



Fungal Planet description sheets: 1383–1435

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Key words

ITS nrDNA barcodes
LSU
new taxa
systematics

Abstract Novel species of fungi described in this study include those from various countries as follows: **Australia**, *Agaricus albofoetidus*, *Agaricus aureolephantii* and *Agaricus parviumbrus* on soil, *Fusarium ramsdenii* from stem cankers of *Araucaria cunninghamii*, *Keissleriella sporoboli* from stem of *Sporobolus natalensis*, *Leptosphaerulina queenslandica* and *Pestalotiopsis chiaroscuro* from leaves of *Sporobolus natalensis*, *Serendipita petricola* as endophyte from roots of *Eriochilus petricola*, *Stagonospora tauntonensis* from stem of *Sporobolus natalensis*, *Teratosphaeria carnegiae* from leaves of *Eucalyptus grandis* × *E. camaldulensis* and *Wongia ficherai* from roots of *Eragrostis curvula*. **Canada**, *Lulworthia fundyensis* from intertidal wood and *Newbrunswickomyces abietophilus* (incl. *Newbrunswickomyces* gen. nov.) on buds of *Abies balsamea*. **Czech Republic**, *Geosmithia funiculosata* from a bark beetle gallery on *Ulmus minor* and *Neoherpotrichiella juglandicola* (incl. *Neoherpotrichiella* gen. nov.) from wood of *Juglans regia*. **France**, *Aspergillus rouenensis* and *Neoacrodontium gallica* (incl. *Neoacrodontium* gen. nov.) from bore dust of *Xestobium rufovillosum* feeding on *Quercus* wood, *Endoradicella communis* (incl. *Endoradicella* gen. nov.) endophytic in roots of *Microthlaspi perfoliatum* and *Entoloma simulans* on soil. **India**, *Amanita konajensis* on soil and *Keithomyces indicus* from soil. **Israel**, *Microascus rothbergiorum* from *Stylophora pistillata*. **Italy**, *Calonarius ligusticus* on soil. **Netherlands**, *Appendopyricularia juncicola* (incl. *Appendopyricularia* gen. nov.), *Eriospora juncicola* and *Tetraploa juncicola* on dead culms of *Juncus effusus*, *Gonatophragmium physciae* on *Physcia caesia* and *Paracosmospora physciae* (incl. *Paracosmospora* gen. nov.) on *Physcia tenella*, *Myrmecridium phragmitigenum* on dead culm of *Phragmites australis*, *Neocalalara lolae* on stems of *Pteridium aquilinum*, *Niesslia nieuwvulvenica* on dead culm of undetermined *Poaceae*, *Nothodevriesia narthecii* (incl. *Nothodevriesia* gen. nov.) on dead leaves of *Narthecium ossifragum* and *Parastenospora pini* (incl. *Parastenospora* gen. nov.) on dead twigs of *Pinus sylvestris*. **Norway**, *Verticillium bjoernoeyanum* from sand grains attached to a piece of driftwood on a sandy beach. **Portugal**, *Collybiopsis cimrmanii* on the base of living *Quercus ilex* and amongst dead leaves of *Laurus* and herbs. **South Africa**, *Paraproliferophorum hyphaenes* (incl. *Paraproliferophorum* gen. nov.) on living leaves of *Hyphaene* sp. and *Saccothecium widdringtoniae* on twigs of *Widdringtonia wallichii*. **Spain**, *Corticarius dryosalor* on soil, *Cyphellophora endoradicis* endophytic in roots of *Microthlaspi perfoliatum*, *Geoglossum laurisilvae* on soil, *Leptographium gemmatum* from fluvial sediments, *Physalacria auricularioides* from a dead twig of *Castanea sativa*, *Terfezia bertae* and *Tuber davidlopezii* in soil. **Sweden**, *Alpova larskersii*, *Inocybe alpestris* and *Inocybe boreogodeyi* on soil. **Thailand**, *Russula banwatchanensis*, *Russula purpureoviridis* and *Russula lilacina* on soil. **Ukraine**, *Nectriella adonis* on overwintered stems of *Adonis vernalis*. **USA**, *Microcyclus jacquiniae* from living leaves of *Jacquinia keyensis* and *Penicillium neoherquei* from a minute mushroom sporocarp. Morphological and culture characteristics are supported by DNA barcodes.

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Fungal Planet 1405 – 12 July 2022

Calonarius ligusticus Calledda, Boccardo & Dovana, sp. nov.

Etymology. The epithet '*ligusticus*' reflects the name Liguria, a region in Italy where the holotype was collected.

Classification — *Cortinariaceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata small-sized, phlegmacioid. *Pileus* 25–45 mm diam, hemispherical, then convex to plano-convex, when old depressed, slightly viscid to dry, radially fibrillose, centre with a few whitish to pale brown veil remnants, margin involute for a long time then inflexed; cream, ochraceous to ochraceous brown with some faint brown hues and with pinkish to violaceous tinges. *Lamellae* moderately distant, emarginate to adnate with decurrent tooth, edge even to slightly crenulated, violet at the beginning, darkening to rusty brown with age. *Stipe* 20–40 × 6–12 mm, cylindrical, with a relatively broad marginate bulb up to 20 mm diam long, violet, later becoming brownish, solid, covered with fibrils of the partial veil; bulipellis whitish with a lilac tinge, that becomes brownish when old; whitish to lilac universal veil on the bulb margin; mycelial strands white. *Cortina* fairly sparse, whitish, with age heavily covered with rust-brown spore powder. *Context* whitish to ochraceous in the pileus, with blue tinge in the stipe, brown in the bulb of older specimens. *Odour* and *taste* not distinctive. Macrochemical reaction 30 % KOH on pileus orange-brown, in context pale orange-brown, brown on bulb edge surface. *Basidiospores* (10.5–)10.7–11.7–12.6(–14.5) × (5.0–)5.9–6.4–6.9(–7.5) µm Q = (1.49–)1.61–1.83–2.05(–2.36) citriform, amygdaliform to subamygdaliform, strongly and coarsely, net-like verrucose, suprahilar plage indistinct, apiculus smooth. *Basidia* 27–32 × 8–10 µm, clavate, four-spored, thin-walled and hyaline in KOH, sterigmata up to 3.5 µm long. *Lamella edge* fertile, presence of cystidioid cylindrical elements. *Cheilocystidia* and *pleurocystidia* not observed. *Pileipellis* as an ixocutis, hyphae hyaline, yellow to brown, cylindrical to slightly moniliform with subcapitate terminal elements 4–6 µm wide. *Pigments* cytoplasmic and parietal. *Clamp connections* frequent at all septa.

Habitat & Distribution — In deciduous forest with *Quercus ilex* on calcareous soil. Found, so far as we know, only in North Italy in Liguria Region.

Typus. ITALY, Zoagli, Località Le Grazie, in a dense forest of *Quercus ilex*, near the path, on calcareous soil, 30 Nov. 2019, F. Calledda, M. Carbone & E. Pini (holotype GDOR5237, ITS and LSU sequences GenBank OM980183 and OM980184, MycoBank MB 843353).

Notes — Spore dimensions are expressed as (a)b–c–d(e), where (a) = minimum value, b = (average – standard deviation), c = average, d = (average + standard deviation) and (e) = maximum value. *Calonarius ligusticus* is characterised by a small basidioma, cream to ochraceous brown fibrillose pileus with pinkish to light violaceous tinges and with some brown hues, lamellae moderately distant and distinctly violaceous, violaceous stipe with broad marginate bulb covered by whitish to lilac universal veil, mycelial strands white and orange alkaline reaction on the pileus surface and in the context. Microscopically, *Calonarius ligusticus* shows high variability in spore shape (citriform, amygdaliform to subamygdaliform, Q = 1.49–2.36) and size (10.5–14.5 × 5.0–7.5 µm) and a pileipellis as an ixocutis, with brown subcapitate terminal elements. In the nrITS phylogenetic analysis, *Calonarius ligusticus* is in a well-supported /*Calochroa* clade (maximum-likelihood bootstrap, MLB = 98 %), and it is a sister species of *Calonarius laberiae* (MLB = 100 %), with which it shares 97 % bp. Based on a megablast search of NCBIs GenBank nucleotide database, the best hit using the LSU sequence is *Calonarius sodagnitus* (voucher AFTOL-ID 811, GenBank; Identities = 909/915 (99 %), no gaps). Morphologically, *C. laberiae* differs mainly from *C. ligusticus* by yellow to yellow-ochraceous pileus, greyish white lamellae, white to ochraceous stipe, smaller basidiospores (9.3–12.2 × 5.9–7.3 µm) and different habitat under *Abies* and *Picea* (Münzmay et al. 2009). *Calonarius sodagnitus* is easily distinguished from *C. ligusticus* by its violaceous pileus with dark spots, intense alkaline reactions on bulipellis and pileus and smaller basidiospores (8.5–10 × 5–6 µm) (Brandrud et al. 1992).

Colour illustrations. Zoagli, Italy, *Quercus ilex* forest. *Calonarius ligusticus* basidiomata in habitat; basidiospores and pileipellis. Scale bars = 10 µm.

Supplementary material

FP1405 Phylogenetic tree.

REFERENCES

- Abdelrazeq, S, Choudhari S, Thimmapuram J, et al. 2020. Changes in the core endophytic mycobiome of carrot taproots in response to crop management and genotype. *Scientific Reports* 10: 13685.
- Agerer R. 1980. Contribution to neotropical cyphellaceous fungi. 11. *Deigloria* gen. nov. (*Physalacriaceae*). *Mycotaxon* 12: 185–200.
- Al Subeh ZY, Raja HA, Burdette JE, et al. 2021. Three diketomorpholines from a *Penicillium* sp. (strain G1071). *Phytochemistry* 189: 112830.
- Alfieri Jr SA, Samuels GJ. 1979. *Nectriella pironii* and its *kutilakesa*-like anamorph, a parasite of ornamental shrubs. *Mycologia* 71: 1178–1185.
- Andjic V, Pegg GS, Carnegie AJ, et al. 2010. Teratosphaeria pseudoeucalypti, new cryptic species responsible for leaf blight of Eucalyptus in subtropical and tropical Australia. *Plant Pathology* 59: 900–912.
- Antonín V, Noordeloos M. 2010. A monograph of marasmoid and collybioid fungi in Europe. IHW Verlag.
- Arauzo S, Iglesias P. 2014. La familia Geoglossaceae ss. str. en la península Ibérica y la Macaronesia. *Errötari* 11: 166–259.
- Arya CP, Manoj Kumar A, Pradeep CK, et al. 2022. *Agaricus brunneodiscus*, a new species of *Agaricus* sect. *Rarolentes* from India. *Phytotaxa* 533: 181–193.
- Arzanalou M, Groenewald JZ, Gams W, et al. 2007. Phylogenetic and morphotaxonomic revision of *Ramichloridium* and allied genera. *Studies in Mycology* 58: 57–93.
- Aylward J, Havenga M, Dreyer LL, et al. 2021. Genetic diversity of *Teratosphaeria pseudoeucalypti* in Eucalyptus plantations in Australia and Uruguay. *Australasian Plant Pathology* 50: 639–649.
- Azevedo E, Barata M, Marques MI, et al. 2017. *Lulworthia atlantica*: a new species supported by molecular phylogeny and morphological analysis. *Mycologia* 109: 287–295.
- Azuddin NF, Mohd MH, Rosely NFN, et al. 2021. Molecular phylogeny of endophytic fungi from Rattan (*Calamus castaneus* Griff.) spines and their antagonistic activities against plant pathogenic fungi. *Journal of Fungi* 7: 301.
- Bandini D, Oertel B, Schüssler C, et al. 2020. Noch mehr Risspilze: Fünfzehn neue und zwei wenig bekannte Arten der Gattung *Inocybe*. *Mycologia* 20: 13–101.
- Bao D-F, Hyde KD, McKenzie EHC, et al. 2021. Biodiversity of lignicolous freshwater hyphomycetes from China and Thailand and description of sixteen species. *Journal of Fungi* 7: 669.
- Barr ME. 1972. Preliminary studies on the Dothideales in temperate North America. Contributions from the University of Michigan Herbarium 9: 523–638.
- Bashir H, Chen J, Jabeen S, et al. 2021. An overview of *Agaricus* section *Hondenses* and *Agaricus* section *Xanthodermatei* with description of eight new species from Pakistan. *Scientific Reports* 11: 12905.
- Bell J, Yokoya K, Kendon JP, et al. 2020. Diversity of root-associated culturable fungi of *Cephalanthera rubra* (Orchidaceae) in relation to soil characteristics. *PeerJ* 8: e8695.
- Bellanger J-M, Moreau P-A, Corriol G, et al. 2015. Plunging hands into the mushroom jar: a phylogenetic framework for Lyophyllaceae (Agaricales, Basidiomycota). *Genetica* 143: 169–194.
- Berger F, Braun U, Heuchert B. 2015. *Gonatophragmium lichenophilum* sp. nov. – a new lichenicolous hyphomycete from Austria. *Mycobiotika* 5: 7–13.
- Berthier J. 1985. Les *Physalacriaceae* du Globe. *Biblioteca Mycologica* 98: 1–128.
- Bidaud A. 2011. Cortinaires rares ou nouveaux de la région Rhône-Alpes (France). *Journal des J.E.C.* 13: 4–24.
- Bonito G, Hameed K, Ventura R, et al. 2016. Isolating a functionally relevant guild of fungi from the root microbiome of *Populus*. *Fungal Ecology* 22: 35–42.
- Brandrud TE, Lindström H, Marklund H, et al. 1992. *Cortinarius*: Flora Photographic. Vol. II (Swedish version). *Cortinarius* HB, Sweden.
- Braun U, Crous PW, Nakashima C. 2015. Cercosporoid fungi (Mycosphaerellaceae) 4. Species on dicots (Acanthaceae to Amaranthaceae). *IMA Fungus* 6: 373–469.
- Braun U, Nakashima C, Crous PW. 2013. Cercosporoid fungi (Mycosphaerellaceae) 1. Species on other fungi, Pteridophyta and Gymnospermae. *IMA Fungus* 4: 265–345.
- Burgess TI, Wingfield MJ. 2017. Pathogens on the move: A 100-year global experiment with planted eucalypts. *Bioscience* 67: 14–25.
- Cannon P, Carmarán C, Romero A. 1995. Studies on biotrophic fungi from Argentina: *Microcyclus porlieriae*, with a key to South American species of *Microcyclus*. *Mycological Research* 99: 353–356.
- Ceruti A, Fontana A, Nosenzo C. 2003. Le specie europee del genere *Tuber*: una revisione storica. Vol. 37. Turin, Italy, Museo Regionale di Scienze Naturali.
- Chang R, Cao W, Wang Y, et al. 2022. *Melanodevriesia*, a new genus of endolichenic oleaginous black yeast recovered from the Inner Mongolia Region of China. *Fungal Systematics and Evolution* 9: 1–9.
- Chen J, Callac P, Parra LA, et al. 2017. Study in *Agaricus* subgenus *Minores* and allied clades reveals a new American subgenus and contrasting phylogenetic patterns in Europe and Greater Mekong Subregion. *Persoonia* 38: 170–196.
- Chen J, Parra L, Kesel A, et al. 2016. Inter- and intra-specific diversity in *Agaricus endoxanthus* and allied species reveals a new taxon, *A. punjabensis*. *Phytotaxa* 252: 1–16.
- Cochard H, Réaudin D. 2019. *Physalacria stilboidea* (Cooke) Sacc., espèce exotique nouvelle pour la France. *Bulletin Mycologique et Botanique Dauphiné-Savoie* 235: 11–16.
- Cooke MC. 1889. New Australian fungi. *Grevillea* 18: 1–8.
- Corner EJH. 1950. A monograph of *Clavaria* and allied genera. *Annals of Botany Memoirs* 1: 1–740.
- Cripps CL, Larsson E, Vauras J. 2020. Nodulose-spored *Inocybe* species from the Rocky Mountain alpine zone: molecularly linked to European type specimens. *Mycologia* 112: 133–153.
- Crous PW, Cowan DA, Maggs-Colling G, et al. 2020a. Fungal Planet description sheets: 1112–1181. *Persoonia* 45: 251–409.
- Crous PW, Cowan DA, Maggs-Colling G, et al. 2021a. Fungal Planet description sheets: 1182–1283. *Persoonia* 46: 313–528.
- Crous PW, Groenewald JZ. 2011. Why everlastings don't last. *Persoonia* 26: 70–84.
- Crous PW, Hernández-Restrepo M, Schumacher RK, et al. 2021b. New and interesting fungi 4. *Fungal Systematics and Evolution* 7: 255–343.
- Crous PW, Osieck ER, Jurjević Ž, et al. 2021c. Fungal Planet description sheets: 1284–1382. *Persoonia* 47: 178–374.
- Crous PW, Schumacher RK, Akulov A, et al. 2019. New and interesting fungi. 2. *Fungal Systematics and Evolution* 3: 57–134.
- Crous PW, Shivas RG, Quaedvlieg W, et al. 2014. Fungal Planet description sheets: 214–280. *Persoonia* 32: 184–306.
- Crous PW, Summerell BA, Shivas RG, et al. 2011. Fungal Planet description sheets: 92–106. *Persoonia* 27: 130–162.
- Crous PW, Wingfield MJ, Burgess TI, et al. 2018. Fungal Planet description sheets: 716–784. *Persoonia* 40: 239–392.
- Crous PW, Wingfield MJ, Chooi Y-H, et al. 2020b. Fungal Planet description sheets: 1042–1111. *Persoonia* 44: 301–459.
- Crous PW, Wingfield MJ, Richardson DM, et al. 2016. Fungal Planet description sheets: 400–468. *Persoonia* 36: 316–458.
- Crous PW, Wingfield MJ, Schumacher RK, et al. 2020c. New and interesting fungi 3. *Fungal Systematics and Evolution* 6: 157–231.
- Da Hora Júnior BT, De Macedo DM, Barreto RW, et al. 2014. Erasing the past: a new identity for the Damoclean pathogen causing South American leaf blight of rubber. *PLoS ONE* 9: e104750.
- De Beer ZW, Wingfield MJ. 2013. Emerging lineages in the Ophiostomatidae. In: Seifert KA, De Beer ZW, Wingfield MJ (eds), *The Ophiostomatoid fungi: expanding frontiers*: 21–46. CBS-KNAW Fungal Biodiversity Centre, Utrecht, The Netherlands.
- De Hoog GS. 1972. The genera *Beauveria*, *Isaria*, *Tritirachium* and *Acrodontium* gen. nov. *Studies in Mycology* 1: 1–41.
- Dereeper A, Guignon V, Blanc G, et al. 2008. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. *Nucleic Acids Research* 1: 36 (Web Server issue): W465–W469.
- Desjardin DE. 1987. New and noteworthy marasmoid fungi from California. *Mycologia* 79: 123–134.
- Desjardin DE, Perry BA. 2017. The gymnopoid fungi (Basidiomycota, Agaricales) from the Republic of São Tomé and Príncipe, West Africa. *Mycosphere* 8: 1317–1391.
- Devadatha B, Calabon MS, Abeywickrama PD, et al. 2020. Molecular data reveals a new holomorphic marine fungus, *Halobyssothecium estuariae*, and the asexual morph of *Keissleriella phragmiticola*. *Mycology* 11:167–183.
- Dutta AK, Wilson AW, Antonín V, et al. 2015. Taxonomic and phylogenetic study on gymnopoid fungi from Eastern India I. *Mycological Progress* 14: 79.
- Eberhart J, Trappe J, Piña Páez C, et al. 2020. *Tuber luomae*, a new spiny-spored truffle species from the Pacific Northwest, USA. *Fungal Systematics and Evolution* 6: 299–304.
- Ellis MB. 1949. *Tetraploa*. *Transactions British Mycological Society* 32: 246–251.
- Esteve-Raventós F, Bandini D, Oertel B, et al. 2018. Advances in the knowledge of the *Inocybe mixtilis* group (Inocybaceae, Agaricomycetes), through molecular and morphological studies. *Persoonia* 41: 213–236.
- Fritsche Y, Lopes ME, Selosse M-A, et al. 2021. *Serendipita restingae* sp. nov. (Sebacinales); an orchid mycorrhizal agaricomycete with wide host range. *Mycorrhiza* 30: 1–15.

- Gams W, Philippi S. 1992. A study of *Cyathicula strobilina* and its Chalara anamorph in vitro. *Persoonia* 14: 547–552.
- Gams W, Stielow B, Gräfenhan T, et al. 2019. The ascomycete genus Niesslia and associated monocillium-like anamorphs. *Mycological Progress* 18: 5–76.
- Garnica S, Schön ME, Abarenkov K, et al. 2016. Determining threshold values for barcoding fungi: lessons from *Cortinarius* (Basidiomycota), a highly diverse and widespread ectomycorrhizal genus. *FEMS Microbiology Ecology* 92: fiw045.
- Geiser DM, Ivery MLL, Hakiza G, et al. 2005. Gibberella xylosoidea (anamorph: *Fusarium xylosoidea*), a causative agent of coffee wilt disease in Africa, is a previously unrecognized member of the *G. fujikuroi* species complex. *Mycologia* 97: 191–201.
- Gerlach W. 1977. *Fusarium robustum* spec. nov., der Erreger einer Stammfäule an Araucaria angustifolia (Bertol.) O. Kuntze in Argentinien? *Journal of Phytopathology* 88: 29–37.
- Gerlach W, Nirenberg H. 1982. The genus *Fusarium* – a pictorial atlas. Mitteilungen aus der Biologischen Bundesanstalt für Land- und Forstwirtschaft Berlin-Dahlem 209: 1–406.
- Glynou K, Ali T, Buch A-K, et al. 2016. The local environment determines the assembly of root endophytic fungi at a continental scale. *Environmental Microbiology* 18: 2418–2434.
- Greif MD, Gibas CFC, Currah RS. 2006. *Leptographium piriforme* sp. nov., from a taxonomically diverse collection of arthropods collected in an aspen-dominated forest in western Canada. *Mycologia* 98: 771–780.
- Gui Y, Zhu GS, Callac P, et al. 2015. Agaricus section Arvenses: three new species in highland subtropical Southwest China. *Fungal Biology* 119: 79–94.
- Guindon S, Dufayard JF, Lefort V, et al. 2010. New algorithms and methods to estimate maximum likelihood phylogenies: Assessing the performance of PhyML 3.0. *Systematic Biology* 59: 307–321.
- Havenga M, Wingfield BD, Wingfield MJ, et al. 2020. Diagnostic markers for *Teratosphaeria destructans* and closely related species. *Forest Pathology* 50: e12645.
- Hayward J, Tourtellot SG, Horton TR. 2014. A revision of the Alpova diaphlophloeus complex in North America. *Mycologia* 106: 846–855.
- Hernández-Restrepo M, Giraldo A, Van Doorn R, et al. 2020. The genera of fungi - G6: Arthrophraphis, Kramasamuha, Melnikomyces, Thysanorea, and Verruconis. *Fungal Systematics and Evolution* 6: 1–24.
- Herrera CS, Rossman AY, Samuels GJ, et al. 2013. Pseudocosmospora, a new genus to accommodate *Cosmospora vilior* and related species. *Mycologia* 105: 1287–1305.
- Hoang DT, Chernomor O, Von Haeseler A, et al. 2018. UFBoot2: Improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution* 35: 518–522.
- Horak E, Desjardin DE. 1994. Reduced marasmoid and mycenoid Agarics from Australasia. *Australian Systematic Botany* 7: 153–170.
- Hou LW, Groenewald JZ, Pfenning LH, et al. 2020. The phoma-like dilemma. *Studies in Mycology* 96: 309–396.
- Houben J, Kocsé S, Visagie CM, et al. 2020. Classification of Aspergillos, Penicillium, Talaromyces and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. *Studies in Mycology* 95: 5–169.
- Hudson O, Buchholz M, Doyle V, et al. 2019. Multilocus phylogeny of Acrospermaceae: New epibiotic species and placement of Gonatophragmium, Pseudovirgaria, and Phaeodactylum anamorphs. *Mycologia* 111: 1041–1055.
- Huelsenbeck JP, Ronquist F. 2001. MrBayes: Bayesian inference of phylogenetic trees. *Bioinformatics* 17: 754–755.
- Hyde KD, Hongsan S, Jeewon R, et al. 2016. Fungal diversity notes 367–490: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 80: 1–270.
- Inderbitzin P, Bostock RM, Davis RM, et al. 2011. Phylogenetics and taxonomy of the fungal vascular wilt pathogen *Verticillium*, with the descriptions of five new species. *PLoS ONE* 6: e28341.
- Jacob M, Bhat DJ. 2000. Two new endophytic fungi from India. *Cryptogamie, Mycologie* 21: 81–88.
- Jacobsson S, Larsson E. 2018. Inocybe (Fr.) Fr. In: Knudsen H, Vesterholt J. (eds), *Funga Nordica*. Agaricoid, boletoid, cyphelloid and gasteroid genera. Second Edition: 981–1021. Nordsvamp, Copenhagen.
- Jankowiak R, Kolařík M. 2010. *Leptographium piriforme* – first record for Europe and of potential pathogenicity. *Biologia* 65: 754–757.
- Jayawardena RS, Hyde KD, McKenzie EH, et al. 2019. One stop shop III: taxonomic update with molecular phylogeny for important phytopathogenic genera: 51–75 (2019). *Fungal Diversity* 98: 77–160.
- Jones EBG, Pang K-L, Abdel-Wahab MA, et al. 2019. An online resource for marine fungi. *Fungal Diversity* 96: 347–433.
- Jones EBG, Suetrong S, Sakayaroj J, et al. 2015. Classification of marine Ascomycota, Basidiomycota, Blastocladiomycota and Chytridiomycota. *Fungal Diversity* 73: 1–72.
- Kalyanaamoorthy S, Minh BQ, Wong TKF, et al. 2017. ModelFinder: Fast model selection for accurate phylogenetic estimates. *Nature Methods* 14: 587–589.
- Kamara AM, El-Lakany MH, Badran OA, et al. 1981. Seed pathology of Araucaria spp. 1. A survey of seed-borne fungi associated with four Araucaria spp. *Australian Forest Research* 11: 269–274.
- Karunaratna A, Dzialak P, Jayawardena RS, et al. 2021. A novel addition to the Pezizellaceae (Rhytismatales, Ascomycota). *Phytotaxa* 480: 251–261.
- Katoh K, Standley DM. 2013. MAFFT Multiple Sequence Alignment Software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772–780.
- Kearse M, Moir R, Wilson A, et al. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28: 1647–1649.
- Kerrigan RW. 2016. Agaricus of North America. *Memoirs of The New York Botanical Garden Volume 114*. NYBG Press.
- Kers LE. 1981. Några anmärkningsvärdा fynd av hypogea svampar i Sverige. *Svensk Botanisk Tidskrift* 75: 129–140.
- Kers LE. 1983. Några svenska fynd av hypogea svampar. *Svensk Botanisk Tidskrift* 77: 259–268.
- Kers LE. 1986. Några norska fynd av hypogea. *Agarica* 7: 30–48.
- Khemmuk W, Geering ADW, Shivas RG. 2016. Wongia gen. nov. (Papuloseae, Sordariomycetes), a new generic name for two root-infecting fungi from Australia. *IMA Fungus* 7: 247–252.
- Klaubauf S, Tharreau D, Fournier E, et al. 2014. Resolving the polyphyletic nature of Pyricularia (Pyriculariaceae). *Studies in Mycology* 79: 85–120.
- Knapp DG, Imrefi I, Boldpurev E, et al. 2019. Root-colonizing endophytic fungi of the dominant grass *Stipa krylovii* from a Mongolian Steppe grassland. *Frontiers in Microbiology* 10: 2565.
- Kolařík M, Hulcr J, Tisserat N, et al. 2017. Geosmithia associated with bark beetles and woodborers in the western USA: taxonomic diversity and vector specificity. *Mycologia* 109: 185–199.
- Kolařík M, Jankowiak R. 2013. Vector affinity and diversity of Geosmithia fungi living on subcortical insects inhabiting Pinaceae species in Central and Northeastern Europe. *Microbial Ecology* 66: 682–700.
- Kolařík M, Kostovčík M, Pažoutová S. 2007. Host range and diversity of the genus Geosmithia (Ascomycota: Hypocreales) living in association with bark beetles in the Mediterranean area. *Mycological Research* 111: 1298–1310.
- Kolařík M, Kubáčová A, Pažoutová S. 2004. Morphological and molecular characterisation of Geosmithia putterillii, G. pallida comb. nov. and G. flava sp. nov., associated with subcorticolous insects. *Mycological Research* 108: 1053–1069.
- Kornerup A, Wanscher JH. 1978. Methuen handbook of colour. 3rd ed. Eyre Methuen, London.
- Kornerup A, Wanscher JH. 1981. Taschenlexikon der Farben. Muster-Schmidt Verlag, Göttingen.
- Kovács G, Calonge FD, Martín MP. 2011. The diversity of Terfezia desert truffles: new species and a highly variable species complex with intra-sporocarpic nrDNA ITS heterogeneity. *Mycologia* 103: 841–853.
- Kozlov AM, Darriba D, Flouri T, et al. 2019. RAxML-NG: a fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. *Bioinformatics* 35: 4453–4455.
- Kumar S, Stecher G, Li M, et al. 2018. MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution* 35: 1547–1549.
- Kumar S, Stecher G, Tamura K. 2016. MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution* 33: 1870–1874.
- Kuyper TW. 1986. A revision of the genus Inocybe in Europe. I. Subgenus Inosperma and the smooth-spored species of subgenus Inocybe. *Persoonia Supplement* 3(1): 1–247.
- Laessøe T, Spooner BM. 1993. New British records. 103. Physalacria cryptomeriae Berthier & Rogerson. 104. Physalacria stilboidea (Cooke) Sacc. *Mycologist* 7: 162–163.
- Letunic I, Bork P. 2019. Interactive Tree Of Life (iTOL) v4: recent updates and new developments. *Nucleic Acids Research* 47: W256–W259.
- Li T, Deng WQ, Li TH, et al. 2018. Endophytic fungal communities associated with leaves, stems and roots of four medicinal plants in South China. *Studies in Fungi* 3: 126–140.
- Li X, Li W, Chu L, et al. 2016. Diversity and heavy metal tolerance of endophytic fungi from *Dysphania ambrosioides*, a hyperaccumulator from Pb–Zn contaminated soils. *Journal of Plant Interactions* 11: 186–192.

- Liu H, Li T, Ding Y, et al. 2017. Dark septate endophytes colonizing the roots of 'non-mycorrhizal' plants in a mine tailing pond and in a relatively undisturbed environment, Southwest China. *Journal of Plant Interactions* 12: 264–271.
- Liu JK, Hyde KD, Gareth EBG, et al. 2015. Fungal diversity notes 1–110: taxonomic and phylogenetic contributions to fungal species. *Fungal Diversity* 72: 1–197.
- Loizides M, Kyriakou T, Tziakouris A. 2011. Edible & toxic Fungi of Cyprus. Published by the authors.
- Madrid H, Hernández M, Gené J, et al. 2016. New and interesting chaetothyrialean fungi from Spain. *Mycological Progress* 15: 1179–1201.
- Maharachchikumbura SSN, Guo LD, Chukeatirote E, et al. 2011. Pestalotiopsis-morphology, phylogeny, biochemistry and diversity. *Fungal Diversity* 50: 167–187.
- Maharachchikumbura SSN, Hyde KD, Groenewald JZ, et al. 2014. Pestalotiopsis revisited. *Studies in Mycology* 79: 121–186.
- McAlpine D. 1902. Fungus diseases of stone-fruit trees in Australia and their treatment. Melbourne, Victorian Department of Agriculture.
- Mello A, Vizzini A, Longato S, et al. 2000. *Tuber borchii* versus *Tuber maculatum*: neotype studies and DNA analyses. *Mycologia* 92: 326–331.
- Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: Proceedings of the Gateway Computing Environments Workshop (GCE), 14 Nov. 2010, New Orleans: 1–8.
- Minh BQ, Schmidt HA, Chernomor O, et al. 2020. IQ-TREE 2: New models and efficient methods for phylogenetic inference in the genomic era. *Molecular Biology and Evolution* 37: 1530–1534.
- Mitkowski NA, Browning M. 2004. Leptosphaerulina australis associated with intensively managed stands of *Poa annua* and *Agrostis palustris*. *Canadian Journal of Plant Pathology* 26: 193–198.
- Mongkolsamrit S, Khonsanit A, Thanakitipattana D, et al. 2020. Revisiting Metarhizium and the description of new species from Thailand. *Studies in Mycology* 95: 171–251.
- Monkai J, Liu J-K, Boonmee S, et al. 2013. Planistromellaceae (Botryosphaeriales). *Cryptogamie, Mycologie* 34: 45–77.
- Moreau PA, Rochet J, Richard F, et al. 2011. Taxonomy of *Alnus*-associated hypogeous species of Alpova and Melanogaster (Basidiomycota, Paxillaceae) in Europe. *Cryptogamie, Mycologie* 32: 33–62.
- Münzmay T, Saar G, Schmidt-Stohn G, et al. 2009. *Cortinarius laberiae* Münzmay, B. Oertel & Saar nov. spec. und zwei weitere, wenig bekannte Arten aus der Gattung *Cortinarius*, Untergattung Phlegmacium in Europa. *Journées Européennes du Cortinaire* 11: 32–40.
- Nag Raj TR, Kendrick WB. 1975. A monograph of Chalara and allied genera. Wilfred Laurier University Press, Waterloo, Ontario, Canada.
- Nguyen L-T, Schmidt HA, Von Haeseler A, et al. 2015. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Molecular Biology and Evolution* 32: 268–274.
- Noordeloos ME. 1980. Entoloma subgenus Nolanea in the Netherlands and adjacent regions with a reconnaissance of its remaining taxa in Europe. *Persoonia* 10: 427–534.
- Noordeloos ME, Hausknecht A. 1998. Rezente Rötlingsfunde aus Österreich und Italien. *Österreichische Zeitschrift für Pilzkunde* 7: 227–261.
- Noordeloos ME, Vila J, Jordal JB, et al. 2022. Contributions to the revision of the genus Entoloma (Basidiomycota, Agaricales) in Europe: six new species from subgenus Cyanula and typification of *E. incarnatofuscescens*. *Fungal Systematics and Evolution* 9: 87–97.
- Oktalira FT, May TW, Dearaley JDW, et al. 2021. Seven new Serendipita species associated with Australian terrestrial orchids. *Mycologia* 113: 968–987.
- Pegler DN. 1977. A preliminary Agaric flora of East Africa. *Kew Bulletin Additional Series* 6: 1–615.
- Peponi AL, Kolařík M, Bettini PP, et al. 2015. Morphological and molecular characterisation of Geosmithia species on European elms. *Fungal Biology* 119: 1063–1074.
- Petersen RH, Hughes KW. 2021. Collybiopsis and its type species, *Coramealis*. *Mycotaxon* 136: 263–349.
- Phookamsak R, Hyde KD, Jeewon R, et al. 2019. Fungal diversity notes 929–1035: taxonomic and phylogenetic contributions on genera and species of fungi. *Fungal Diversity* 95: 1–273.
- Pitt JI. 1980. The genus *Penicillium* and its teleomorphic states *Eupenicillium* and *Talaromyces*. London, Academic Press.
- Quaedvlieg W, Binder M, Groenewald JZ, et al. 2014. Introducing the Consolidated Species Concept to resolve species in the Teratosphaeriaceae. *Persoonia* 33: 1–40.
- Quin J, Yang ZL. 2016. Three new species of *Physalacria* from China, with a key to the Asian taxa. *Mycologia* 108: 215–226.
- Radic T, Likar M, Hancevic K, et al. 2021. Root-associated community composition and co-occurrence patterns of fungi in wild grapevine. *Fungal Ecology* 50: 101034.
- Ramirez C. 1982. Manual and atlas of the Penicillia. Amsterdam-New York-Oxford, Elsevier Biomedical Press.
- Raper KB, Thom C. 1949. A manual of the Penicillia. Baltimore, Waverly Press, INC for The Williams & Wilkins Company.
- Redhead SA. 1979. *Physalacria subpelata* sp. nov. from Hawaii. *Mycotaxon* 10: 46–48.
- Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574.
- Ronquist F, Teslenko M, Van der Mark P, et al. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61: 539–542.
- Rossman AY, Samuels GJ, Rogerson CT, et al. 1999. Genera of Bionectriaceae, Hypocreaceae and Nectriaceae (Hypocreales, Ascomycetes). *Studies in Mycology* 42: 1–248.
- Roux C. 1986. *Leptosphaerulina chartarum* sp.nov., the teleomorph of *Pithomyces chartarum*. *Transactions of the British Mycological Society* 86: 319–323.
- Sandoval-Denis, Gené J, Sutton DA, et al. 2016. Redefining Microascus, Scopulariopsis and allied genera. *Studies in Mycology* 36: 1–36.
- Santana-Ortiz B, Chen J, Parra L, et al. 2021. The genus Agaricus in the Caribbean II. Refined phylogeny of Agaricus subg. Spissicaules with description of two new sections and eight new species. *Mycological Progress* 20: 381–411.
- Seifert KA, Nickerson NL, Corlett M, et al. 2004. Devriesia, a new hyphomycete genus to accommodate heat-resistant, cladosporium-like fungi. *Canadian Journal of Botany* 82: 914–926.
- Shemesh H, Boaz BE, Millar CI, et al. 2020. Symbiotic interactions above treeline of long-lived pines: Mycorrhizal advantage of limber pine (*Pinus flexilis*) over Great Basin bristlecone pine (*Pinus longaeva*) at the seedling stage. *Journal of Ecology* 108: 908–916.
- Singer R. 1969. Mycoflora australis. *Beihefte zur Nova Hedwigia* 29: 1–405.
- Singer R. 1973. The genera Marasmiellus, Crepidotus, and Simocybe in the Neotropics. *Beihefte zur Nova Hedwigia* 44: 1–517.
- Singer R. 1976. Marasmeiae (Basidiomycetes-Tricholomataceae). *Flora Neotropica Monograph* 17: 1–347.
- Singer R. 1986. The Agaricales in modern taxonomy. 4th edn. Koeltz Scientific Books, Koenigstein, Germany.
- Singer R, Digilio APL. 1951. Pródromo de la Flora Agaricina Argentina. Lilloa 25: 5–461.
- Soop K, Dima B, Cooper JA, et al. 2019. A phylogenetic approach to a global supraspecific taxonomy of *Cortinarius* (Agaricales) with an emphasis on the southern mycota. *Persoonia* 42: 261–290.
- Stamatakis A. 2014. RAxML version 8: A toll for phylogenetic analysis and post-analysis of large phylogenomes. *Bioinformatics* 30: 1312–1313.
- Stamatakis A, Hoover P, Rougemont J. 2008. A rapid bootstrap algorithm for the RAxML web servers. *Systematic Biology* 57: 758–771.
- Stangl J. 1989. Die Gattung Inocybe in Bayern. *Hoppea* 46: 5–388.
- Strzałka B, Jankowiak R, Bilański P, et al. 2020. Two new species of Ophiostomatales (Sordariomycetes) associated with the bark beetle Dryocoetes alni from Poland. *MycoKeys* 68: 23.
- Strzałka B, Kolařík M, Jankowiak R. 2021. Geosmithia associated with hardwood-infesting bark and ambrosia beetles, with the description of three new species from Poland. *Antonie van Leeuwenhoek* 114: 169–194.
- Sutton BC. 1980. The Coelomycetes. *Fungi Imperfecti with Pycnidia, Acervuli and Stromata*. CMI, Kew, UK.
- Swofford DL. 2003. PAUP* 4.0b10. Phylogenetic Analysis Using Parsimony ("and other methods). Version 4. Sinauer Associates, Sunderland, MA, USA.
- Takahashi H. 2000. Two new species of Marasmiellus from eastern Honshu, Japan. *Mycoscience* 41: 467–472.
- Tamura K, Stecher G, Kumar S. 2021. MEGA 11: Molecular Evolutionary Genetics Analysis Version 11. *Molecular Biology and Evolution* 38: 3022–3027.
- Tamura K, Stecher G, Peterson D, et al. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725–2729.
- Tanaka K, Hirayama K, Yonezawa H, et al. 2009. Molecular taxonomy of bambusicolous fungi: Tetraplosphaeriaceae, a new pleosporalean family with tetraploa-like anamorphs. *Studies in Mycology* 64: 175–209.
- Tennakoon DS, Thambugala KM, De Silva ED, et al. 2019. Leaf litter saprobic Didymellaceae (Dothideomycetes): *Leptosphaerulina longiflora* sp. nov. and *Didymella sinensis*, a new record from *Roystonea regia*. *Asian Journal of Mycology* 2: 87–100.

- Thambugala KM, Wanasinghe DN, Phillips AJL, et al. 2017. Mycosphere notes 1–50: Grass (Poaceae) inhabiting Dothideomycetes. *Mycosphere* 8: 697–796.
- Thongklang N, Nawaz R, Khalid AN, et al. 2014. Morphological and molecular characterization of three *Agaricus* species from tropical Asia (Pakistan, Thailand) reveals a new group in section Xanthodermatei. *Mycologia* 106: 1220–1232.
- Tulasne LR, Tulasne C. 1851. *Fungi Hypogaei*. In: Klincksieck F (ed.), *Histoire et Monographie des Champignons Hypogés*. Paris, France.
- Vauras J, Larsson E. 2016. *Inocybe caprimulgi* and *I. lacunarum*, two new nodulose-spored species from Fennoscandia. *Karstenia* 55: 1–18.
- Videira S, Groenewald JZ, Nakashima C, et al. 2017. Mycosphaerellaceae – chaos or clarity? *Studies in Mycology* 87: 257–421.
- Videira SIR, Groenewald JZ, Braun U, et al. 2016. All that glitters is not Ramularia. *Studies in Mycology* 83: 49–163.
- Vila J, Carbó J, Caballero F, et al. 2013. A first approach to the study of the genus *Entoloma* subgenus *Nolanea* sensu lato using molecular and morphological data. *Fungi non Delineati* 66: 3–62.
- Visagie CM, Goodwell M, Nkwe DO. 2021. Aspergillus diversity from the Gcwihaba Cave in Botswana and description of one new species. *Fungal Systematics and Evolution* 8: 81–89.
- Walker J, Sutton BC, Pascoe IG. 1992. *Phaeoseptoria eucalypti* and similar fungi on Eucalyptus, with description of *Kirramyces* gen. nov. (Coelomycetes). *Mycological Research* 96: 911–924.
- Walz A, De Hoog GS. 1987. A new species of Cyphellophora. *Antonie van Leeuwenhoek* 53: 143–146.
- Wanasinghe DN, Phukhamsakda C, Hyde KD, et al. 2018. Fungal diversity notes 709–839: taxonomic and phylogenetic contributions to fungal taxa with an emphasis on fungi on Rosaceae. *Fungal Diversity* 89: 1–236.
- Wang XC, Chen K, Zeng ZQ, et al. 2017. Phylogeny and morphological analyses of *Penicillium* section *Sclerotiora* (Fungi) lead to the discovery of five new species. *Scientific Reports* 7: 8233.
- Warcup JH, Talbot PHB. 1967. Perfect states of Rhizoctonias associated with orchids. *New Phytologist* 66: 631–641.
- Weiss M, Waller F, Zuccaro A, et al. 2016. Sebacinales – one thousand and one interactions with land plants. *New Phytologist* 211: 20–40.
- Wijayawardene NN, Hyde KD, Wanasinghe DN, et al. 2016. Taxonomy and phylogeny of dematiaceous coelomycetes. *Fungal Diversity* 77: 1–316.
- Wijayawardene NN, Phillips AJL, Tibpromma S, et al. 2021. Looking for the undiscovered asexual taxa; case studies from lesser studied life modes and habitats. *Mycosphere* 12: 1290–1333.
- Wong PTW, Dong C, Stirling AM, et al. 2012. Two new *Magnaporthe* species pathogenic to warm-season turfgrasses in Australia. *Australasian Plant Pathology* 41: 321–329.
- Woudenberg JHC, Meijer M, Houbraken J, et al. 2017. *Scopulariopsis* and *scopulariopsis*-like species from indoor environments. *Studies in Mycology* 88: 1–35.
- Yaguchi T, Miyadoh S, Udagawa S. 1993. *Chromocleista*, a new cleistothelial genus with a *Geosmithia* anamorph. *Transactions of the Mycological Society of Japan* 34: 101–108.
- Yen LTH, Dung NL, Van Hop D, et al. 2012. *Condylospora vietnamensis*, a new Ingoldian hyphomycete isolated from fallen leaves in Vietnam. *Mycoscience* 53: 326–329.
- Yin M, Wingfield MJ, Zhou X, et al. 2019. Taxonomy and phylogeny of the *Leptographium olivaceum* complex (Ophiostomatales, Ascomycota), including descriptions of six new species from China and Europe. *MycoKeys* 60: 93.
- Yokoya K, Postel S, Fang R, et al. 2017. Endophytic fungal diversity of *Fragaria vesca*, a crop wild relative of strawberry, along environmental gradients within a small geographical area. *PeerJ* 5: e2860.
- Zhang D, Gao F, Jaković I, et al. 2020. PhyloSuite: An integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. *Molecular Ecology Resources* 20: 348–355.
- Zhang X, Li Y, Si H, et al. 2022. *Geosmithia* species associated with bark beetles from China, with the description of eleven new species. *Frontiers in Microbiology* 124. <https://doi.org/10.3389/fmicb.2022.820402>.
- Zhang Z-F, Zhou S-Y, Eurwilaichitr L, et al. 2021. Culturable mycobiota from Karst caves in China II, with descriptions of 33 new species. *Fungal Diversity* 106: 29–136.
- Zhou J-L, Su S-Y, Su H, et al. 2016. A description of eleven new species of *Agaricus* sections Xanthodermatei and Hondenses collected from Tibet and the surrounding areas. *Phytotaxa* 257: 99–121.