EG. Vegetative remediation process offers advantages over traditional pump-and-treat technologies. Remediation 1994; 4: 343-52. [<http://dx.doi.org/10.1002/rem.3440040307>]
- [12] Dec J, Bollag JM. Use of plant material for the decontamination of water polluted with phenols. Biotechnol Bioeng 1994; 44: 1132-9. [<http://dx.doi.org/10.1002/bit.260440915>]
- [13] Roper JC, Dec J, Bollag J. Using minced horseradish roots for the treatment of polluted waters. J Environ Qual 1996; 25: 1242-7. [<http://dx.doi.org/10.2134/jeq1996.00472425002500060011x>]
- [14] Strand SE, Newman L, Ruszaj M, *et al.* Proceedings of the National Conference of the environmental engineering Division of the American Society of Civil Engineers. Pittsburgh, PA, New York 1995.
- [15] Newman LA, Strand SE, Choe N, *et al.* Uptake and biotransformation of trichloroethylene by hybrid poplars. Environ Sci Technol 1997; 31: 1062-7. [<http://dx.doi.org/10.1021/es960564w>]
- [16] Hedge RS, Fletcher JS. Influence of plant growth stage and season on the release of root phenolics by mulberry as related to development of phytoremediation technology. Chemosphere 1996; 32: 2471-9. [[http://dx.doi.org/10.1016/0045-6535\(96\)00144-0](http://dx.doi.org/10.1016/0045-6535(96)00144-0)]

- [17] Zorrig W, Rabhi M, Ferchichi S, Smaoui A, Abdely C. Phytodesalination: a solution for salt-affected soils in arid and semi-arid regions. *J Arid Land Stud* 2012; 22: 299-302.
- [18] Sakai Y, Ma Y, Xu C, Wu H, Zhu W, Yang J. Phytodesalination of a salt-affected soil with four halophytes in China. *J Arid Land Stud* 2012; 22: 17-20.
- [19] Colborn T, vom Saal FS, Soto AM. Developmental effects of endocrine-disrupting chemicals in wildlife and humans. *Environ Health Perspect* 1993; 101: 378-84.
[<http://dx.doi.org/10.1289/ehp.93101378>]
- [20] Kavlock RJ, Daston GP, Derosa C, *et al.* Research needs for the risk assessment of health and environmental effects of endocrine disruptors: a report of the U.S.-EPA-sponsored workshop. *Environ Health Perspect Suppl* 1996; 104: 715-40.
[<http://dx.doi.org/10.1289/ehp.96104s4715>]
- [21] Lintelmann J, Katayama A, Kurihara N, Shore L, Wenzel A. Endocrine disruptors in the environment. *Pure Appl Chem* 2003; 75: 631-81.
[<http://dx.doi.org/10.1351/pac200375050631>]
- [22] Nolan C. Ecosystem Research Reports Series No 29: Endocrine-Disrupters Research in the EU, Report EUR 18345, Environ Clim Res Progr. Brussels, Luxembourg: Office for Official Publication of the EC 1998.
- [23] Colborn T, Clement C. Chemically-induced alterations in sexual and functional development: the wildlife/human connection. New Jersey: Princeton Sci Publ 1992.
- [24] Staples CA, Dorn PB, Klečka GM, O'Block ST, Hariis LR. A review of the environmental fate, effects, and exposures of bisphenol A. *Chemosphere* 1998; 36: 2149-73.
[[http://dx.doi.org/10.1016/S0045-6535\(97\)10133-3](http://dx.doi.org/10.1016/S0045-6535(97)10133-3)]
- [25] Loffredo E, Senesi N. The role of humic substances in the fate of anthropogenic organic pollutants in soil with emphasis on endocrine disruptor compounds. In: Twardowska I, Allen HE, Häggblom MM, Stefaniak S, Eds. *Soil and water pollution monitoring, protection and remediation*, NATO sciences series. Dordrecht: Springer 2006; Vol. 69: pp. 69-92.
- [26] Loffredo E, Senesi N. Fate of organic contaminants in soils with emphasis on adsorption/desorption processes of endocrine disruptor compounds. *Pure Appl Chem* 2006; 78: 947-61. b
[<http://dx.doi.org/10.1351/pac200678050947>]
- [27] Fürhacker M, Scharf S, Weber H. Bisphenol A: emissions from point sources. *Chemosphere* 2000; 41: 751-6.
[[http://dx.doi.org/10.1016/S0045-6535\(99\)00466-X](http://dx.doi.org/10.1016/S0045-6535(99)00466-X)]
- [28] Soares A, Guieyette B, Jefferson B, Cartmell E, Lester JN. Nonylphenol in the environment: a critical review on occurrence, fate, toxicity and treatment in wastewaters. *Environ Int* 2008; 34: 1033-49.
[<http://dx.doi.org/10.1016/j.envint.2008.01.004>]
- [29] Stevenson FJ. Humus chemistry: Genesis, composition, reactions. New York: Wiley 1994; pp. 1-443.
- [30] Senesi N, Loffredo E. The chemistry of soil organic matter. In: Sparks DL, Ed. *Soil physical chemistry*. 2nd ed. Boca Raton, Florida, USA: CRC Press 1999; pp. 239-370.
- [31] Schnitzer M. Humic substances chemistry and reactions soil organic matter. Boca Raton, Florida, USA: Elsevier: Amsterdam 1978; pp. 1-64.
- [32] Hayes MH, Swift RS. The chemistry of soil organic colloids. In: Greenland DJ, Hayes MH, Eds. *The chemistry of soil constituents*. New York: Wiley 1978; pp. 179-320.
- [33] Senesi N, Chen Y. Interactions of toxic organic chemicals with humic substances. In: Gerstl Z, Chen Y, Mingelgrin V, Yaron B, Eds. *Toxic organic chemicals in porous media, ecological studies*. Berlin: Springer-Verlag 1989; Vol. 73: pp. 37-90.
[http://dx.doi.org/10.1007/978-3-642-74468-6_3]
- [34] Senesi N. Binding mechanisms of pesticides to soil humic substances. *Sci Total Environ* 1992; 123/124: 63-76.
[[http://dx.doi.org/10.1016/0048-9697\(92\)90133-D](http://dx.doi.org/10.1016/0048-9697(92)90133-D)]
- [35] Senesi N. Organic pollutant migration in soils as affected by soil organic matter. Molecular and mechanistic aspects. In: Petruzzelli D, Helfferich FG, Eds. *Migration and fate of pollutants in soils and subsoils*, NATO-ASI series. Berlin: Springer-Verlag 1993; Vol. G32: pp. 47-74.
- [36] Senesi N. Nature of interactions between organic chemicals and dissolved humic substances and the influence of environmental factors. In: Beck AJ, James KC, Hyes MH, Eds. *Organic substances in soil and water: natural constituents and their influence on contaminant behaviour*. London: Royal Society of Chemistry 1993; pp. 73-101.
- [37] Senesi N, Miano TM. The role of abiotic interactions with humic substances on the environmental impact of organic pollutants. In: Huang PM, Berthelin J, Bollag JM, McGill WB, Page AL, Eds. *Environmental impact of soil component interactions natural and anthropogenic organics*. Boca Raton: CRC-Lewis 1995; Vol. I: pp. 311-35.
- [38] Ke L, Wong TW, Wong AH, Wong YS, Tam NF. Negative effects of humic acid addition on phytoremediation of pyrene-contaminated sediments by mangrove seedlings. *Chemosphere* 2003; 52: 1581-91.
[[http://dx.doi.org/10.1016/S0045-6535\(03\)00498-3](http://dx.doi.org/10.1016/S0045-6535(03)00498-3)]
- [39] Gattullo CE, Traversa A, Senesi N, Loffredo E. Phytodecontamination of the endocrine disruptor 4-nonylphenol in water also in the presence of two natural organic fractions. *Water Air Soil Pollut* 2012; 223: 6035-44.
[<http://dx.doi.org/10.1007/s11270-012-1337-8>]

- [40] Gattullo CE, Cunha B, Rosa AH, Loffredo E. Removal of a combination of endocrine disruptors from aqueous systems by seedlings of radish and ryegrass. *Environ Technol* 2013; 34: 3129-36. [<http://dx.doi.org/10.1080/09593330.2013.807854>]
- [41] Loffredo E, Gattullo CE, Traversa A, Senesi N. Potential of various herbaceous species to remove the endocrine disruptor bisphenol A from aqueous media. *Chemosphere* 2010; 80: 1274-80. [<http://dx.doi.org/10.1016/j.chemosphere.2010.06.054>]
- [42] Schmidt B, Schuphan I. Metabolism of the environmental estrogen bisphenol A by plant cell suspension cultures. *Chemosphere* 2002; 49: 51-9. [[http://dx.doi.org/10.1016/S0045-6535\(02\)00142-X](http://dx.doi.org/10.1016/S0045-6535(02)00142-X)]
- [43] Ferrara G, Loffredo E, Senesi N. Phytotoxic, clastogenic and bioaccumulation effects of the environmental endocrine disruptor bisphenol A in various crops grown hydroponically. *Planta* 2006; 223: 910-6. [<http://dx.doi.org/10.1007/s00425-005-0147-2>]
- [44] Dettenmaier E, Doucette WJ. Mineralization and plant uptake of ¹⁴C-labeled nonylphenol, nonylphenol tetraethoxylate, and nonylphenol nonylethoxylate in biosolids/soil systems planted with crested wheatgrass. *Environ Toxicol Chem* 2007; 26: 193-200. [<http://dx.doi.org/10.1897/06-268R.1>]
- [45] Domene X, Ramírez W, Solà L, Alcañiz JM, Andrés P. Soil pollution by nonylphenol and nonylphenol ethoxylates and their effects to plants and invertebrates. *J Soils Sediments* 2009; 9: 555-67. [<http://dx.doi.org/10.1007/s11368-009-0117-6>]
- [46] Nardi S, Pizzeghello D, Reniero F, Rascio N. Chemical and biochemical properties of humic substances isolated from forest soils and plant growth. *Soil Sci Soc Am J* 2000; 64: 639-45. [<http://dx.doi.org/10.2136/sssaj2000.642639x>]
- [47] Vaughan D, Malcolm RE, Ord BG. Soil organic matter and biological activity. Dordrecht: Nijhoff-Junk Publishers 1985; pp. 77-108. [http://dx.doi.org/10.1007/978-94-009-5105-1_3]
- [48] Senesi N, Loffredo E, Padovano G. Effects of humic acid-herbicide interactions on the growth of *Pisum sativum* in nutrient solution. *Plant Soil* 1990; 127: 41-7. [<http://dx.doi.org/10.1007/BF00010835>]
- [49] Bokern M, Raid P, Harms H. Toxicity, uptake and metabolism of 4-n-nonylphenol in root cultures and intact plants under septic and aseptic conditions. *Environ Sci Pollut Res* 1998; 5: 21-7. [<http://dx.doi.org/10.1007/BF02986370>]
- [50] Shi W, Lizheng W, Rousseau DP, Lens PN. Removal of estrone, 17 α -ethinylestradiol, and 17 β -estradiol in algae and duckweed-based wastewater treatment systems. *Environ Sci Pollut Res* 2010; 17: 824-33. [<http://dx.doi.org/10.1007/s11356-010-0301-7>]
- [51] Garcinuño RM, Fernández-Hernando P, Cámara C. Removal of carbaryl, linuron, and permethrin by *Lupinus angustifolius* under hydroponic conditions. *J Agric Food Chem* 2006; 54: 5034-9. [<http://dx.doi.org/10.1021/jf060850j>]
- [52] Garcinuño RM, Fernández-Hernando P, Cámara C. Evaluation of pesticide uptake by *Lupinus* seeds. *Water Res* 2003; 37: 3481-9. [[http://dx.doi.org/10.1016/S0043-1354\(03\)00210-0](http://dx.doi.org/10.1016/S0043-1354(03)00210-0)]
- [53] Imai S, Shiraishi A, Gamo K, *et al.* Removal of phenolic endocrine disruptors by *Portulaca oleracea*. *J Biosci Bioeng* 2007; 103: 420-6. [<http://dx.doi.org/10.1263/jbb.103.420>]
- [54] González PS, Agostini E, Milrad SR. Comparison of the removal of 2,4-dichlorophenol and phenol from polluted water, by peroxidases from tomato hairy roots, and protective effect of polyethylene glycol. *Chemosphere* 2008; 70: 982-9. [<http://dx.doi.org/10.1016/j.chemosphere.2007.08.025>]
- [55] Matsui T, Nomura Y, Takano M, *et al.* Molecular cloning and partial characterization of a peroxidase gene expressed in the roots of *Portulaca oleracea* cv., one potentially useful in the remediation of phenolic pollutants. *Biosci Biotechnol Biochem* 2011; 75: 882-90. [<http://dx.doi.org/10.1271/bbb.100823>]
- [56] Gattullo CE, Bährs H, Steinberg EW, Loffredo E. Removal of bisphenol A by the freshwater green alga *Monoraphidium braunii* and the role of natural organic matter. *Sci Total Environ* 2012; 416: 501-6. [<http://dx.doi.org/10.1016/j.scitotenv.2011.11.033>]
- [57] Li R, Chen GZ, Tam NF, *et al.* Toxicity of bisphenol A and its bioaccumulation and removal by a marine microalga *Stephanodiscus hantzschii*. *Ecotoxicol Environ Saf* 2009; 72: 321-8. [<http://dx.doi.org/10.1016/j.ecoenv.2008.05.012>]
- [58] Hirooka T, Nagase H, Uchida K, *et al.* Biodegradation of bisphenol A and disappearance of its estrogenic activity by the green alga *Chlorella fusca* var. *vacuolata*. *Environ Toxicol Chem* 2005; 24: 1896-901. [<http://dx.doi.org/10.1897/04-259R.1>]
- [59] Hirooka T, Akiyama Y, Tsuji N, *et al.* Removal of hazardous phenols by microalgae under photoautotrophic conditions. *J Biosci Bioeng* 2003; 95: 200-3. [[http://dx.doi.org/10.1016/S1389-1723\(03\)80130-5](http://dx.doi.org/10.1016/S1389-1723(03)80130-5)]

- [60] Nakajima N, Teramoto T, Kasai F, *et al.* Glycosylation of bisphenol A by freshwater microalgae. *Chemosphere* 2007; 69: 934-41. [<http://dx.doi.org/10.1016/j.chemosphere.2007.05.088>]
- [61] Saiyood S, Vangnai AS, Thiravetyan P, Inthorn D. Bisphenol A removal by the *Dracaena* plant and the role of plant-associating bacteria. *J Hazard Mater* 2010; 178: 777-85. [<http://dx.doi.org/10.1016/j.jhazmat.2010.02.008>]
- [62] Hamada H, Tomi R, Asada Y, Furuya T. Phytoremediation of bisphenol A by cultured suspension cells of *Eucalyptus perriniana* – regioselective hydroxylation and glycosylation. *Tetrahedron Lett* 2002; 43: 4087-9. [[http://dx.doi.org/10.1016/S0040-4039\(02\)00647-0](http://dx.doi.org/10.1016/S0040-4039(02)00647-0)]
- [63] Ávila C, Reyes C, Bayona JM, García J. Emerging organic contaminant removal depending on primary treatment and operational strategy in horizontal subsurface flow constructed wetlands: influence of redox. *Water Res* 2013; 47: 315-25. [<http://dx.doi.org/10.1016/j.watres.2012.10.005>]
- [64] Okuhata H, Ikeda K, Miyasaka H, *et al.* Floricultural *Salvia* plants have a high ability to eliminate bisphenol A. *J Biosci Bioeng* 2010; 1: 99-101. [<http://dx.doi.org/10.1016/j.jbiosc.2009.12.014>]

Received: June 7, 2014

Revised: May 20, 2015

Accepted: June 5, 2015

© Loffredo and Traversa; Licensee *Bentham Open*.

This is an open access article licensed under the terms of the Creative Commons Attribution-Non-Commercial 4.0 International Public License (CC BY-NC 4.0) (<https://creativecommons.org/licenses/by-nc/4.0/legalcode>), which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.