

Development and Validation of a Simplified Score to Predict Early Relapse in Newly Diagnosed Multiple Myeloma in a Pooled Dataset of 2,190 Patients



Gian Maria Zaccaria^{1,2}, Luca Bertamini¹, Maria Teresa Petrucci³, Massimo Offidani⁴, Paolo Corradini⁵, Andrea Capra¹, Alessandra Romano⁶, Anna Marina Liberati⁷, Donato Mannina⁸, Paolo de Fabritiis⁹, Nicola Cascavilla¹⁰, Marina Ruggeri¹, Roberto Mina¹, Francesca Patriarca¹¹, Giulia Benevolo¹², Angelo Belotti¹³, Gianluca Gaidano¹⁴, Arnon Nagler¹⁵, Roman Hájek^{16,17}, Andrew Spencer¹⁸, Pieter Sonneveld¹⁹, Pellegrino Musto^{20,21}, Mario Boccadoro¹, and Francesca Gay¹

ABSTRACT

Purpose: Despite the improvement of therapeutic regimens, several patients with multiple myeloma (MM) still experience early relapse (ER). This subset of patients currently represents an unmet medical need.

Experimental Design: We pooled data from seven European multicenter phase II/III clinical trials enrolling 2,190 patients with newly diagnosed MM from 2003 to 2017. Baseline patient evaluation included 14 clinically relevant features. Patients with complete data ($n = 1,218$) were split into training ($n = 844$) and validation sets ($n = 374$). In the training set, a univariate analysis and a multivariate logistic regression model on ER within 18 months (ER18) were made. The most accurate model was selected on the validation set. We also developed a dynamic version of the score by including response to treatment.

Results: The Simplified Early Relapse in Multiple Myeloma (S-ERMM) score was modeled on six features weighted by a

score: 5 points for high lactate dehydrogenase or t(4;14); 3 for del17p, abnormal albumin, or bone marrow plasma cells >60%; and 2 for λ free light chain. The S-ERMM identified three patient groups with different risks of ER18: Intermediate (Int) versus Low (OR = 2.39, $P < 0.001$) and High versus Low (OR = 5.59, $P < 0.001$). S-ERMM High/Int patients had significantly shorter overall survival (High vs. Low: HR = 3.24, $P < 0.001$; Int vs. Low: HR = 1.86, $P < 0.001$) and progression-free survival-2 (High vs. Low: HR = 2.89, $P < 0.001$; Int vs. Low: HR = 1.76, $P < 0.001$) than S-ERMM Low. The Dynamic S-ERMM (DS-ERMM) modulated the prognostic power of the S-ERMM.

Conclusions: On the basis of simple, widely available baseline features, the S-ERMM and DS-ERMM properly identified patients with different risks of ER and survival outcomes.

Introduction

In the past few years, the prognosis of patients with multiple myeloma (MM) has been markedly improved by the introduction of new drugs and better therapeutic strategies both at diagnosis and at relapse (1–4). Traditionally, the maximal benefit in terms of duration of remission has been observed with first-line therapies. With the use of

high-dose chemotherapy and autologous stem cell transplantation (ASCT) combined with novel agents or the adoption of multi-targeted agents including immunomodulatory (IMiD) agents, proteasome inhibitors (PI), and mAbs, the current median progression-free survival (PFS) of patients with newly diagnosed MM (NDMM) ranged between 41 and 50 months (4, 5). Despite this remarkable improvement, still a significant proportion of patients experiences an early

¹Myeloma Unit, Division of Hematology, University of Torino, Azienda Ospedaliero-Universitaria Città della Salute e della Scienza di Torino, Torino, Italy.

²Hematology and Cell Therapy Unit, IRCCS Istituto Tumori ‘Giovanni Paolo II,’ Bari, Italy.

³Hematology, Department of Translational and Precision Medicine, Azienda Ospedaliera Policlinico Umberto I, Sapienza University of Rome, Rome, Italy.

⁴Clinica di Ematologia, AOU Ospedali Riuniti di Ancona, Ancona, Italy.

⁵Divisione di Ematologia, Fondazione IRCCS Istituto Nazionale dei Tumori di Milano; Università degli Studi di Milano, Milano, Italy.

⁶Department of General Surgery and Medical-Surgical Specialties, Haematology Section, University of Catania, Catania, Italy.

⁷Università degli Studi di Perugia, Azienda Ospedaliera Santa Maria, Terni, Italy.

⁸Division of Hematology, Azienda Ospedaliera Papardo, Messina, Italy.

⁹Hematology, St. Eugenio Hospital ASL Roma 2, Tor Vergata University, Rome, Italy.

¹⁰Ematologia, Ospedale ‘Casa Sollievo della Sofferenza’ IRCCS, San Giovanni Rotondo, Italy.

¹¹Clinica Ematologica e Unità di Terapie Cellulari, Azienda Sanitaria Universitaria Friuli Centrale (ASUFC), Dipartimento di Area Medica (DAME), Università di Udine, Udine, Italy.

¹²SC Hematology, AO Città della Salute e della Scienza, Turin, Italy.

¹³Hematology Division, ASST Spedali Civili Brescia, Brescia, Italy.

¹⁴Division of Hematology, Department of Translational Medicine, University of Eastern Piedmont, Novara, Italy.

¹⁵Hematology Division, Chaim Sheba Medical Center, Tel Hashomer, Israel.

¹⁶Department of Haematology, University Hospital Ostrava, Ostrava, Czech Republic.

¹⁷Faculty of Medicine, University of Ostrava, Ostrava, Czech Republic.

¹⁸Alfred Health-Monash University, Melbourne, Australia.

¹⁹Department of Hematology, Erasmus MC Cancer Institute, Rotterdam, the Netherlands.

²⁰Department of Emergency and Organ Transplantation, ‘Aldo Moro’ University School of Medicine, Bari, Italy.

²¹Unit of Hematology and Stem Cell Transplantation, AOUC Policlinico, Bari, Italy.

Note: Supplementary data for this article are available at Clinical Cancer Research Online (<http://clincancerres.aacrjournals.org/>).

G.M. Zaccaria and L. Bertamini contributed equally as co-first authors of this article.

Supplementary documents: TRIPOD Checklist - Prediction Model Development

Corresponding Author: Francesca Gay, Myeloma Unit, Division of Hematology, University of Torino, Azienda Ospedaliero-Universitaria Città della Salute e della Scienza di Torino, Torino 10126, Italy. Phone: 0039-011-633-4279; Fax: 0039-011-633-4187; E-mail: fgay@ciudadellasalute.to.it

Clin Cancer Res 2021;27:3695–703

doi: 10.1158/1078-0432.CCR-21-0134

©2021 American Association for Cancer Research.

Translational Relevance

Despite the huge amount of literature, there is a lack of consensus on how to better predict early relapse (ER) in patients with multiple myeloma (MM). We pooled data from seven European clinical trials enrolling 2,190 patients with newly diagnosed MM from October 2003 to March 2017 to develop the Simplified Early Relapse in Multiple Myeloma (S-ERMM) score. This analysis provided further evidence of the critical role of predicting ER in patients with MM, which is strongly associated with poor outcome. The S-ERMM predicted ER by using simple and widely available baseline features. After external validation, the future development of this prognostic index may consider its combination with other static-risk features (genomic abnormalities, circulating tumor cells) and dynamic risk evaluation (response to the therapy such as minimal residual disease) for ER prediction. The identification of high-risk patients with dismal prognosis is the first step toward a better design of therapeutic approaches for this patient subgroup.

relapse (ER), which has been associated with a dismal prognosis. Several studies reported the association of baseline clinical features with ER (6–12), but there is no clear consensus on what the most important determinants are; as a matter of fact, even patients without well-known high-risk features at baseline may relapse early (7, 13). Data published so far mainly come from registries or from retrospective analyses that do not systematically consider updated standard-of-care risk assessments [e.g., cytogenetics, Revised International Staging System (R-ISS); ref. 12].

Unfortunately, a consensus on the appropriate definition of ER is also lacking. So far, it has been defined as relapse within 12 or 18 months from the start of induction treatment (10, 12), or else within 12 or 24 months from transplantation (6–9, 11, 14, 15).

Indeed, patients with ER have an inferior prognosis, as compared with patients who relapse later and, as such, represent a high-risk group and an unmet medical need (12, 14).

The correct identification of patient risk at baseline is the first step toward a risk-adaptive therapeutic approach. The aim of our analysis was to develop and validate the Simplified Early Relapse in Multiple Myeloma (S-ERMM), a score to predict the risk of ER based on widely available clinical and biological features. Thereafter, the S-ERMM score was remodulated during the patient clinical course by integrating response to therapy. We also aimed to correlate the S-ERMM with the long-term outcomes overall survival (OS) and progression-free survival-2 (PFS2).

Materials and Methods

Source of data and participants

Individual patient data from 2,190 patients with NDMM enrolled in seven multicenter European, open-label, phase II/III clinical trials evaluating novel agent-based therapies from October 2003 to March 2017 were pooled together and analyzed: NCT01093196, NCT01346787, NCT01857115, NCT01190787, NCT00551928, NCT01091831, and NCT02203643 (2, 3, 16–20). Each study was approved by ethics committees or institutional review boards at the respective study sites and was conducted in accordance with the Declaration of Helsinki; all patients provided written informed consent. All patients received new drugs (IMiD agents and/or PIs)

as upfront treatment, with or without transplantation. Trial details, treatment schedules, and eligibility criteria are reported in Supplementary Tables S1A and S1B.

Prognostic factors and outcomes

Data were retrieved from electronic case-report forms. All the available individual baseline features were analyzed. Age, creatinine levels, albumin, β 2-microglobulin (β 2m), and monoclonal plasma cells in the bone marrow (BMPC) were evaluated as continuous features. According to the International Myeloma Working Group recommendations, the percentage of BMPCs considered was the highest in case of discrepancy between BM biopsy and BM aspirate (21).

Free light chain (FLC; λ vs. κ), M-component subtype (IgA vs. others), lactate dehydrogenase (LDH) levels $>/\leq$ upper limit of normal (ULN), presence versus absence of plasmacytomas, presence versus absence of chromosomal abnormalities (CA) detected by interphase fluorescence *in situ* hybridization [iFISH; del17p, t(4;14), t(14;16), t(11;14)] were evaluated as categorical values. iFISH analysis was centralized in one laboratory (see the Supplementary Methods). High-risk CAs were defined as the presence of 17p deletion (del17p) and/or t(4;14) translocation and/or t(14;16) translocation (22). The cutoffs for del17p and IgH translocation were 10% and 15%, respectively. Baseline R-ISS stage (II/III vs. I) was also included in the prognostic factor evaluation (23). Patients with complete data were then split into training and validation sets. In the validation set, patients treated with more innovative and effective therapies were included. These patients also had a shorter median follow-up.

On the basis of the available literature, two cutoffs for ER were evaluated: 18 (ER18) and 24 (ER24) months from diagnosis. In the ER18 analysis, patients who died for reasons other than progressive disease (PD) or who withdrew consent within 18 months were excluded from the analysis because they were not at risk of progression for the entire first 18 months. Patients experiencing PD within 18 months from diagnosis were included in the ER18 population; those not experiencing PD within 18 months were included in the reference population. The reference population was then divided into two groups: patients experiencing PD after 18 months from diagnosis at the time of their last follow-up (Late Relapse group) and patients who were free from progression at the time of their last follow-up (No PD group).

Methods for the ER24 analysis were similar, but they included a cutoff of 24 months after diagnosis (see the Supplementary Appendix). The results regarding the best cutoff are reported in the main text of this contribution. For the sake of completeness, the other analyses are included in the Supplementary Appendix.

OS was calculated from the start of treatment until the date of death or the date the patient was last known to be alive. PFS2 was calculated from the start of treatment until the date of PD after the second line of treatment (second PD) or death (regardless of the cause of death), whichever came first. Other clinical endpoints are detailed in the Supplementary Methods.

Statistical analysis

From the training set, a univariate (UV) analysis on ER18 as outcome was performed according to chi-square and Kruskal-Wallis tests, as appropriate. Features with $P < 0.1$ were then tested in a multivariate (MV) logistic regression model. We compared two MV analyses, one including the R-ISS and the other including individual features defining the R-ISS (LDH, albumin, β 2m, and CAs). To account for potential confounders, each MV analysis was adjusted for age. Subsequently, each MV analysis was identified through a

backward selection based on the minimization of the Akaike information criterion to identify independent prognostic factors. Continuous parameters were not categorized *a priori* because this would have negatively affected the power of the analysis. After selecting the best MV model, the optimal cutoffs for the most significant continuous features were re-evaluated by spline function. MV models were used to estimate OR for ER18 risk, 95% confidence intervals (CI), and *P* values.

Each model was tested on the validation set by assessing the AUC, to select the most accurate model including individual features or features aggregated into the R-ISS.

Once the most accurate model was selected, three prognostic groups of patients with Low, Intermediate (Int), and High risk of ER were defined by categorizing the linear predictors of the final MV logistic model. Hence, two optimal cut-points were found maximizing the ORs defined by the MV model in the training set. A scalar score was thus proportionally assigned to each predictor according to the coefficients of the final MV model. As the linear score, two optimal cut-points were found maximizing the ORs defined by the MV model in both the training and validation sets. Thus, we developed the S-ERMM score, which identified three different groups of patients with Low, Int, and High risks of ER18. Other statistical survival analyses are detailed in the Supplementary Methods.

To integrate baseline prognostic evaluation and response to treatment, we developed the Dynamic S-ERMM (DS-ERMM), a logistic model that included S-ERMM score and achievement of at least a very good partial response (\geq VGPR). Because this score includes response, it should not be assessed at baseline, but at a subsequent timepoint after treatment, to remodulate patient risk during therapy (dynamic risk score). We therefore analyzed data from a landmark point, which was set at the median time to achieve \geq VGPR and included only patients who did not relapse before the landmark point. We assessed the role of \geq VGPR and S-ERMM in a MV logistic regression model to predict ER18. The DS-ERMM was modeled on the proportional coefficients obtained from the MV model. To measure the prognostic performances on this subcohort, we compared the concordance (C)-index assessed in both models (24).

The statistical analysis was performed using R (v.3.5.2). We used the Transparent Reporting of a Multivariable Prediction Model for Individual Prognosis Or Diagnosis (TRIPOD) criteria to validate our methods (25, 26).

Data sharing

After the publication of this article, data collected for this analysis and related documents will be made available to others upon reasonably justified request, which has to be written and addressed to the attention of the corresponding author Francesca Gay at the following e-mail address: fgay[at]cittadellasalute.to.it. The corresponding author Francesca Gay is responsible to evaluate and eventually accept or refuse every request to disclose data and their related documents, in compliance with the ethical approval conditions, in compliance with applicable laws and regulations, and in conformance with the agreements in place with the involved subjects, the participating institutions, and all the other parties directly or indirectly involved in the participation, conduct, development, management, and evaluation of this analysis.

Results

Patient characteristics

Data from 2,190 patients were available; 3 patients were excluded because of screening failure.

In the ER18 analysis, 50 patients died for reasons other than PD and 51 withdrew their consent within 18 months and were excluded; patients eligible for the analyses were 2,086. Patients with complete data ($n = 1,218$) were then split into training ($n = 844$) and validation ($n = 374$) sets and included in the logistic regression analysis.

Training set: in the overall population [median follow-up 70 months; interquartile range (IQR) = 48–81 months], the median age was 66 years, 73% of patients presented with R-ISS stage II/III, 10% with LDH>ULN, 14% with del17p, and 14% with t(4;14). Twenty-nine percent of patients had BMPCs > 60% and 36% had λ FLC. A total of 312 of 844 (37%) patients experienced ER18. Patients in the ER18 versus the reference population were significantly older ($P = 0.026$), and had higher β 2m ($P < 0.001$) and lower albumin ($P < 0.001$) levels; a higher proportion of patients had LDH>ULN ($P = 0.001$), t(4;14) ($P < 0.001$), R-ISS stage II/III ($P < 0.001$), del17p ($P = 0.005$), and BMPCs > 60% ($P = 0.001$; **Table 1**).

Validation set: in the overall population (median follow-up 35 months; IQR, 29–41), the median age was 57 years, which was significantly lower ($P < 0.001$) than that in the training set. Patients who experienced ER18 were 61 of 374 (16%). The distribution of baseline features between ER18 and the reference population was similar to that in the training set, except for the absence of significant differences in the proportion of patients with BMPCs > 60% and del17p, although this may be related to the smaller sample size (**Table 1**).

The median time to \geq VGPR was 9 months in the training set and 3 months in the validation set. \geq VGPR at 9 months was achieved in 40% and 81% of patients in the training and validation sets, respectively.

Best model of ER

On the basis of the UV analysis of patients who experienced ER18, 10 of 14 features were included in the MV analysis: age, FLC, BMPCs, del17p, t(4;14), t(14;16), albumin, β 2m, LDH, and R-ISS stage. In the MV analysis incorporating the R-ISS, age, R-ISS II/III versus I, and increased BMPCs increased the risk of ER18. When the MV analysis was performed including single features defining the R-ISS, increased BMPCs, λ FLC, LDH>ULN, presence of del17p, and t(4;14) increased the probability of ER18 (**Table 2**).

Each MV model was then tested on the validation set. The AUC was 0.62 (95% CI = 0.55–0.69) for the ER18 model including the R-ISS and 0.66 (95% CI = 0.58–0.73) for the ER18 model incorporating individual features. The ER18 model incorporating individual features resulted in the highest AUC (0.66) and was therefore selected to develop the S-ERMM score.

UV and MV ER24 analyses and the AUC in the validation set are reported in the Supplementary Results and in Supplementary Table S3.

S-ERMM score

The ER18 linear index was calculated as $0.047 \times \text{BMPCs \%}/5 + 0.589 \times \text{LDH}/\text{ULN}$ (IF LDH>ULN) + $0.459 \times \text{del17p}$ (IF present) + $0.705 \times \text{t}(4;14)$ (IF present) + $0.293 \times \text{FLC}$ (IF λ) – $0.284 \times \text{albumin}$.

In the training set, the linear score significantly discriminated three patient groups (High, Int, and Low risk) with significantly different risks of ER18 (**Fig. 1**).

BMPC and albumin levels were dichotomized according to the optimal cutoff: high BMPC level if >60% and abnormal albumin level if ≤ 3.5 or ≥ 5 (Supplementary Fig. S1).

The S-ERMM score (<https://sermm.emnitaly.org/>) was mathematically consistent with the linear index and was defined including six features identified in the MV analysis: 5 points for LDH>ULN or the

Table 1. Patient characteristics of the overall population, stratified according to the ER18 outcome as training set and validation set.

	POOLED SET Overall population	TRAINING SET ^a				VALIDATION SET ^a			
		Overall population	ER18 population	Reference population	P	Overall population	ER18 population	Reference population	P
No. of patients (%)	2,190	844	312 (37)	532 (63)	—	374	61 (16)	313 (84)	—
Age, y: median (IQR)	63.0 (56.0–72.0)	66.0 (57.0–73.0)	68.0 (58.0–75.0)	65.0 (57.0–73.0)	0.026	57.0 (51.0–62.0)	56.0 (48.0–62.0)	58.0 (52.0–62.0)	0.173
Missing, N (%)	0 (0)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
Albumin, g/dL: median (IQR)	3.8 (3.4–4.2)	3.8 (3.4–4.2)	3.7 (3.2–4.1)	3.9 (3.5–4.2)	<0.001	3.9 (3.5–4.3)	3.7 (3.4–4.1)	3.9 (3.5–4.3)	0.046
Missing, N (%)	10 (0.5)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
β2m, mg/dL: median (IQR)	3.4 (2.4–5.1)	3.6 (2.5–5.4)	4.1 (2.8–6.1)	3.3 (2.4–5.0)	<0.001	2.9 (2.0–4.2)	3.7 (2.2–5.8)	2.8 (2.0–4.0)	0.015
Missing, N (%)	8 (0.4)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
LDH>ULN, N (%)	211 (9.6)	83 (10)	45 (14)	38 (7)	0.001	54 (14)	17 (28)	37 (12)	0.002
Missing, N (%)	267 (12.2)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
del17p, N (%)	246 (11.2)	121 (14)	59 (19)	62 (12)	0.005	52 (14)	11 (18)	41 (13)	0.414
Missing, N (%)	477 (21.8)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
t(4;14), N (%)	228 (10.4)	117 (14)	63 (20)	54 (10)	<0.001	57 (15)	17 (28)	40 (13)	0.005
Missing, N (%)	482 (22.0)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
t(11;14), N (%)	341 (15.5)	156 (18)	50 (16)	106 (20)	0.188	87 (23)	14 (23)	73 (23)	1
Missing, N (%)	522 (23.8)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
t(14;16), N (%)	78 (3.6)	32 (4)	16 (5)	16 (3)	0.171	19 (5)	4 (7)	15 (5)	0.798
Missing, N (%)	506 (23.1)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
R-ISS, II/III: N (%)	1388 (63.5)	613 (73)	255 (82)	358 (67)	<0.001	250 (67)	53 (87)	197 (63)	<0.001
Missing, N (%)	388 (17.7)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
Creatinine, mg/dL: median (IQR)	0.9 (0.7–1.1)	0.9 (0.8–1.2)	1.0 (0.8–1.2)	0.9 (0.8–1.1)	0.136	0.8 (0.7–1.0)	0.8 (0.7–1.2)	0.8 (0.7–1.0)	0.427
Missing, N (%)	67 (3.1)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
BMPCs >60%, N (%)	612 (28)	243 (29)	112 (36)	131 (25)	0.001	139 (37)	27 (44)	112 (36)	0.267
Missing, N (%)	121 (5.5)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
FLC, λ, N (%)	727 (36)	301 (36)	123 (39)	178 (33)	0.095	143 (38)	18 (30)	125 (40)	0.165
Missing, N (%)	0 (0)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
M component, IgA, N (%)	451 (21)	186 (22)	65 (21)	121 (23)	0.575	57 (15)	10 (16)	47 (15)	0.937
Missing, N (%)	3 (0.1)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	
Plasmacytomas, N (%)	266 (12)	78 (9)	27 (9)	51 (10)	0.743	49 (13)	10 (16)	39 (12)	0.532
Missing, N (%)	0 (0)	0 (0)	0 (0)	0 (0)		0 (0)	0 (0)	0 (0)	

Abbreviations: BMPCs, bone marrow plasma cells; β2m, β2-microglobulin; del17p, 17p deletion; ER18, early relapse within 18 months from diagnosis; FLC, free light chains; IQR, interquartile range; LDH, lactate dehydrogenase; M, monoclonal; N, number; P, P value; R-ISS, Revised International Staging System stage; t, translocation; ULN, upper limit of normal; y, years.

^aPatients with complete data only.

presence of t(4;14); 3 points for the presence of del17p, abnormal albumin, and BMPCs > 60%; and 2 points for the presence of λ FLC (Fig. 1).

The Low-risk group included patients with a total score ≤5 (68% of patients in the training set, 29% of whom experienced an ER18); the Int-risk group included patients with a total score between 6 and 10 (25% of patients in the training set, 50% of whom with ER18); and the High-risk group included patients with a total score ≥11 (7% of patients in the training set, 70% of whom with ER18). In the training set, the S-ERMM significantly discriminated three groups of patients with different risks of ER18: Int versus Low (OR = 2.39, 95% CI = 1.73–3.30, *P* < 0.001) and High versus Low (OR = 5.59, 95% CI = 3.08–10.16, *P* < 0.001). The S-ERMM was confirmed in the validation set: Int versus Low (OR = 2.27, 95% CI = 1.23–4.17, *P* = 0.008) and High versus Low (OR = 4.87, 95% CI = 2.01–11.76, *P* = 0.001). The impact of the S-ERMM on the ER18 risk was higher than that of each single feature.

In the DS-ERMM analyses, the training population (*n* = 673) included patients evaluable for response at 9 months (median time to ≥VGPR), 162 (24%) of whom experienced ER18. In this population, both the S-ERMM and the achievement of ≥VGPR were statistically independent predictors of ER in the MV analysis (Supplementary Fig. S2).

The DS-ERMM score was defined as the S-ERMM score obtained at baseline minus 4 points in case of achievement of ≥VGPR. Patients who reached the 9-month cutoff (which was not reached by 171 patients, 150 of whom relapsed/died before), were thus reclassified in three groups: the Low-risk group included 250 patients (37%) with a total score ≤0 (only 12% of whom experienced an ER18); the Int-risk group included 271 patients (40%) with a total score between 1 and 5 (only 24% of whom with ER18); and the High-risk group included 152 patients (23%) with a total score ≥6 (45% of whom with ER18). These three groups had different risks of ER18 (Supplementary Fig. S3): Int versus Low (OR = 2.36, 95% CI = 1.46–3.80, *P* < 0.001) and High versus Low (OR = 6.34, 95% CI = 3.84–10.46, *P* < 0.001).

In the validation set, there were no significant differences in terms of risk of ER18 between DS-ERMM Int versus Low, while a trend toward a higher risk was observed in the High versus Low comparison (OR = 2.40, *P* = 0.09; see the Supplementary Results).

Following the application of the S-ERMM score at baseline and the remodulation of patient risk at 9 months according to DS-ERMM (for those patients who did not relapse during the first 9 months), 20% of patients in the total population of the training set were classified as High-risk patients, 39% as Int-risk patients, and 41% as Low-risk patients.

Table 2. UV and MV analyses of the baseline features to predict ER18.

	ER18 ^a					
	Analysis including R-ISS			Analysis including single features		
	UV Analysis	MV Analysis	P	UV Analysis	MV Analysis	P
	P	OR (95% CI)	P	P	OR (95% CI)	P
Age (increased by 1 y)	0.026	1.01 (1.00-1.02)	0.113	0.026	1.01 (1.00-1.03)	0.094
Albumin (increased by 1 mg/dL)				<0.001	0.75 (0.60-0.95)	0.015
β2m (increased by 1 mg/dL)				<0.001	— ^b	— ^b
LDH (> vs. ≤ ULN)				0.001	2.03 (1.27-3.26)	0.003
del17p (presence vs. no)				0.005	1.65 (1.10-2.47)	0.016
t(4;14) (presence vs. no)				<0.001	2.12 (1.40-3.19)	<0.001
R-ISS (II/III vs. I)	<0.001	1.91 (1.35-2.71)	<0.001			
BMPCs % (increased by 5%)	<0.001	1.05 (1.02-1.08)	<0.001	<0.001	1.06 (1.03-1.09)	<0.001
FLC (λ vs. κ)	0.095	— ^b	— ^b	0.095	1.31 (0.97-1.78)	0.076

Abbreviations: BMPCs, bone marrow plasma cells; β2m, β2-microglobulin; CI, confidence interval; del17p, 17p deletion; ER18, early relapse within 18 months from diagnosis; FLC, free light chains; LDH, lactate dehydrogenase; M, monoclonal; MV, multivariate; OR, odds ratio; P, P value; R-ISS, Revised International Staging System stage; t, translocation; ULN, upper limit of normal; UV, univariate; y, years.

^aOnly significant features (P < 0.1) in UV analysis and age were included.

^bExcluded before the MV analysis by the Akaike information criterion.

Survival analysis

A landmark analysis with landmark point at 18 months was performed. OS and PFS2 were significantly shorter in the ER18 population than in the reference population and the Late Relapse and No Relapse populations (Supplementary Figs. S4A-S4D). Similarly, ER18 patients showed an inferior outcome after relapse (Supplementary Results; Supplementary Fig. S5).

The median OS was 31.5 months in patients with S-ERMM High, 59.5 with S-ERMM Int, and not reached (NR) with S-ERMM Low. Median PFS2 was 19.8 months in patients with S-ERMM High, 40.0 months with S-ERMM Int, and 62.3 months with S-ERMM

Low. OS and PFS2 were significantly shorter in S-ERMM Int versus S-ERMM Low patients (OS, HR = 1.86, 95% CI = 1.48-2.33; PFS2, HR = 1.76, 95% CI = 1.45-2.14; both P < 0.001) and in S-ERMM High versus S-ERMM Int patients (OS, HR = 1.74, 95% CI = 1.22-2.50, P = 0.002; PFS2, HR = 1.64, 95% CI = 1.18-2.28; P = 0.003; Fig. 2). The median PFS was 31.6 months in S-ERMM Low, 17.3 months in S-ERMM Int, and 13.2 months in S-ERMM High patients.

Subgroup analyses for OS according to first-line treatment confirmed the prognostic role of the S-ERMM in ASCT-ineligible patients (Int vs. Low, HR = 1.75, 95% CI = 1.30-2.35, P < 0.001; High

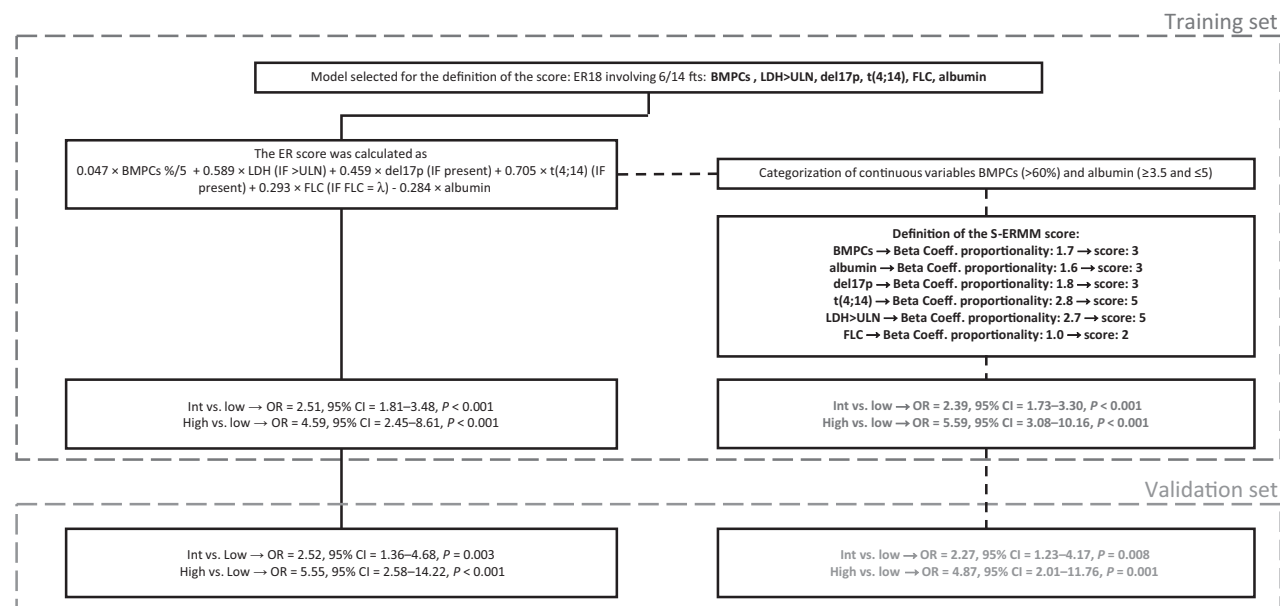


Figure 1.

Flowchart of the S-ERMM score construction. alb, albumin; BMPCs, bone marrow plasma cells; CI, confidence interval; coeff., coefficient; del17p, 17p deletion; ER, early relapse; FLC, free light chains; fts, features; High, high; Int, intermediate; LDH, lactate dehydrogenase; Low, low; OR, odds ratio; P, P value; S-ERMM, Simplified Early Relapse in Multiple Myeloma score; t, translocation; ULN, upper limit of normal.

Downloaded from http://aacrjournals.org/clinccancerres/article-pdf/27/13/3695/3086079/3695.pdf by guest on 11 December 2024

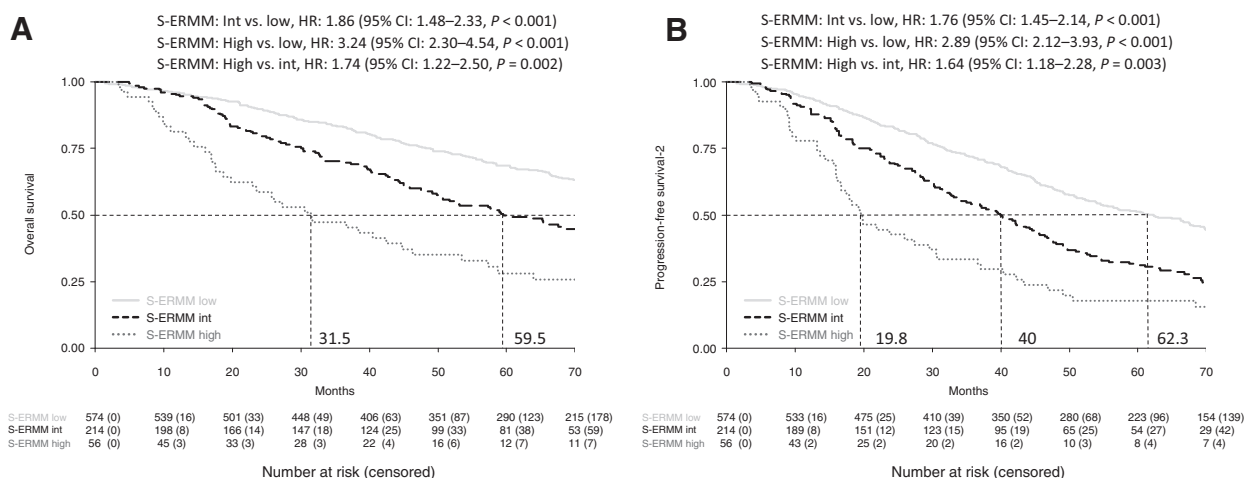


Figure 2. OS (A) and PFS2 (B) stratified by S-ERMM score. A, OS: S-ERMM Int versus S-ERMM Low, S-ERMM High versus S-ERMM Low, and S-ERMM High versus S-ERMM Int. B, PFS2: S-ERMM Int versus S-ERMM Low, S-ERMM High versus S-ERMM Low, and S-ERMM High versus S-ERMM Int. CI, confidence interval; HR, hazard ratio; Int, intermediate; OS, overall survival; P, P value; PFS2, progression-free survival-2; S-ERMM, Simplified Early Relapse in Multiple Myeloma score.

vs. Int, HR = 1.85, 95% CI = 1.10–3.11, $P = 0.020$); in ASCT-eligible patients (Int vs. Low, HR = 1.94, 95% CI = 1.36–2.78, $P < 0.001$; High vs. Int, HR = 1.81, 95% CI = 1.09–3.01, $P = 0.022$; Supplementary Fig. S6); in patients treated with PIs (Int vs. Low, HR = 1.73, 95% CI = 0.93–3.20, $P = 0.083$; High vs. Int, HR = 3.13, 95% CI = 1.21–8.08, $P = 0.018$); and in patients treated with IMiD agents (Int vs. Low, HR = 1.85, 95% CI = 1.45–2.37, $P < 0.001$; High vs. Int, HR = 1.64, 95% CI = 1.11–2.43, $P = 0.013$).

According to the DS-ERMM, OS and PFS2 were significantly shorter in DS-ERMM Int versus DS-ERMM Low patients (OS, HR = 1.96, 95% CI = 1.44–2.66; PFS2, HR = 1.86, 95% CI = 1.44–2.38; both $P < 0.001$) and in DS-ERMM High versus DS-ERMM Low

patients (OS, HR = 3.28, 95% CI = 2.37–4.54, $P < 0.001$; PFS2, HR = 2.91, 95% CI = 2.22–3.82; $P < 0.001$; Fig. 3).

Discussion

Several studies reported dismal survival outcomes in patients with MM experiencing an ER; however, the definition of ER varies from study to study, and consensus is still lacking. Also, the impact of well-known disease-related risk factors (e.g., albumin, β 2m, CAs by iFISH, and LDH) on the risk of ER has not been thoroughly assessed in patients with NDMM. The correct evaluation of baseline ER risk thus remains an unmet medical need.

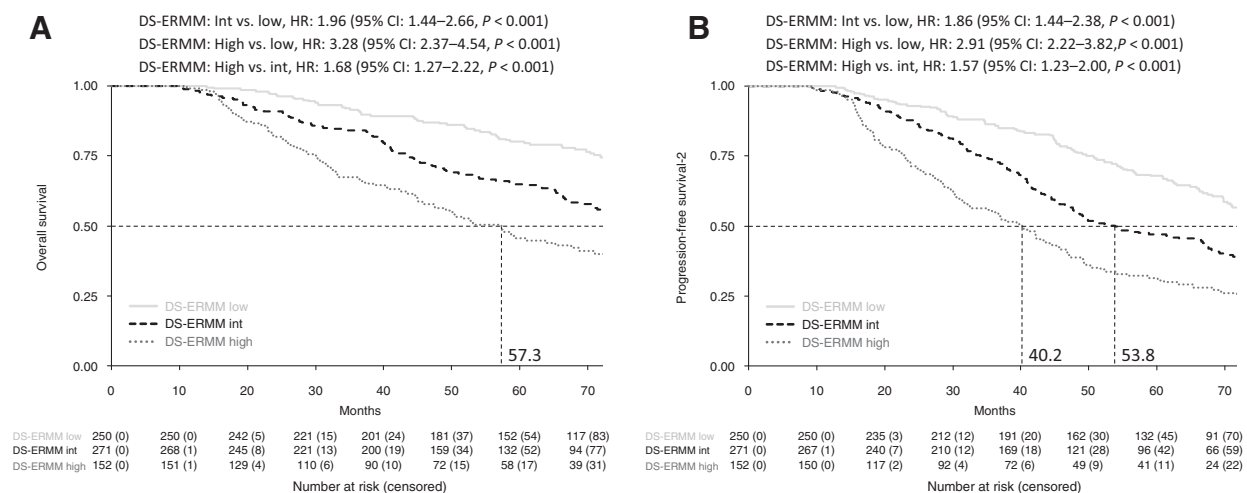


Figure 3. OS (A) and PFS2 (B) according to the DS-ERMM score. Landmark analysis at 9 months, corresponding to the median time to achieve \geq VGPR. A, OS: DS-ERMM Int versus DS-ERMM Low, DS-ERMM High versus DS-ERMM Low, and DS-ERMM High versus DS-ERMM Int. B, PFS2: DS-ERMM Int versus DS-ERMM Low, DS-ERMM High versus DS-ERMM Low, and DS-ERMM High versus DS-ERMM Int. CI, confidence interval; DS-ERMM, Dynamic Simplified Early Relapse in Multiple Myeloma score; HR, hazard ratio; Int, intermediate; OS, overall survival; P, P value; PFS2, progression-free survival-2; VGPR, very good partial response.

We confirmed ER with an 18-month cutoff as a strong predictor of the long-term outcomes OS and PFS2 in the context of novel treatment approaches (e.g., PIs, IMiD agents, and ASCT). Our decision of adopting the 18-month cutoff for the definition of ER was also supported by the available literature, in which most of the studies defined ER as both 18 months from diagnosis and 12 months from ASCT (6–10, 12, 27).

We developed the S-ERMM score by identifying and integrating six features that predicted the risk of ER [presence of t(4;14), del17p, LDH>ULN, BMPCs > 60%, abnormal albumin, and λ FLC]. The S-ERMM is a simple tool enabling the identification of three patient groups with significantly different risks of ER (Fig. 2) and significantly different PFS2 and OS. In particular, S-ERMM High patients had a median OS of 31 months, significantly shorter than that of S-ERMM Int (median, 60 months) and S-ERMM Low patients (median NR after 6 years of follow-up).

Although several studies tried to correlate the ER risk with clinical and biological features (6–9, 11, 12, 14), most of them did not cover all of the recognized MM prognostic factors [ref. 23; e.g., extensive CA analysis (8, 9, 14) and LDH assessment (10)]. In our series, we included widely available and well-recognized baseline features. The S-ERMM score included albumin levels (which reflect the inflammatory state at diagnosis; ref. 28), high-risk CAs (del17p and t(4;14), associated with a biologically aggressive disease), and high LDH (29) and BMPC levels (associated with tumor burden; ref. 30).

The majority of analyses published so far on ER in MM are single-center or retrospective studies. To the best of our knowledge, only Bygrave and colleagues analyzed young ASCT-eligible patients enrolled in a single clinical trial (7). Indeed, in our analysis, data from clinical trials underwent a systematic data assessment, with baseline features assessment, centralized laboratory analyses, and uniform evaluation of response and clinical outcomes (31). In this light, the development and validation of the S-ERMM score in a population consisting of both young (transplant-eligible) and elderly (>65 years) patients enrolled in four phase II/III clinical trials treated with novel agents from different drug classes with or without ASCT support the application of this score to patients with NDMM. On the other hand, this is a selected population of European clinical trials that indeed needs validation in real-life settings.

Response to therapy is a strong predictor of better OS and PFS2 (15), and the achievement of a deep response [minimal residual disease (MRD) negativity] may abrogate the poor prognosis conferred by high-risk FISH at diagnosis. Therefore, the importance of integrating static (baseline) and dynamic (response) prognostic features led to the incorporation of response to treatment (\geq VGPR) into the S-ERMM score. The assessment of the S-ERMM score at the time of diagnosis and the remodulation of patient risk at 9 months (for those patients who did not relapse during the first 9 months) improved our ability to detect ER patients. In fact, in the initial population of our analysis, only 7% of patients were included in the high-risk group (S-ERMM High), while 68% of patients were included in the S-ERMM Low group (29% of whom had an ER18) and 25% in the S-ERMM Int group (50% of whom had an ER18). Of note, in the DS-ERMM analysis, the Low-risk group included only 37% of patients (with only 12% who had an ER18), and the Int-risk group 40% of patients (with only 24% who had an ER18). Ultimately, using sequentially these two scores in the overall population, 20% of patients were determined to be at High risk, 39% at Int risk, and 41% at Low risk. This improvement in the evaluation of patient risk of ER highlighted the role of the dynamic modulation of patient risk at baseline.

Unfortunately, in the validation set, there were no significant differences in terms of risk of ER18 between DS-ERMM Int versus Low, while trends toward a higher risk were observed in the High versus Low comparison.

Of note, the optimal response (degree and timing) to be incorporated as a dynamic factor should consider the type of patient population and the availability of treatment options: these two factors determine the choice of a specific therapy, with different degrees of efficacy and time to best response. In the validation set, the rate of \geq VGPR was definitely higher than in the training set, and the median time to response was lower. We presume that the assessment of a deeper response, such as the achievement of MRD negativity, could better discriminate patients in the context of novel, highly effective therapies. Unfortunately, MRD evaluation was not available in most of the trials included in the training set and could not be used as optimal response to recalculate the risk of ER after therapy. Still, our main aim was to identify patients at risk of ER using risk assessment at diagnosis, and the S-ERMM score was prognostic in the context of both older (training set) and more recent (validation set) drug regimens.

Our analysis has some limitations. First, the risk classification based on the S-ERMM score was designed to better identify patients at high risk of ER and, as a consequence, was unbalanced, with only a small proportion of patients in the S-ERMM High group. Nevertheless, the risk group stratification improved after the remodulation of risk assessment by using the DS-ERMM score.

Another limitation was the low number of patients treated upfront with a combination of PIs and IMiD agents in the training set, since this currently represents a standard of care for both young and elderly patients. Nevertheless, our results were validated in a population who received intensive and effective induction and consolidation therapies including the second-generation PI carfilzomib with or without IMiD agents and ASCT intensification. In this context, the S-ERMM maintained its prognostic role, but the percentage of patients experiencing ER was definitely lower than that reported in the training set.

In conclusion, we were able to correctly classify a good proportion of patients who experienced ER by assessing the S-ERMM score at baseline and remodulating patient risk at 9 months with the DS-ERMM score. An external validation of the S-ERMM and DS-ERMM scores is warranted, especially in patients treated with combinations of PIs, IMiD agents, and anti-CD38 mAbs. Our ability to predict ER could also be improved by the inclusion of other risk features at baseline with known prognostic impact, such as amp(1q21), TP53 mutational status, and circulating plasma cells (27, 32, 33). Unfortunately, these data were not available for this analysis.

The correct identification of patient risk at diagnosis and during therapy is an essential step toward a risk-adapted approach, the cure of patients, and the prevention of overtreatment and undertreatment.

Authors' Disclosures

M.T. Petrucci reports personal fees from Celgene, Janssen-Cilag, BMS, Amgen, Takeda, Sanofi, and Karyopharm outside the submitted work. M. Offidani reports grants and personal fees from Amgen and Celgene outside the submitted work. A.M. Liberati reports grants and other support from Bristol-Myers Squibb, Takeda, Roche, Celgene, and AbbVie; other support from Sanofi, Novartis, Iqvia, Verastem, and Servier; and grants from Novartis, Janssen, Amgen, Incyte, Beigene, Oncopeptides, Karyopharm, Archigen, Debiopharm, Morphosys, Fibrogen, and Onconova Therapeutics, Inc. outside the submitted work. R. Mina reports personal fees from Sanofi, Celgene, Takeda, and Janssen outside the submitted work. A. Belotti reports other support from Amgen, Janssen, and Celgene outside the submitted work. G. Gaidano reports personal fees from AbbVie, Janssen, and AstraZeneca outside the

submitted work. R. Hájek reports personal fees from Janssen, Abbvie, BMS, and Pharma Mar, as well as grants and personal fees from Amgen, Celgene, Takeda, and Novartis outside the submitted work. P. Sonneveld has served on the advisory boards for Amgen, Celgene, Genenta, Janssen, Seattle Genetics, Takeda, and Karyopharm. P. Musto reports personal fees from Celgene, Janssen-Cilag, Takeda, Novartis, Bristol-Myers Squibb, AbbVie, Amgen, Jazz, Gilead, Sanofi, and Glaxo-SmithKline outside the submitted work. M. Boccadoro reports personal fees and other support from Sanofi, Celgene, Amgen, Janssen, Novartis, and Bristol-Myers Squibb; personal fees from AbbVie and GSK; and other support from Mundipharma outside the submitted work. F. Gay reports personal fees from Amgen, Celgene, Janssen, Takeda, Bristol-Myers Squibb, AbbVie, GSK, Roche, Adaptive Biotechnologies, Oncopeptides, and Bluebird bio outside the submitted work. No disclosures were reported by the other authors.

Authors' Contributions

G.M. Zaccaria: Conceptualization, resources, data curation, software, formal analysis, validation, investigation, visualization, methodology, writing—original draft, writing—review and editing. **L. Bertamini:** Conceptualization, resources, data curation, formal analysis, validation, investigation, visualization, methodology, writing—original draft, writing—review and editing. **M.T. Petrucci:** Resources, data curation, validation, investigation, writing—review and editing. **M. Offidani:** Resources, data curation, validation, investigation, writing—review and editing. **P. Corradini:** Resources, data curation, validation, investigation, writing—review and editing. **A. Capra:** Resources, data curation, software, formal analysis, validation, investigation, visualization, writing—original draft, writing—review and editing. **A. Romano:** Resources, data curation, validation, investigation, writing—review and editing. **A.M. Liberati:** Resources, data curation, validation, investigation, writing—review and editing. **D. Mannina:** Resources, data curation, validation, investigation, writing—review and editing. **P. de Fabritiis:** Resources, data curation, validation, investigation, writing—review and editing. **N. Cascavilla:** Resources, data curation, validation, investigation, writing—review and editing. **M. Ruggeri:** Resources, data curation, validation,

investigation, writing—review and editing. **R. Mina:** Resources, data curation, validation, investigation, writing—original draft, writing—review and editing. **F. Patriarca:** Resources, data curation, validation, investigation, writing—review and editing. **G. Benevolo:** Resources, data curation, validation, investigation, writing—review and editing. **A. Belotti:** Resources, data curation, validation, investigation, writing—review and editing. **G. Gaidano:** Resources, data curation, validation, investigation, writing—review and editing. **A. Nagler:** Resources, data curation, validation, investigation, writing—review and editing. **R. Hájek:** Resources, data curation, validation, investigation, writing—review and editing. **A. Spencer:** Resources, data curation, validation, investigation, writing—review and editing. **P. Sonneveld:** Resources, data curation, validation, investigation, writing—review and editing. **P. Musto:** Resources, data curation, validation, investigation, writing—review and editing. **M. Boccadoro:** Conceptualization, resources, data curation, supervision, validation, investigation, writing—review and editing. **F. Gay:** Conceptualization, resources, data curation, formal analysis, supervision, validation, investigation, methodology, writing—original draft, writing—review and editing.

Acknowledgments

G.M. Zaccaria acknowledges the support of European Myeloma Network (EMN) Research Italy. The authors wish to thank all the study participants and referring clinicians for their valuable contributions.

No funding was provided for this contribution.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received January 12, 2021; revised March 12, 2021; accepted April 26, 2021; published first April 29, 2021.

References

- Kumar SK, Rajkumar SV, Dispenzieri A, Lacy MQ, Hayman SR, Buadi FK, et al. Improved survival in multiple myeloma and the impact of novel therapies. *Blood* 2008;111:2516–20.
- Palumbo A, Cavallo F, Gay F, Di Raimondo F, Yehuda DB, Petrucci MT, et al. Autologous transplantation and maintenance therapy in multiple myeloma. *N Engl J Med* 2014;371:895–905.
- Gay F, Oliva S, Petrucci MT, Conticello C, Catalano L, Corradini P, et al. Chemotherapy plus lenalidomide versus autologous transplantation, followed by lenalidomide plus prednisone versus lenalidomide maintenance, in patients with multiple myeloma: a randomised, multicentre, phase 3 trial. *Lancet Oncol* 2015;16:1617–29.
- Attal M, Lauwers-cances V, Hulin C, Leleu X, Caillot D, Escoffre M, et al. Lenalidomide, bortezomib, and dexamethasone with transplantation for myeloma. *N Engl J Med* 2017;376:1311–20.
- Durie BGM, Hoering A, Sexton R, Abidi MH, Epstein J, Rajkumar SV, et al. Longer term follow-up of the randomized phase III trial SWOG S0777: bortezomib, lenalidomide and dexamethasone vs. lenalidomide and dexamethasone in patients (Pts) with previously untreated multiple myeloma without an intent for immediate autologous stem. *Blood Cancer J* 2020;10:53.
- Kumar S, Mahmood ST, Lacy MQ, Dispenzieri A, Hayman SR, Buadi FK, et al. Impact of early relapse after auto-SCT for multiple myeloma. *Bone Marrow Transplant* 2008;42:413–20.
- Bygrave C, Pawlyn C, Davies F, Craig Z, Cairns D, Hockaday A, et al. Early relapse after high-dose melphalan autologous stem cell transplant predicts inferior survival and is associated with high disease burden and genetically high-risk disease in multiple myeloma. *Br J Haematol* 2021;193:151–55.
- Jimenez-Zepeda VH, Reece DE, Trudel S, Chen C, Tiedemann R, Kukreti V. Early relapse after single auto-SCT for multiple myeloma is a major predictor of survival in the era of novel agents. *Bone Marrow Transplant* 2015;50:204–8.
- Ong SY, De Mel S, Chen YX, Ooi MG, Surendran S, Lin A, et al. Early relapse post autologous transplant is a stronger predictor of survival compared with pretreatment patient factors in the novel agent era: analysis of the Singapore Multiple Myeloma Working Group. *Bone Marrow Transplant* 2016;51:933–7.
- Corre J, Montes L, Martin E, Perrot A, Caillot D, Leleu X, et al. Early relapse after autologous transplant for myeloma is associated with poor survival regardless of cytogenetic risk. *Haematologica* 2019;105:e480–3.
- Helm-Petersen S, Sørrig R, Klausen TW, Preiss B, Frolund UC, Helleberg C, et al. Early relapsed disease of multiple myeloma following up-front HDM-ASCT: a study based on the Danish Multiple Myeloma Registry in the period 2005 to 2014. *Leukemia* 2018;32:2054–7.
- Majithia N, Rajkumar SV, Lacy MQ, Buadi FK, Dispenzieri A, Gertz MA, et al. Early relapse following initial therapy for multiple myeloma predicts poor outcomes in the era of novel agents. *Leukemia* 2016;30:2208–13.
- Durie BGM, Jacobson J, Barlogie B, Crowley J. Magnitude of response with myeloma frontline therapy does not predict outcome: importance of time to progression in Southwest Oncology Group chemotherapy trials. *J Clin Oncol* 2004;22:1857–63.
- Kumar SK, Dispenzieri A, Fraser R, Mingwei F, Akpek G, Cornell R, et al. Early relapse after autologous hematopoietic cell transplantation remains a poor prognostic factor in multiple myeloma but outcomes have improved over time. *Leukemia* 2018;32:986–95.
- Gopalakrishnan S, D'Souza A, Scott E, Fraser R, Davila O, Shah N, et al. Revised international staging system is predictive and prognostic for early relapse (<24 months) after autologous transplantation for newly diagnosed multiple myeloma. *Biol Blood Marrow Transplant* 2019;25:683–8.
- Magarotto V, Bringhen S, Offidani M, Benevolo G, Patriarca F, Mina R, et al. Triplet vs doublet lenalidomide-containing regimens for the treatment of elderly patients with newly diagnosed multiple myeloma. *Blood* 2016;127:1102–8.
- Bringhen S, Petrucci MT, Larocca A, Conticello C, Rossi D, Magarotto V, et al. Carfilzomib, cyclophosphamide, and dexamethasone in patients with newly diagnosed multiple myeloma: a multicenter, phase 2 study. *Blood* 2014;124:63–9.
- Larocca A, Bringhen S, Petrucci MT, Oliva S, Falcone AP, Caravita T, et al. A phase 2 study of three low-dose intensity subcutaneous bortezomib regimens in elderly frail patients with untreated multiple myeloma. *Leukemia* 2016;30:1320–6.

19. Palumbo A, Bringhen S, Rossi D, Cavalli M, Larocca A, Ria R, et al. Bortezomib-melphalan-prednisone-thalidomide followed by maintenance with bortezomib-thalidomide compared with bortezomib-melphalan-prednisone for initial treatment of multiple myeloma: a randomized controlled trial. *J Clin Oncol* 2010;28:5101-9.
20. Gay F, Bertamini L, Zaccaria GM, Aquino S, Spadano A, Siniscalchi A, et al. Predictors of early relapse in young multiple myeloma patients: integrated analysis of baseline features and achievement of minimal residual disease negativity [abstract]. In: Proceedings of the 25th Congress of the European Hematology Association; 2020 Jun 11-Oct 15; Virtual. *Hemasphere* 2020;4:457. Abstract nr EP999.
21. Caers J, Garderet L, Kortüm KM, O'Dwyer ME, van de Donk NWCJ, Binder M, et al. European Myeloma Network recommendations on tools for the diagnosis and monitoring of multiple myeloma: what to use and when. *Haematologica* 2018;103:1772-84.
22. Sonneveld P, Avet-Loiseau H, Lonial S, Usmani S, Siegel D, Anderson KC, et al. Treatment of multiple myeloma with high-risk cytogenetics: a consensus of the International Myeloma Working Group. *Blood* 2016;127:2955-62.
23. Palumbo A, Avet-Loiseau H, Oliva S, Lokhorst HM, Goldschmidt H, Rosinol L, et al. Revised international staging system for multiple myeloma: a report from international myeloma working group. *J Clin Oncol* 2015;33:2863-9.
24. Therneau TM, Watson DA. The concordance statistic and the Cox model. Technical Report #85. Rochester, MN: Department of Health Sciences, Mayo Clinic; 2017. Available from: <https://www.mayo.edu/research/documents/bsi-techreport-85/doc-20433003>.
25. Collins GS, Reitsma JB, Altman DG, Moons KGM. Transparent reporting of a multivariable prediction model for individual prognosis or diagnosis (TRIPOD): the TRIPOD statement. *Ann Intern Med*. 2015;162:55-63.
26. Moons KGM, Altman DG, Reitsma JB, Ioannidis JPA, Macaskill P, Steyerberg EW, et al. Transparent reporting of a multivariable prediction model for individual prognosis or diagnosis (TRIPOD): explanation and elaboration. *Ann Intern Med* 2015;162:W1-73.
27. D'Agostino M, Zaccaria GM, Ziccheddu B, Rustad EH, Genuardi E, Capra A, et al. Early relapse risk in patients with newly diagnosed multiple myeloma characterized by next-generation sequencing. *Clin Cancer Res* 2020;26:4832-41.
28. Greipp PR, San-Miguel J, Durie BGM, Crowley JJ, Barlogie B, Bladé J, et al. International staging system for multiple myeloma. *J Clin Oncol* 2005;23:3412-20.
29. Dimopoulos MA, Barlogie B, Smith TL, Alexanian R. High serum lactate dehydrogenase level as a marker for drug resistance and short survival in multiple myeloma. *Ann Intern Med* 1991;115:931-5.
30. Rajkumar SV, Dimopoulos MA, Palumbo A, Blade J, Merlini G, Mateos M-V, et al. International Myeloma Working Group updated criteria for the diagnosis of multiple myeloma. *Lancet Oncol* 2014;15:e538-48.
31. Zaccaria GM, Ferrero S, Rosati S, Ghislieri M, Genuardi E, Evangelista A, et al. Applying data warehousing to a phase III clinical trial from the Fondazione Italiana Linfomi (FIL) ensures superior data quality and improved assessment of clinical outcomes. *JCO Clin Cancer Inform* 2019;3:1-15.
32. Schmidt TM, Barwick BG, Joseph N, Heffner LT, Hofmeister CC, Bernal L, et al. Gain of Chromosome 1q is associated with early progression in multiple myeloma patients treated with lenalidomide, bortezomib, and dexamethasone. *Blood Cancer J* 2019;9:94.
33. Bertamini L, Grasso M, D'Agostino M, Pascarella A, Tosi P, Monaco F, et al. Poor prognosis of multiple myeloma predicted by high levels of circulating plasma cells is independent from other high-risk features but is modulated by the achievement of minimal residual disease negativity [abstract]. In: Proceedings of the 62nd Annual Meeting of the American Society of Hematology; 2020 Dec 2-11; Virtual. *Blood* 2020;136:12-13. Abstract nr 720.