

Review

# Sarcopenia and Diabetes: A Detrimental Liaison of Advancing Age

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**Abstract:** Sarcopenia is an age-related clinical complaint characterized by the progressive deterioration of skeletal muscle mass and strength over time. Type 2 diabetes (T2D) is associated with faster and more severe skeletal muscle impairment. Both conditions influence each other, leading to negative consequences on glycemic control, cardiovascular risk, general health status, risk of falls, frailty, overall quality of life, and mortality. PubMed/Medline, Scopus, Web of Science, and Google Scholar were searched for research articles, scientific reports, observational studies, clinical trials, narrative and systematic reviews, and meta-analyses to review the evidence on the pathophysiology of diabetes-induced sarcopenia, its relevance in terms of glucose control and diabetes-related outcomes, and diagnostic and therapeutic challenges. This paper comprehensively addresses key elements for the clinical definition and diagnostic criteria of sarcopenia, instruments of assessment of skeletal muscle mass and strength, the pathophysiological correlation between T2D, sarcopenia, and related outcomes, a critical review of the role of antihyperglycemic treatment on skeletal muscle health, and perspectives on the role of specific treatment targeting myokine signaling pathways involved in glucose control and the regulation of skeletal muscle metabolism and trophism. Prompt diagnosis and adequate management, including lifestyle intervention, health diet programs, micronutrient supplementation, physical exercise, and pharmacological treatment, are needed to prevent or delay skeletal muscle deterioration in T2D.

**Keywords:** sarcopenia; diabetes mellitus; obesity; aging; physical exercise; protein supplementation; vitamin D; glucagon-like peptide 1; irisin; myostatin



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## 1. Introduction

Sarcopenia is defined as an age-related impairment of skeletal muscle performance, resulting in progressive deterioration of mobility, increased risk of falls and fractures, impaired ability to carry out daily activities [1]. According to the European Working Group on Sarcopenia in Older People, sarcopenia may occur with one or more three specific criteria: (a) low muscle strength, (b) low muscle quantity or quality, and (c) low physical performance [2]. Sarcopenia should be suspected if one criterion is satisfied. Two criteria confirm the diagnosis, and three define “severe sarcopenia”. Similarly, according to the

Third National Health and Nutrition Examination Survey, muscle mass should be assessed by bioimpedance analysis and be expressed as skeletal muscle index (skeletal muscle mass-to-body mass index  $\times 100$ ). Sarcopenia is defined when individual skeletal muscle index is lower than one standard deviation compared to reference values [3].

Type 2 diabetes mellitus (T2D) is a chronic multifactorial and systemic disease characterized by hyperglycemia and hyperglycemia-induced deterioration of microcirculation and macrovascular complications. The prevalence of T2D increases with age; hence, an overlap between T2D and sarcopenia is anticipated.

Sarcopenia is more prevalent in patients with chronic diseases, such as T2D, indicating that the age-related decline in skeletal muscle performance is faster than in healthy individuals [4,5]. Poor glucose control, longer diabetes evolution, and the presence of chronic diabetes-related complications also increase the risk of sarcopenia in T2D [6–9].

This review describes the pathophysiological relationship between T2D, sarcopenia, and related outcomes, a critical reexamination of the effect of pharmacological and non-pharmacological interventions in T2D on skeletal muscle health, and a perspective on the myokine signaling pathways involved in glucose control and skeletal muscle metabolism and trophism.

## 2. Methods

PubMed/Medline, Scopus, Web of Science, and Google Scholar were searched for research articles, scientific reports, observational studies, clinical trials, narrative and systematic reviews, and meta-analyses. Appropriate keywords or medical subject headings used in the research strings were as follows: “sarcopenia”; “diabetes mellitus/type 2 diabetes mellitus/T2D”; “aging”; “physical activity/physical exercise”; “protein supplementation”; “vitamin D”; “antihyperglycemic agent\*”; “biguanide”; “dipeptidyl peptidase type IV inhibitor\*/DPPIV inhibitor\*”; “sodium-glucose transporter type 2 inhibitor\*/SGLT2 inhibitor\*”; “glucagon-like peptide one receptor agonist\*/GLP-1RA”; “thiazolidinedione\*”; “insulin analog\*”; “acarbose”; “myokine,” “irisin”, “myostatin”.

## 3. Mechanism of Diabetes-Induced Sarcopenia

Evidence suggests that insulin resistance is associated with impaired skeletal muscle glucose uptake and utilization and intracellular accumulation of triglycerides/fatty acids, both associated with sarcopenia [10]. Lipid accumulation in myocytes further reduces skeletal muscle sensitivity to insulin [11]. Insulin resistance, hyperglycemia, and T2D per se induce mitochondrial dysfunction, impaired oxidative metabolism, and energetic utilization, contributing to sarcopenia [12]. In addition, insulin resistance impairs post-prandial myofibrillar protein synthesis due to an imbalance between catabolic and anabolic stimuli at the skeletal muscle site [13].

Proinflammatory cytokines, such as interleukin 1b (IL1b) and tumor necrosis factor  $\alpha$  (TNF $\alpha$ ), exacerbate protein imbalance by acting as catabolic stimuli [9]. Circulating levels of these cytokines are elevated in T2D and related co-morbidities, thus contributing to background systemic inflammation [14]. Resident macrophages are known to induce and sustain inflammation in the adipose tissue, pancreatic islets, liver, and other peripheral tissues [10], with a significant contribution to the pathophysiology of T2D. As another mechanism, proinflammatory macrophages promote lipolysis, thus exacerbating skeletal muscle steatosis and insulin resistance [12]. Advanced glycation end products (AGEs) also contribute to systemic inflammation and sarcopenia. The effect is attributable to the AGE-mediated activation of scavenger receptors (RAGEs), leading to the activation of proinflammatory pathways associated with systemic inflammation (NF- $\kappa$ B) and oxidative stress (NADPH oxidase) [15]. Hence, background systemic inflammation fosters sarcopenia in patients with T2D and related co-morbidities.

Gut dysbiosis plays a role in chronic intestinal and systemic diseases, including T2D. Notably, the balance between Bifidobacteria and Bacteroides is crucial in maintaining a healthy intestinal barrier. Children develop the best composition of the gut microbiome

when vaginally born and breastfed with maternal milk. Once solid foods are consumed daily, Bacteroides and Firmicutes become the prevalent species [16]. Subsequent changes in microbiome composition are related to genetic predisposition and diet. A hypercaloric Western diet predisposes one to T2D and cardiometabolic complications and is characterized by a significant shift in microbiome composition, expressed as a high Bacteroides to Bifidobacterial ratio. The mechanisms explaining the relation between gut dysbiosis and T2D are related to a significant change in intestinal mucosal membrane permeability that facilitates bacterial leakage and translocation from the gut lumen to the subepithelial space, eventually promoting endotoxemia, systemic inflammation, impaired insulin synthesis, and insulin resistance [17]. In addition, some specific species, such as *Akkermansia muciniphila*, are also less expressed in the gut of individuals with T2D, and this phenomenon is associated with impaired intestinal membrane permeability due to reduced synthesis of mucins, exacerbating bacterial leakage as mentioned above [18]. Moreover, gut dysbiosis is associated with defective synthesis of essential micronutrients, such as vitamin B12 and tryptophan, that play a crucial role in skeletal muscle homeostasis, with the latter phenomenon explaining the role of gut dysbiosis in T2D and sarcopenia [19].

Suboptimal chronic protein intake is an age-related nutritional concern. Several factors influence protein intake with advancing age, including physiological changes, such as reduced daily energy requirement, genetic predispositions to low appetite, dental issues, impaired gastric acid secretion and slow gastric emptying, pathological conditions, including physical and mental disabilities, inability to prepare or consume food, dysphagia, and environmental factors such as financial concerns or loneliness [20]. Moreover, these background conditions also affect bromatological diet composition in favor of carbohydrates (and rapidly adsorbed carbohydrates), thus increasing the risk of T2D, obesity, and sarcopenia.

Vitamin D (Vit-D) deficiency and insufficiency are frequently observed in old people. The leading causes of the age-related fall in Vit-D levels are attributable to low intake of naturally Vit-D-rich foods (e.g., meats, fish, eggs, milk, and milk-derived foods) and impaired dermal Vit-D metabolism. Vit-D deficiency is a usual finding in T2D. Vit-D deficiency contributes to impaired insulin synthesis and insulin resistance, increasing the risk of prediabetes and T2D [21]. Vit-D deficiency also contributes to sarcopenia, osteomalacia, osteoporosis, and the risk of falls and fractures [22].

Hormonal changes occur along with aging. The decline in the frequency and amplitude of growth hormone (GH) peaks and insulin-like growth factor (IGF) 1 is observed in old patients [23]. A similar imbalance is also known for testosterone in men and estrogen, progesterone, and ovarian- and adrenal-derived androgens in women [24,25]. T2D is frequently associated with male hypogonadism, with both conditions fostering sarcopenia in affected men [26]. Figure 1 depicts the pathogenesis of diabetes-related sarcopenia and the potential mechanisms to restore healthy skeletal muscle from sarcopenia.

T2D is associated with insulin resistance, chronic (low-grade) systemic inflammation, unhealthy lifestyle, malnutrition, and microbiome changes that represent concurrent factors of sarcopenia (indicated in bold red above the horizontal red arrow). Concurrent factors induce a significant perturbation in the physiological functions and biochemical activities of skeletal muscle (shown in red below the horizontal red arrow). A healthy lifestyle, including diet, protein and vitamin supplementation, regular physical exercise, and anabolic supplementation when necessary (indicated in bold green below the horizontal green arrow), attenuates skeletal muscle catabolism and may revert sarcopenia to healthy skeletal muscle (shown in green above the horizontal green arrow).



## 5. Sarcopenia: A Determinant of Glucose Deterioration and Poor Outcomes

Sarcopenia is associated with low glucose disposal at the skeletal muscle site [33]. Skeletal muscle is responsible for around 80% of glucose uptake during experimental conditions of euglycemic hyperinsulinemic clamp [34]. Skeletal muscle serves as a sort of buffer against hyperglycemia after a glucose load, as observed in the post-prandial phase under physiological conditions [35]. Preserving skeletal muscle mass prevents the onset of prediabetes and progression to T2D [34], as healthy insulin-sensitive skeletal muscle is essential to regulate glucose disposal. First, insulin stimulates the endothelial expression of nitric oxide synthase, nitric oxide production, and peripheral vasodilation. This mechanism ensures adequate blood flow and nutrient supply to skeletal muscle. Second, insulin stimulates the Akt/PKB-mediated translocation of glucose transporters, such as GLUT4, on myocyte membranes. Therefore, insulin is essential in increasing overall glucose uptake in skeletal muscle [36]. The third mechanism is insulin-independent and involves an extracellular matrix interposing the space between microvascular vessels and myocytes. In T2D, myocyte steatosis prompts insulin resistance, oxidative stress, cell injury, necrosis, and apoptosis. All these events stimulate the recruitment and translocation of peripheral monocyte/macrophage-derived proinflammatory cells into skeletal muscles, resulting in local inflammation, accumulation of cellular debris, and fibrillar amorphous matrix. The extracellular matrix becomes a thicker tissue, hindering glucose transport from vessels to myocytes. Inflammation, insulin resistance, and impaired regulation of intramuscular blood flow significantly affect glucose disposal by skeletal muscle [37].

Myokines are a group of proteins with autocrine, paracrine, and endocrine activities, and are produced and released by myocytes, whose expression increases with healthy skeletal muscle [38,39]. These molecules control muscle metabolism and growth, and have immunoregulatory effects [40]. Myokines can be classified as positive and negative regulators of muscle growth, differentiation, and repair. Bone-morphogenic proteins and irisin are the leading positive regulators, along with follistatin, which is secreted at the liver site. Myostatin, transforming growth factor  $\beta$ , activins, and growth differentiation factor are the foremost negative regulators [41,42]. A negative balance between myokines affects the differentiation, proliferation, and repair of myocytes and impairs myofibrillar synthesis, leading to sarcopenia [43]. Myokines, such as IL6, IL10, IL15, irisin, myonectin, osteocorin, and secreted proteins acidic and rich in cysteine (SPARC), are also involved in the crosstalk between skeletal muscle and peripheral tissues, such as pancreatic islets, the liver, adipose tissue, and in the regulation of insulin sensitivity, glucose metabolism, metabolite utilization, and energy expenditure [44–46].

Physical inactivity and a sedentary lifestyle are associated with insulin resistance, poor glucose control, and metabolically related consequences, including metabolic syndrome, T2D, obesity, and cardiovascular diseases [47,48]. A sedentary lifestyle is associated with loss of mechanical stimuli, consequent impairment of skeletal muscle trophism [49], skeletal muscle loss [50], and impaired myokine secretion. All these events are involved in impaired glucose metabolism and T2D pathophysiology. Overall, sarcopenia is an independent risk factor of new-onset T2D in normal-weight older people [51], as sarcopenic compared to non-sarcopenic patients require frequently a multipharmacological approach to manage chronic diseases [52] and related outcomes [53].

## 6. Modifiable Risk Factors

Aging is the foremost non-modifiable risk factor for sarcopenia and T2D. However, it is not the only risk factor associated with skeletal muscle health deterioration. Physical exercise, education about a healthy lifestyle, and social status are important and potentially modifiable risk factors of both sarcopenia [54–56] and T2D [57]. Adequate management of modifiable risk factors has positive consequences in the prevention and treatment of sarcopenia and T2D in the general population. Nonetheless, managing these non-modifiable risk factors in a community-based manner takes work, especially considering that current healthcare policies usually focus on individually centered management of patients. In



addition, healthcare policies do not routinely supply healthcare facilities with sufficient time, space, adequate specialists, and appropriate technological support to improve the quality and efficacy of prescriptions for lifestyle changes, especially exercise programs [58]. Consequently, most patients usually receive less specific advice without supervision on lifestyle adjustments. This approach is characterized by a considerable heterogeneity in the results because of patients' background differences, such as financial resources and the possibility of adequate access to care. Keeping in mind the current limitations and given the importance of structured and supervised lifestyle change interventions, health policies should endorse more education and specific psychosocial and financial support to facilitate adherence to interventions for most.

## 7. Preventing Sarcopenia: A Therapeutic Target in Primary and Secondary Prevention of T2D

### 7.1. The Physiological Role of Healthy Skeletal Muscle in Preventing Sarcopenia and Glucose Metabolism Deterioration

Preserving myokine syntheses, such as irisin, IL6, myonectin, decorin, fibroblast growth factor (FGF) 19, IL15, SPARC, and brain-derived neurotrophic factor or BDNF, results in a significant improvement in mitochondrial function and skeletal muscle metabolism, myofibrillar synthesis and skeletal muscle growth, insulin secretion, peripheral glucose and lipid utilization, with overall improvement in body composition (including fat mass loss) [59–61]. Moreover, suppressing the synthesis of myostatin, a potent skeletal muscle-derived transforming factor that acts as an endogenous inhibitor of myofibrillar synthesis and muscle growth, or increasing the hepatic synthesis of follistatin, a potent endogen myostatin inhibitor, can be additional strategies to improve muscle trophism. High circulating myostatin levels are observed in sarcopenic patients, in whom myostatin concentration is inversely related to follistatin, GH, IGF 1, testosterone, and estradiol [62]. Physical exercise, directly or through indirect metabolic changes, such as low insulin-to-glucagon ratio, GH, and IGF1, enhances the synthesis of myokines and promotes the liver-mediated secretion of follistatin with a net effect on skeletal muscle gain [63,64].

Satellite cells are muscle-derived stem cells that play an essential role in skeletal muscle repair and regeneration [65]. Preserving satellite cells would result in an antiaging effect, an important ally against sarcopenia.

Suppressing the adenosine monophosphate-activate protein kinase (AMPK)–mammalian target of rapamycin (mTOR) pathway by the TGF- $\beta$ -small mother against decapentaplegic (Smad) signaling results in impaired muscle synthesis and muscle atrophy [66,67]. Therefore, reinforcing the mTOR pathway is beneficial for skeletal muscle health.

Testosterone, estradiol, and GH and IGF1 provide essential anabolic stimuli to increase skeletal muscle mass and reverse skeletal muscle impairment [68]. As the efficiency of the hypothalamus–pituitary–gonadal and GH-IGF-1 axes declines considerably over time, skeletal muscle trophism is significantly impaired by aging. Anabolic hormones are dampened in T2D, especially functional male hypogonadism, with both conditions considered significant contributors to sarcopenia [24].

### 7.2. Non-Pharmacological Intervention: The Role of Lifestyle Changes and Supplements

Evidence suggests that adequate protein intake, monounsaturated acid supplementation, and anti-inflammatory diets have a therapeutical potential to improve muscle health and prevent sarcopenia [69–73] (Table 1). Ramified aminoacidic supplementation attenuates skeletal muscle catabolism and induces skeletal muscle mass gain when combined with regular training, especially resistance training [74,75]. Generally, dietary intervention and physical exercise improve body composition and skeletal muscle strength at all ages [76,77].

**Table 1.** Summary of the leading mechanisms by which non-pharmacologic intervention may affect skeletal muscle health in T2D.

Type of Intervention	Possible Positive Effects on Skeletal Muscle	Possible Detrimental Effects on Skeletal Muscle	Overall Effect
Protein supplementation	Attenuates myofibrillar catabolism	-	Prevent sarcopenia
Vitamin D supplementation	Improve insulin sensitivity Boost testosterone synthesis Boost myokine synthesis (e.g., irisin)	-	Prevent sarcopenia
Diets	Insulin-sensitizing effect Improve glucose utilization Reduce systemic inflammation Prevent muscle steatosis Induce weight loss (Facilitate adherence and resistance to physical exercise)	Impairment of testosterone synthesis (low-fat diets, intermittent fasting protocols, vegetable-based diets)	Prevent sarcopenia
Physical exercise (high-intensity more than low-to-moderate intensity)	Insulin-sensitizing effect Improve glucose utilization Prevent muscle steatosis Induce weight loss Boost testosterone synthesis Boost myokine synthesis Increase myofibrillar synthesis Reduce myofibrillar catabolism	-	Improve muscle mass and strength

Most diet protocols have been demonstrated to affect testosterone synthesis in men. Intermittent fasting protocols are associated with serum total testosterone decline. A reduction in serum testosterone is usually not associated with short-term lean and skeletal muscle mass and loss of strength [78], even though more study is needed to clarify the long-term effects of such protocols. Low-fat diets are associated with significant weight loss and improvement in insulin sensitivity but are also associated with a considerable decline in testosterone concentration with potentially adverse effects on lean mass and body composition [79]. A mild but significant reduction in serum testosterone has also been observed with the Mediterranean diet [80]. Conversely, a low-carb diet with moderate-to-high protein intake not exceeding 3.4 g/kg/day is usually associated with a neutral or even ameliorating effect on serum testosterone [81–83]. Very low-carb diets induce a significant increase in serum testosterone, even if the magnitude of this effect is strictly associated with weight loss and the patient's age [84].

Estradiol is also essential for skeletal muscle health [85]. Phytoestrogens, polyphenols, and hormonal replacement therapy can be considered in post-menopausal women to reinforce muscle health [86,87].

Vit-D is essential for skeletal muscle health [88]. Low circulating levels of 25OH-Vit-D were found in sarcopenic compared to healthy individuals [89]. Vit-D supplementation is associated with gain in muscle strength and, possibly, skeletal muscle mass in healthy and sarcopenic people [90–92]. Combining resistance exercise with adequate protein intake and Vit-D supplementation ensures better results on skeletal muscle performance in sarcopenic people [93,94]. Vit-D supplementation is also proven to boost testosterone synthesis in men. Vit-D receptors were found in testicular tissue, especially Leydig cells, where the vitamin is locally activated [95] and stimulates the synthesis of testosterone [96,97]. Men with Vit-D deficiency and insufficiency display reduced levels of serum testosterone and lower testosterone-to-luteinizing hormone ratio, indicating that sufficient exposure to vitamin D is required to sustain the testicular synthesis of testosterone [98]. Vit-D deficiency and male hypogonadism usually coexist and are both independent risk factors for frailty [99].

On the other hand, testosterone affects the peripheral metabolism of Vit-D by enhancing the synthesis of 1,25OH-Vit-D, such as in the kidneys, intestine, and bone tissue [100,101]. This mechanism is probably an additional contributor to the pathogenesis of osteopenia/osteoporosis in hypogonadal men [95]. Compared to standard supplementation of VIT-D (800–1000 IU/day), high-dose Vit-D (>3000 IU/day per 1 year) increases circulating levels of testosterone in healthy individuals [102,103]. Primitive testicular damage is associated with impaired testosterone synthesis and 1,25OH-hydroxylase activity [104]; therefore, sufficient levels of active Vit-D (calcitriol) are required to stimulate testosterone synthesis [105].

7.3. Pharmacological Intervention

7.3.1. Biguanides

Metformin, a biguanide approved for T2D, is widely used as a first-line treatment of T2D [106]. Metformin induces controversial results on body composition, skeletal muscle health, and strength. On the one hand, metformin suppresses hepatic glucose output and improves glucose metabolism in skeletal muscle, consequently ameliorating energy utilization and preventing muscle steatosis, two key biochemical elements to prevent or treat sarcopenia [107]. Metformin also exhibits anti-inflammatory and antioxidative properties, improves satellite cell viability and regenerative effects, and promotes myofibrillar repair [108–110]. The mechanism by which metformin improves muscle repair could be attributable to the drug-induced attenuation of Smad 2 and 3 activities in the context of the TGF-β signaling pathway [111], as the attenuation of this pathway stimulates insulin secretion and myofibrillar synthesis [112,113]. On the other hand, metformin affects the AMPK/mTORc1 pathway, thus reducing glucose output from the liver and fasting glucose levels. The inhibition of mTORc1 is also associated with impaired protein synthesis and autophagy, which results in defective myofibrillar synthesis and skeletal muscle hypotrophy [114]. Moreover, metformin activates the forkhead box O3a or FoxO3a, via AMPK, a key transcription factor of myostatin, a potent inhibitor of myofibrillar synthesis and skeletal muscle growth [115]. Observational data suggest that metformin may have a protective effect against sarcopenia [116–118]. However, the level of evidence is slight and possibly affected by confounding factors, such as exercise, diets, or nutraceuticals (Table 2).

**Table 2.** Summary of the leading mechanisms by which pharmacologic intervention affects skeletal muscle health in T2D.

Classes of Antihyperglycemic Agents	Possible Positive Effects on Skeletal Muscle	Possible Detrimental Effects on Skeletal Muscle	Overall Effect
Biguanides (e.g., metformin)	Insulin-like sensitizing effect Improve glucose metabolism Ameliorate energy utilization Anti-inflammatory/antioxidative properties Improve satellite cell viability/regenerative effects Antiproteolytic effect (Inhibition of TGF-β/Smad signaling)	Proteolytic effect (Inhibition of AMPK/mTORc1 pathway) Stimulate myostatin synthesis (AMPK/FoxO3a transcription factor)	Neutral or favors sarcopenia
Secretagogues (e.g., sulfonylureas, glinides)	Unclear	Inhibit ATP-sensitive potassium channels (Muscle atrophy) Enhance caspase-3 activity (Apoptosis)	Favors sarcopenia



Table 2. Cont.

Classes of Antihyperglycemic Agents	Possible Positive Effects on Skeletal Muscle	Possible Detrimental Effects on Skeletal Muscle	Overall Effect
Thiazolidinediones (e.g., pioglitazone)	Insulin-sensitizing effect Improve glucose utilization Prevent muscle steatosis	Direct muscle toxicity? (Rhabdomyolysis, rare adverse event)	Neutral
Intestinal glucosidase inhibitors (e.g., acarbose)	Unclear	Unclear	Unclear
DPPIVIs	Potentiate microvascular supply Insulin-sensitizing effect Improve glucose utilization Antioxidative/anti-inflammatory effects Enhance the synthesis of PGC-1 $\alpha$ (Mitochondrial biogenesis)	Unclear	Neutral
SGLT2is	Metabolic shift toward fatty acids and ketones Improve tissue oxygenation Antioxidative/anti-inflammatory effects Improve cardiac pump efficiency Improve exercise tolerance Boost myokine secretion	Clinical evidence of fat-free mass loss	Favors sarcopenia. Prevent sarcopenia in heart failure
GLP-1RAs	Improve glucose utilization Antioxidative/anti-inflammatory effects Stimulate hepatic synthesis of IGF1 (myogenesis) Boost myokine secretion Improve satellite cell viability Improve satellite cell viability/regenerative effects Promote myofiber repair Boost testosterone synthesis	Excessive weight loss Reduce appetite (might hamper sufficient caloric and protein intake)	Prevent sarcopenia or improve skeletal muscle
Insulin analogues	Improve glucose utilization Antioxidative/anti-inflammatory effects Potentiate microvascular supply Direct stimulation of myofibrillar synthesis Direct inhibition of myofiber proteolysis	Long-term, dose-dependent impairment of insulin sensitivity Muscle steatosis Weight gain and hypoglycemia (Facilitate discontinuation of physical exercise and sedentarism)	Unclear

Abbreviations: transforming growth factor  $\beta$  = TGF- $\beta$ ; small mother against decapentaplegic = Smad; adenosine monophosphate-activate protein kinase = AMPK; mammalian target of rapamycin = mTOR; forkhead box O3 = FoxO3; adenosine triphosphate = ATP; peroxisome proliferator co-activator 1 alpha = PGC-1 $\alpha$ ; insulin-like growth factor 1 = IGF1; dipeptidyl peptidase type IV inhibitors = DPPIVIs; glucagon-like peptide 1 receptor agonists = GLP-1RAs; sodium-glucose (co)transporter type 2 inhibitors = SGLT2is.

### 7.3.2. Secretagogues

Sulfonylureas have been widely used for treating hyperglycemia in T2D. The drugs bind to a specific site of the ATP-sensitive K-channel in the  $\beta$ -cell plasma membrane and close it, consequently blocking the potassium outflow, depolarizing the cell membrane, and initiating the signal cascade, eventually resulting in insulin release [119]. Sulfonylureas potentiate glucose disposal in skeletal muscles [120]. Nevertheless, preclinical studies have found that inhibiting ATP-sensitive potassium channels is associated with muscle atrophy. In addition, sulfonylureas may enhance caspase-3 activity and reduce the protein content

in skeletal muscle [121]. Overall, these data indicate that sulfonylureas have detrimental effect on skeletal muscle health.

### 7.3.3. Intestinal Glucosidase Inhibitor

Acarbose, an  $\alpha$ -glucosidase inhibitor, is an antihyperglycemic agent able to attenuate post-prandial glycemic excursion after food intake [122]. Data suggest that acarbose could be associated with impaired muscle trophism and strength, with unclear mechanisms [123]. Caution should be taken while prescribing  $\alpha$ -glucosidase inhibitors in patients at risk for or diagnosed with sarcopenia [124].

### 7.3.4. Dipeptidyl Peptidase Type IV Inhibitors

Dipeptidyl peptidase type IV inhibitors (DPPiVIs) belong to the incretin family, a class of drugs affecting the incretin system. DPPiVIs compete with endogen incretins, such as the glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide 1 (GLP-1), to the catalytic site of the enzyme and inhibit their degradation [125]. Therefore, DPPiVIs improve glucose control by extending the circulating half-life of endogenous incretins, especially in the post-prandial phase [126]. Besides their well-established effect on glucose control in terms of absolute efficacy and durability [127], DPPiVIs are associated with a neutral effect on body weight and composition [128]. However, mechanistic data suggest that DPPiVIs may sustain skeletal muscle trophism. First, they have the potential to regulate the arteriolar diameter and, consequently, increase blood flow in skeletal muscle [129]. Second, they enhance the insulin-mediated translocation of glucose transporters on the myocyte surface [130], have antioxidative and anti-inflammatory effects, and improve mitochondrial function and oxidative phosphorylation [131]. The latter effect could be mediated by DPPiV-induced sympathetic activation rather than a direct effect, as observed in the post-prandial phase in T2D individuals [132]. DPPiVIs improve exercise tolerance in patients with heart failure by stimulating mitochondrial biogenesis in skeletal muscles [133,134]. The peroxisome proliferator co-activator 1 alpha (PGC-1 $\alpha$ ) plays a direct role in mitochondrial biogenesis and mitophagy, two fundamental biological events related to mitochondrial viability and function [135]. Notably, PGC-1 $\alpha$  is downregulated in the skeletal muscle of patients with T2D and DPPiVIs, such as sitagliptin, stimulate the PGC-1 $\alpha$  synthesis [136] with protective effects against insulin resistance, skeletal muscle hypotrophy, and impaired glucose and lipid metabolism [137]. Overall, clinical data indicate that DPPiVIs do not improve skeletal muscle mass or cardiometabolic fitness, with an uncertain effect on sarcopenia [138].

### 7.3.5. Thiazolidinediones

Thiazolidinediones are insulin-sensitizer agents as they increase significantly glucose uptake in skeletal muscle at rest and after exercise [139]. Specifically, thiazolidinediones increase the phosphorylation of protein kinase B and insulin-stimulated phosphoinositide 3-kinase activity in skeletal muscle [140,141]. This insulin-sensitizing effect is also mediated by an increase in serum adiponectin concentration and adiponectin receptor 1 in skeletal muscle and adipose tissue, which is associated with enhanced glucose uptake [142,143]. Moreover, thiazolidinediones are well-known peroxisome proliferator-activated receptors  $\gamma$  which regulate adipocyte differentiation, fatty acid storage, and glucose metabolism [144]. Thiazolidinediones affect lipid metabolism by increasing HDL, decreasing LDL and triglycerides, and attenuating liver steatosis, which is associated with sarcopenia [145,146]. As demonstrated in mouse models, the fructose transporter GLUT5 is markedly enhanced in skeletal muscle. It is a possible adaptive mechanism to overcome an impaired glucose metabolism, but it predisposes to steatosis [147]. Rosiglitazone reduces the expression of GLUT5, hence playing a protective role in both the liver and skeletal muscle. Pioglitazone has anti-inflammatory properties in skeletal muscle [148] and improves mitochondrial function in T2D [149]. Moreover, thiazolidinediones reduce circulating and intramuscular levels of ceramides [150], which play a pathophysiological role in insulin resistance in

skeletal muscle and accentuate the risk of age-related sarcopenia in T2D [151]. Clinical data indicate that pioglitazone increases whole-body aerobic capacity and skeletal muscle energy metabolism, thus providing beneficial effects on muscle trophism and physical performance [152]. Because of the adipogenic potential of thiazolidinediones, fat mass gain at the subcutaneous but not visceral adipose tissue site [153,154] is a common finding in T2D. Moreover, pioglitazone stimulates the commitment of skeletal muscle satellite cells to adipocytes [155]. Nevertheless, these phenomena are not associated with skeletal muscle impairment [156]. Thiazolidinediones significantly increase total body water content, as they stimulate sodium retention. However, they did not affect the level of skeletal muscle hydration [157]. A decline in bone mineral density has also been reported in patients on thiazolidinediones [158]. A few cases in the literature of thiazolidinedione-induced rhabdomyolysis have been described. The mechanisms are unclear, also considering that neither statins nor physical exercise were ascertained as promoting factors [159]. Besides sporadic cases, thiazolidinediones should be considered safe and possibly effective medications for preserving muscle health in T2D.

### 7.3.6. Gliflozins

Gliflozins or sodium-glucose (co)transporter type 2 inhibitors (SGLT2is) act as antihyperglycemic agents by blunting glucose resorption at the proximal renal tubule site through an insulin-independent mechanism [160]. SGLT2is reduce glucose resorption by 30 to 50% [161] and are thus responsible for a moderate but significant caloric dissipation of approximately 200 Kcal/day [162]. Glycosuria is also responsible for osmotic diuresis, which leads to transient extracellular water and sodium depletion [163]. SGLT2is are associated with broad positive effects beyond glucose control, including a mild reduction in blood pressure and weight loss and cardiovascular and renal benefits. Moreover, SGLT2is have antioxidative effects, improve mitochondrial function, provide a favorable metabolic shift towards fatty acids and ketone bodies rather than glucose in myocytes, stimulate erythropoiesis, attenuate the sympathetic tone [164], insulin resistance, and systemic inflammation [165]. These effects ameliorate cardiac pump efficiency and improve long-term outcomes related to chronic heart failure, regardless of hyperglycemia and background cardiac pump efficiency [166]. SGLT2is affect body composition by specifically reducing both fat and fat-free mass [167]. Despite causing a significant reduction in fat mass in both subcutaneous and visceral adipose areas, these medications have had some controversial results [168], including increasing the long-term loss of lean mass and skeletal muscle mass [169]. Because of this phenomenon, physical exercise, diet, and an adequate protein intake are needed to minimize muscle mass impairment in T2D patients on SGLT2is [170]. On the other hand, SGLT2is have been demonstrated to improve skeletal muscle performance in patients with T2D and heart failure. Low cardiac output, tissue hypoxia, hormonal and metabolic imbalance, and forced inactivity or immobilization are the leading mechanisms explaining skeletal muscle deterioration in T2D with heart failure [171]. As SGLT2is improve cardiac function, tissue perfusion, and oxygenation and provide a metabolic boost to skeletal muscle mass metabolism [172], this class of medications may be helpful in preserving skeletal muscle mass and performance in T2D with chronic heart failure [173].

### 7.3.7. Glucagon-like Peptide 1 Receptor Agonists

Glucagon-like peptide 1 receptor agonists (GLP-1RAs) belong to the incretin class approved for T2D. Apart from semaglutide, which includes injectable and oral formulations, GLP-1RAs are administered subcutaneously with simple-to-use devices. Clinical trials showed that GLP-1RAs, especially long-acting analogs, are potent antihyperglycemic agents and induce significant weight loss without affecting the risk of hypoglycemia [174]. GLP-1RAs work in several domains. The leading therapeutic effects of GLP-1RAs include the enhancement of glucose-dependent insulin release and suppression of glucagon secretion during hyperglycemia, but not hypoglycemia, delay of gastric emptying, and sup-

pression of appetite [175,176]. GLP-1RAs also provide cardiovascular and renal protection. As GLP-1RAs work efficaciously and safely, the prescription of this class of medications has gained success in T2D, even compared to other equally potent but more expensive and composite regimens, such as insulin therapy [177–179]. GLP-1RAs are expected to affect body composition significantly. Patients on GLP-1RAs lose weight, fat mass, and visceral adipose tissue instead of fat-free and skeletal muscle masses [180–182]. Real-life studies have confirmed previous findings in T2D patients with and without weight excess up to one year of treatment [183–185]. Interestingly, GLP-1RAs improve endothelial function in T2D with indirect (improving glucose control and insulin signaling) and direct (receptor agonism) mechanisms, thus improving muscle perfusion and angiogenesis [186]. Some uncertainty remains on the role of GLP-1RAs in modulating skeletal muscle viability and metabolism, since previous evidence did not confirm the presence of GLP-1 receptors on skeletal muscle myocytes. One study found that semaglutide inhibited ubiquitin-proteasome-mediated skeletal muscle proteolysis, thus promoting myogenesis in murine myocytes. These effects were attributable to a GLP-1RA-induced decrease in proinflammatory cytokines and oxidative stress, which in turn was associated with the attenuation of ubiquitin-proteasome muscle wasting and increase in the hepatic synthesis of IGF 1 (myogenesis). Nevertheless, semaglutide was found to improve skeletal muscle atrophy by directly stimulating GLP-1 receptors in myocytes by the cAMP-mediated activation of PKA and AKT [187]. Another study found that liraglutide and semaglutide improved glucose tolerance and insulin sensitivity, reduced body weight gain and excessive lipid accumulation, and enhanced muscle atrophy in a high-fat diet model of obesity by activating the SIRT1 pathway [188]. Evidence suggests that GLP-1RAs boost irisin release and reduce IL6 secretion after 6 months of treatment, indicating favorable effects on skeletal muscle and adipose tissue, as both irisin deficiency and chronic exposure to moderate-high levels of IL6 are associated with insulin resistance, impaired insulin secretion, poor glucose control, weight gain, expansion of visceral adipose tissue, and muscle hypotrophy [189,190]. GLP-1RAs also attenuate the expression of atrophic factors in mice, thus decreasing the skeletal muscle catabolism associated with advancing age and T2D [191] and stimulating the expression of antiatrophic factors and differentiation of satellite stem cells to improve skeletal muscle regenerative potential [192]. In addition, GLP-1RAs boost the hypothalamus–pituitary–testicle axis and increase serum testosterone concentration in T2D and functional hypogonadism, positively affecting weight loss and body composition [193,194].

#### 7.3.8. Dual GLP-1/GIP Co-Agonists

Dual GLP-1/GIP co-agonists have recently been approved for T2D [195], and GLP-1/glucagon co-agonists and triple (glucagon, GLP-1, and GIP) agonists are under investigation [196,197]. It has been demonstrated that co-agonists provide a synergic effect on appetite and energy intake compared to GLP-1RAs alone [198]. Clinical trials indicate that dual GLP-1/GIP co-agonists, such as tirzepatide, should be considered the most effective agents compared to other antihyperglycemic drugs in the early stage as well as long-lasting T2D [199–204]. Clinical trials and meta-analyses also indicated that tirzepatide, compared to placebo, reduces body weight by 7.5 to 12 kg (5 to 15 mg/weekly) and 1.7 to 7.2 kg compared to GLP-1RAs [205,206]. Ongoing investigations confirm these impressive results on weight loss also in obese individuals. In the SURMOUNT-1 trial, the mean percentage change in weight after 72 weeks of treatment was −15% with 5 mg weekly doses of tirzepatide, −19.5% with 10 mg doses, and −20.9% with 15 mg doses compared to −3.1% with placebo. The number of patients who lost more than 20% of baseline body weight while on 10 mg (50%) and 15 mg doses (57%) was significantly higher compared to placebo (3%) [207]. Slightly lower but significant results were also found in obese individuals with T2D, as demonstrated by the SURMOUNT-2 trial [208]. Tirzepatide in addition to a lifestyle intervention also boosted weight loss in patients who had achieved a satisfactory weight reduction (i.e.,  $\geq 5.0\%$ ) after a 12-week intensive lifestyle intervention [209]. Overall, the promising results of dual agonists are approaching those obtained with bariatric

surgery [210,211], in a magnitude never seen until today [212]. Despite remarkable weight loss, tirzepatide was found to selectively reduce fat, but not free-fat mass, as recently demonstrated [213,214]. Thanks to these results, tirzepatide is expected to prevent and dramatically change the clinical course of cardio-nephron-metabolic diseases [215], but specific trials are needed to ascertain the magnitude of positive effects on skeletal muscle health and strength.

### 7.3.9. Insulin Analogues

Insulin promotes muscle growth by stimulating myofibrillar synthesis and increasing skeletal muscle blood flow, amino acid delivery, and availability [216]. The leading mechanism by which insulin promotes skeletal muscle protein synthesis and muscle growth is attributable to the activation of the phosphatidylinositol 3-kinase–mTOR pathway, in an opposite way compared to how metformin works [217]. Insulin analogs act exactly like human insulin to stimulate glucose and amino acid uptake in skeletal muscle myocytes [218]. Also, insulin is essential to the promotion of glucose utilization, oxidative mitochondrial respiration, and substrate accumulation after energy replacement in skeletal muscles in a dose-dependent manner [219]. Experimental models suggest that insulin therapy reduces myocyte apoptosis and attenuates skeletal muscle wasting in rats by alleviating reticulum endoplasmic stress [220]. However, long-term insulin treatment was found to produce significant histological changes in skeletal muscle myocytes, including the level of expression of myosin heavy chains, shift toward type II fibers, and reduced expression of several myokines, such as IL6, myostatin, and irisin [221]. Despite the potential for improving skeletal muscle health, these changes are involved in insulin resistance and the deterioration of skeletal muscle performance, which depends on chronic exposure to endogen insulin or exogen analogs and occurs in a dose-dependent manner. Insulin treatment must be adjusted over time, as most of its hypoglycemic potential is obtained when a treat-to-target approach is carried out. The need for intensifying insulin regimens depends, at least in part, on a progressive decline in insulin sensitivity, which, in turn, is related to several factors, including obesity and chronic insulin exposure per se. Progressive titration of insulin analogs leads to a considerable increase in the total daily dose of insulin, which results in weight gain, risk of hypoglycemic events, and further deterioration of skeletal muscle mass and insulin sensitivity. Strategies to improve insulin sensitivity are therefore necessary to attenuate this vicious circle [222] and preserve skeletal muscle health.

Given the pathophysiology of T2D, most patients receive combined treatment in their lifetime to achieve tailored control of glycemic and extra-glycemic targets. Metformin is the background treatment of T2D; adding DPP-IVis, thiazolidinediones, and SGLT2is is expected to improve skeletal muscle health by balancing the positive and potentially detrimental effects of each class of drugs individually. Adding GLP-1RAs or more composite regimens, such as basal insulin or basal-GLP-1RAs, may induce interesting results as well [223].

## 8. Future Directions

Tailored interventions are the new frontiers of precision- and evidence-based medicine. Targeting energy expenditure, fat oxidation, appetite regulation, and lean mass preservation are key elements for sustainable weight loss in metabolic disorders, including T2D [224]. The pharmacological treatment of T2D induces different effects on skeletal muscle health. Metformin, acarbose, and secretagogues do not improve body composition and skeletal muscle mass, as they have neutral or even detrimental effects on muscle health. Thanks to a wide range of metabolic and non-metabolic effects, thiazolidinediones and DPP-IVis have the potential for attenuating age-related skeletal muscle mass decline in T2D and, possibly, in earlier stages of impaired glucose metabolism. However, they have not been demonstrated to increase skeletal muscle mass or strength. Moreover, DPP-IVis do not affect body weight and composition, while thiazolidinediones induce weight and fat mass gain over time. SGLT2is have been demonstrated to improve body composition, espe-



cially by providing mild-to-moderate weight loss more than other oral antihyperglycemic agents. Nevertheless, some data suggest that gliflozins might impair muscle mass and strength, leading clinicians to consider appropriate lifestyle adjustments or avoid SGLT2is prescription in patients at risk of or with sarcopenia [225]. However, these agents improve skeletal muscle health, exercise tolerance, and overall physical performance in patients T2D with heart failure. GLP-1RAs have the potential to affect body composition healthier. They reduce body weight, fat mass, and visceral adipose tissue while preserving or even improving skeletal muscle mass and strength regardless of baseline body composition and BMI. Nevertheless, GLP-1RAs significantly reduce food intake and appetite, and their use in sarcopenic patients may be complicated by further weight loss and the inability to consume hypercaloric diets and protein supplementations. Prescribing GLP-1RAs may be, therefore, intricate in sarcopenic patients or those at high risk of sarcopenia. Insulin therapy can potentially improve skeletal muscle mass and induce weight gain. However, it is less likely to be handled easily compared to non-insulin regimens, raises hypoglycemic risk, and requires adequate daily glucose monitoring and proper adherence by patients or caregivers.

Future research is needed to address more evidence on the management of sarcopenia in T2D. Aside from biomechanical function, skeletal muscle has significant endocrine activities. Preserving skeletal muscle endocrine functions means maintaining essential crosstalk between skeletal muscle and several tissues, such as the brain, adipose tissue, bones, the liver, gut microbiome, pancreatic islets, microvasculature, skin, and muscle itself [226]. A few studies have been conducted in the long term, highlighting non-significant or controversial results on skeletal muscle mass and strength [227]. Among antihyperglycemic agents, SGLT2is were found to preserve healthy myokine secretion, which is essential to the maintenance of both metabolic and functional activity of skeletal muscles. Nevertheless, they probably have neutral or detrimental effects on atrophic factors, such as myostatin. As indicated in preclinical studies, GLP-1RAs may induce anabolic stimuli in skeletal muscles and potentiate muscle-regenerative properties. Evidence suggests that combining physical exercise with diet ensures more significant effects on skeletal muscle health. Although similar positive results can be anticipated while considering the combination of physical activity and antihyperglycemic treatments, no specific studies have been carried out to confirm or deny this hypothesis.

Myokine-based therapy has the potential to be a new therapeutic frontier. It is widely accepted that physical exercise has antidegenerative and renewal properties, and myokines are the leading mediators of these beneficial effects. Irisin is primarily involved in maintaining muscle cell and bone health thanks to its anti-inflammatory [228] and regenerative properties [229]. Mechanistic studies have shown a close association between irisin deficiency and the development of insulin resistance and cardiometabolic complications, such as pathological myocardial remodeling [230]. Aerobic, high-intensity interval training and combined aerobic–resistance workouts increase irisin levels considerably in different settings and regardless of background characteristics in terms of cardiorespiratory fitness and weight status [231,232]. Caloric restriction alone seems not to preserve lean mass and slightly reduces irisin concentration [233]. Interestingly, Vit-D supplementation increases the level of circulating irisin by directly stimulating the intramuscular synthesis of its precursor (fibronectin type III domain-containing protein 5 or FNDC5) [234], which probably mediates both metabolic and functional parameters [235,236]. There is a direct relationship between serum testosterone concentration and irisin levels in men with metabolic syndrome [237,238]. Other data show that insulin resistance is associated with increased basal but not post-exercise levels of irisin [239,240], indicating that the condition of irisin resistance may play a role in the early stages of T2D. Testosterone replacement treatment in male functional hypogonadism, including patients with T2D, is associated with a significant increase in irisin concentration [241]. Conversely, peripheral but not intracerebral irisin administration improve testosterone levels and biologically related consequences such as sexual function and spermatogenesis [242–244].

Fibroblast growth factor 19 (FGF19) has significant regenerative, metabolic, and anti-inflammatory properties associated with skeletal muscle growth and hypertrophy [245–247], while low levels of FGF19 are associated with sarcopenia [248]. FGF19 reverts obesity-induced muscle atrophy and restores irisin levels [249], thus playing a role in improving skeletal muscle health. One study found that short-term administration of FGF19 improved skeletal muscle growth regardless of food intake in mice [250].

Last, myostatin inhibitors and follistatin analogs can improve skeletal muscle mass and strength. Interesting results can be obtained by monoclonal antibodies targeting the myostatin/activin signaling pathway by antagonizing activin type II receptors, which mediate muscle breakdown [251]. Activin type II receptor antagonism is expected to maximize muscle hypertrophy in the presence of chronic muscle training [252]. Moreover, testosterone, estradiol, and GH suppress myostatin synthesis by downregulating gene expression and stimulate the synthesis of follistatin [253,254]. This mechanism is thought to explain, at least in part, the anabolic effect of sexual steroids and GH on skeletal muscle trophism. Although aging is associated with a progressive decline in gonadal and hypophyseal function, no evidence indicates that replacing dysfunctional axes may result in a long-term and safe amelioration of skeletal and muscle endpoints, including muscle strength, prevention of falls, and frailty [24]. Therefore, targeting myostatin with specific monoclonal antibodies may have a therapeutic rationale, as demonstrated by several trials in patients with primitive or secondary myopathies [255,256] and age-related sarcopenia [257].

## 9. Conclusions

The close interconnection between sarcopenia and T2D is well known. Both conditions are expected to increase in prevalence due to the elongation of life expectancy, as aging is one of the leading contributing factors of T2D and sarcopenia.

Identifying patients at risk of or with sarcopenia is essential for individualizing comprehensive therapeutic programs in T2D, including education, lifestyle adjustments, healthy diet, micronutrients, and protein supplements, regular physical exercise, and appropriate pharmacological treatment to remove risk factors, revert skeletal muscle depletion and, possibly, improve skeletal muscle mass and strength.

Preserving skeletal muscle mass and strength positively affects overall physical performance and independence during daily activities. It also holds important endocrine and metabolic mechanisms underlying significant improvements in glucose control, durability of pharmacological effectiveness, prevention of complications, and amelioration of the quality of life in T2D.

From a research viewpoint and in terms of future directions, more evidence is needed to address the role of pharmacological management of T2D on long-term skeletal muscle health. Myokine-based treatment has the potential to improve skeletal muscle health and provide reliable therapeutic strategies to ameliorate glucose control, positively affect body composition, and prevent and treat sarcopenia in T2D.

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## Abbreviations

AMPK	Adenosine monophosphate-activate protein kinase
DPPIV <sub>is</sub>	Dipeptidyl peptidase type IV inhibitors
FGF	Fibroblast growth factor
FGF19	Fibroblast growth factor 19
GLP-1	Glucagon-like peptide 1
GLP-1RAs	GLP-1 receptor agonists
GIP	Glucose-dependent insulinotropic polypeptide
GH	Growth hormone
IGF	Insulin-like growth factor
IL	Interleukin
mTOR	Mammalian target of rapamycin
PGC-1 $\alpha$	Peroxisome proliferator co-activator 1 alpha
SPARC	Secreted proteins acidic and rich in cysteine
Smad	Small mother against decapentaplegic
SGLT2 <sub>is</sub>	Sodium-glucose (co) transporter type 2 inhibitors
TNF $\alpha$	Tumor necrosis factor $\alpha$
T2D	Type 2 diabetes
Vit-D	Vitamin D

## References

- Santilli, V.; Bernetti, A.; Mangone, M.; Paoloni, M. Clinical definition of sarcopenia. *Clin. Cases Miner. Bone Metab.* **2014**, *11*, 177–180. [[CrossRef](#)] [[PubMed](#)]
- Cruz-Jentoft, A.J.; Bahat, G.; Bauer, J.; Boirie, Y.; Bruyère, O.; Cederholm, T.; Cooper, C.; Landi, F.; Rolland, Y.; Sayer, A.A.; et al. Sarcopenia: Revised European consensus on definition and diagnosis. *Age Ageing* **2019**, *48*, 16–31. [[CrossRef](#)] [[PubMed](#)]
- Janssen, I.; Heymsfield, S.B.; Ross, R. Low Relative Skeletal Muscle Mass (Sarcopenia) in Older Persons Is Associated with Functional Impairment and Physical Disability. *J. Am. Geriatr. Soc.* **2002**, *50*, 889–896. [[CrossRef](#)] [[PubMed](#)]
- Pacifico, J.; Geerlings, M.A.; Reijnierse, E.M.; Phassouliotis, C.; Lim, W.K.; Maier, A.B. Prevalence of sarcopenia as a comorbid disease: A systematic review and meta-analysis. *Exp. Gerontol.* **2019**, *131*, 110801. [[CrossRef](#)] [[PubMed](#)]
- Anagnostis, P.; Gkekas, N.K.; Achilla, C.; Pananastasiou, G.; Taoukidou, P.; Mitsiou, M.; Kenanidis, E.; Potoupnis, M.; Tsiridis, E.; Goulis, D.G. Type 2 Diabetes Mellitus is Associated with Increased Risk of Sarcopenia: A Systematic Review and Meta-analysis. *Calcif. Tissue Int.* **2020**, *107*, 453–463. [[CrossRef](#)] [[PubMed](#)]
- Qiao, Y.-S.; Chai, Y.-H.; Gong, H.-J.; Zhuldyz, Z.; Stehauer, C.D.A.; Zhou, J.-B.; Simó, R. The Association Between Diabetes Mellitus and Risk of Sarcopenia: Accumulated Evidences From Observational Studies. *Front. Endocrinol.* **2021**, *12*, 782391. [[CrossRef](#)] [[PubMed](#)]
- Ida, S.; Kaneko, R.; Imataka, K.; Murata, K. Association between Sarcopenia and Renal Function in Patients with Diabetes: A Systematic Review and Meta-Analysis. *J. Diabetes Res.* **2019**, *2019*, 1365189. [[CrossRef](#)]
- Wannarong, T.; Sukpornchairak, P.; Naweera, W.; Geiger, C.D.; Ungprasert, P. Association between diabetic peripheral neuropathy and sarcopenia: A systematic review and meta-analysis. *Geriatr. Gerontol. Int.* **2022**, *22*, 785–789. [[CrossRef](#)]
- Feng, L.; Gao, Q.; Hu, K.; Wu, M.; Wang, Z.; Chen, F.; Mei, F.; Zhao, L.; Ma, B. Prevalence and risk factors of sarcopenia in patients with diabetes: A meta-analysis. *J. Clin. Endocrinol. Metab.* **2021**, *107*, 1470–1483. [[CrossRef](#)]
- Corcoran, M.P.; Lamon-Fava, S.; Fielding, R.A. Skeletal muscle lipid deposition and insulin resistance: Effect of dietary fatty acids and exercise. *Am. J. Clin. Nutr.* **2007**, *85*, 662–677. [[CrossRef](#)]
- Samuel, V.T.; Shulman, G.I. The pathogenesis of insulin resistance: Integrating signaling pathways and substrate flux. *J. Clin. Invest.* **2016**, *126*, 12–22. [[CrossRef](#)] [[PubMed](#)]
- Kalyani, R.R.; Corriere, M.; Ferrucci, L. Age-related and disease-related muscle loss: The effect of diabetes, obesity, and other diseases. *Lancet Diabetes Endocrinol.* **2014**, *2*, 819–829. [[CrossRef](#)] [[PubMed](#)]
- Buford, T.W.; Anton, S.D.; Judge, A.R.; Marzetti, E.; Wohlgemuth, S.E.; Carter, C.S.; Leeuwenburgh, C.; Pahor, M.; Manini, T.M. Models of accelerated sarcopenia: Critical pieces for solving the puzzle of age-related muscle atrophy. *Ageing Res. Rev.* **2010**, *9*, 369–383. [[CrossRef](#)] [[PubMed](#)]
- Rohm, T.V.; Meier, D.T.; Olefsky, J.M.; Donath, M.Y. Inflammation in obesity, diabetes, and related disorders. *Immunity* **2022**, *55*, 31–55. [[CrossRef](#)] [[PubMed](#)]
- Luevano-Contreras, C.; Chapman-Novakofski, K. Dietary Advanced Glycation End Products and Aging. *Nutrients* **2010**, *2*, 1247–1265. [[CrossRef](#)] [[PubMed](#)]
- Peroni, D.G.; Nuzzi, G.; Trambusti, I.; Di Cicco, M.E.; Comberati, P. Microbiome Composition and Its Impact on the Development of Allergic Diseases. *Front. Immunol.* **2020**, *11*, 700. [[CrossRef](#)] [[PubMed](#)]
- Di Ciaula, A.; Bonfrate, L.; Khalil, M.; Garruti, G.; Portincasa, P. Contribution of the microbiome for better phenotyping of people living with obesity. *Rev. Endocr. Metab. Disord.* **2023**, *24*, 839–870. [[CrossRef](#)]

18. Sharma, S.; Tripathi, P. Gut microbiome and type 2 diabetes: Where we are and where to go? *J. Nutr. Biochem.* **2019**, *63*, 101–108. [[CrossRef](#)]
19. Shimizu, Y. Gut microbiota in common elderly diseases affecting activities of daily living. *World J. Gastroenterol.* **2018**, *24*, 4750–4758. [[CrossRef](#)]
20. Krok-Schoen, J.L.; Price, A.A.; Luo, M.; Kelly, O.J.; Taylor, C.A. Low Dietary Protein Intakes and Associated Dietary Patterns and Functional Limitations in an Aging Population: A NHANES Analysis. *J. Nutr. Health Aging* **2019**, *23*, 338–347. [[CrossRef](#)]
21. McCarthy, K.; Laird, E.; O'Halloran, A.M.; Walsh, C.; Healy, M.; Fitzpatrick, A.L.; Walsh, J.B.; Hernández, B.; Fallon, P.; Molloy, A.M.; et al. Association between vitamin D deficiency and the risk of prevalent type 2 diabetes and incident prediabetes: A prospective cohort study using data from The Irish Longitudinal Study on Ageing (TILDA). *EClinicalMedicine* **2022**, *53*, 101654. [[CrossRef](#)] [[PubMed](#)]
22. Kupisz-Urbańska, M.; Płudowski, P.; Marcinowska-Suchowierska, E. Vitamin D Deficiency in Older Patients—Problems of Sarcopenia, Drug Interactions, Management in Deficiency. *Nutrients* **2021**, *13*, 1247. [[CrossRef](#)]
23. Cappola, A.R.; Auchus, R.J.; Fuleihan, G.E.-H.; Handelsman, D.J.; Kalyani, R.R.; McClung, M.; A Stuenkel, C.; O Thorner, M.; Verbalis, J.G. Hormones and Aging: An Endocrine Society Scientific Statement. *J. Clin. Endocrinol. Metab.* **2023**, *108*, 1835–1874. [[CrossRef](#)] [[PubMed](#)]
24. Maliszewska, K.; Adamska-Patruno, E.; Kretowski, A. The interplay between muscle mass decline, obesity, and type 2 diabetes. *Pol. Arch. Intern. Med.* **2019**, *129*, 809–816. [[CrossRef](#)] [[PubMed](#)]
25. Buckinx, F.; Aubertin-Leheudre, M. Sarcopenia in Menopausal Women: Current Perspectives. *Int. J. Women's Health* **2022**, *14*, 805–819. [[CrossRef](#)] [[PubMed](#)]
26. McKee, A.; Morley, J.E.; Matsumoto, A.M.; Vinik, A. Sarcopenia: An Endocrine Disorder? *Endocr. Pract.* **2017**, *23*, 1143–1152. [[CrossRef](#)] [[PubMed](#)]
27. Wang, M.; Tan, Y.; Shi, Y.; Wang, X.; Liao, Z.; Wei, P. Diabetes and Sarcopenic Obesity: Pathogenesis, Diagnosis, and Treatments. *Front. Endocrinol.* **2020**, *11*, 568. [[CrossRef](#)] [[PubMed](#)]
28. Remelli, F.; Maietti, E.; Abete, P.; Bellelli, G.; Bo, M.; Cherubini, A.; Corica, F.; Di Bari, M.; Maggio, M.; Rizzo, M.R.; et al. Prevalence of obesity and diabetes in older people with sarcopenia defined according to EWGSOP2 and FNHI criteria. *Aging Clin. Exp. Res.* **2021**, *34*, 113–120. [[CrossRef](#)]
29. Baumgartner, R.N. Body Composition in Healthy Aging. *Ann. N. Y. Acad. Sci.* **2006**, *904*, 437–448. [[CrossRef](#)]
30. Davison, K.K.; Ford, E.S.; Cogswell, M.E.; Dietz, W.H.; DrPH, M.E.C. Percentage of Body Fat and Body Mass Index Are Associated with Mobility Limitations in People Aged 70 and Older from NHANES III. *J. Am. Geriatr. Soc.* **2002**, *50*, 1802–1809. [[CrossRef](#)]
31. Stenholm, S.; Harris, T.B.; Rantanen, T.; Visser, M.; Kritchevsky, S.B.; Ferrucci, L. Sarcopenic obesity: Definition, cause and consequences. *Curr. Opin. Clin. Nutr. Metab. Care* **2008**, *11*, 693–700. [[CrossRef](#)] [[PubMed](#)]
32. Hong, S.-H.; Choi, K.M. Sarcopenic Obesity, Insulin Resistance, and Their Implications in Cardiovascular and Metabolic Consequences. *Int. J. Mol. Sci.* **2020**, *21*, 494. [[CrossRef](#)] [[PubMed](#)]
33. Mesinovic, J.; Zengin, A.; De Courten, B.; Ebeling, P.R.; Scott, D. Sarcopenia and type 2 diabetes mellitus: A bidirectional relationship. *Diabetes Metab. Syndr. Obesity: Targets Ther.* **2019**, *12*, 1057–1072. [[CrossRef](#)] [[PubMed](#)]
34. Xu, Y.; Hu, T.; Shen, Y.; Wang, Y.; Bao, Y.; Ma, X. Association of skeletal muscle mass and its change with diabetes occurrence: A population-based cohort study. *Diabetol. Metab. Syndr.* **2023**, *15*, 53. [[CrossRef](#)] [[PubMed](#)]
35. Srikanthan, P.; Karlamangla, A.S. Relative Muscle Mass Is Inversely Associated with Insulin Resistance and Prediabetes. Findings from The Third National Health and Nutrition Examination Survey. *J. Clin. Endocrinol. Metab.* **2011**, *96*, 2898–2903. [[CrossRef](#)] [[PubMed](#)]
36. Hulett, N.A.; Scalzo, R.L.; Reusch, J.E.B. Glucose Uptake by Skeletal Muscle within the Contexts of Type 2 Diabetes and Exercise: An Integrated Approach. *Nutrients* **2022**, *14*, 647. [[CrossRef](#)] [[PubMed](#)]
37. Merz, K.E.; Thurmond, D.C. Role of Skeletal Muscle in Insulin Resistance and Glucose Uptake. *Compr. Physiol.* **2020**, *10*, 785–809. [[CrossRef](#)] [[PubMed](#)]
38. Henningsen, J.; Rigbolt, K.T.G.; Blagoev, B.; Pedersen, B.K.; Kratchmarova, I. Dynamics of the Skeletal Muscle Secretome during Myoblast Differentiation. *Mol. Cell. Proteom.* **2010**, *9*, 2482–2496. [[CrossRef](#)]
39. Raschke, S.; Eckel, J. Adipo-Myokines: Two Sides of the Same Coin—Mediators of Inflammation and Mediators of Exercise. *Mediat. Inflamm.* **2013**, *2013*, 320724. [[CrossRef](#)]
40. Duggal, N.A.; Niemi, G.; Harridge, S.D.R.; Simpson, R.J.; Lord, J.M. Can physical activity ameliorate immunosenescence and thereby reduce age-related multi-morbidity? *Nat. Rev. Immunol.* **2019**, *19*, 563–572. [[CrossRef](#)]
41. Anand, A.C. Nutrition and Muscle in Cirrhosis. *J. Clin. Exp. Hepatol.* **2017**, *7*, 340–357. [[CrossRef](#)] [[PubMed](#)]
42. Sartori, R.; Romanello, V.; Sandri, M. Mechanisms of muscle atrophy and hypertrophy: Implications in health and disease. *Nat. Commun.* **2021**, *12*, 330. [[CrossRef](#)] [[PubMed](#)]
43. Geladari, E.; Alexopoulos, T.; Kontogianni, M.D.; Vasilieva, L.; Mani, I.; Alexopoulou, A. Mechanisms of sarcopenia in liver cirrhosis and the role of myokines. *Ann. Gastroenterol.* **2023**, *36*, 392–404. [[CrossRef](#)] [[PubMed](#)]
44. Garneau, L.; Aguer, C. Role of myokines in the development of skeletal muscle insulin resistance and related metabolic defects in type 2 diabetes. *Diabetes Metab.* **2019**, *45*, 505–516. [[CrossRef](#)] [[PubMed](#)]
45. Eckel, J. Myokines in metabolic homeostasis and diabetes. *Diabetologia* **2019**, *62*, 1523–1528. [[CrossRef](#)] [[PubMed](#)]



46. Gianoudis, J.; Bailey, C.A.; Daly, R.M. Associations between sedentary behaviour and body composition, muscle function and sarcopenia in community-dwelling older adults. *Osteoporos. Int.* **2014**, *26*, 571–579. [[CrossRef](#)] [[PubMed](#)]
47. Hamilton, M.T.; Hamilton, D.G.; Zderic, T.W. Sedentary behavior as a mediator of type 2 diabetes. *Med. Sport Sci.* **2014**, *60*, 11–26. [[CrossRef](#)] [[PubMed](#)]
48. Eckardt, K.; Görgens, S.W.; Raschke, S.; Eckel, J. Myokines in insulin resistance and type 2 diabetes. *Diabetologia* **2014**, *57*, 1087–1099. [[CrossRef](#)]
49. Buford, T.W.; Cooke, M.B.; Manini, T.M.; Leeuwenburgh, C.; Willoughby, D.S. Effects of Age and Sedentary Lifestyle on Skeletal Muscle NF- $\kappa$ B Signaling in Men. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2010**, *65*, 532–537. [[CrossRef](#)]
50. Joseph, J.J.; Echouffo-Icheugui, J.B.; Golden, S.H.; Chen, H.; Jenny, N.S.; Carnethon, M.R.; Jacobs, D.; Burke, G.L.; Vaidya, D.; Ouyang, P.; et al. Physical activity, sedentary behaviors and the incidence of type 2 diabetes mellitus: The Multi-Ethnic Study of Atherosclerosis (MESA). *BMJ Open Diabetes Res. Care* **2016**, *4*, e000185. [[CrossRef](#)]
51. Luo, C.; Liu, R.-Y.; Zhang, G.-W.; Hu, F.; Jin, Y.-H.; Liu, B.-Y. Possible sarcopenia and risk of new-onset type 2 diabetes mellitus in older adults in China: A 7-year longitudinal cohort study. *BMC Geriatr.* **2023**, *23*, 404. [[CrossRef](#)] [[PubMed](#)]
52. Prokopidis, K.; Giannos, P.; Reginster, J.Y.; Bruyere, O.; Petrovic, M.; Cherubini, A.; Triantafyllidis, K.K.; Kechagias, K.S.; Dionyssiotis, Y.; Cesari, M.; et al. Sarcopenia is associated with a greater risk of polypharmacy and number of medications: A systematic review and meta-analysis. *J. Cachex-Sarcopenia Muscle* **2023**, *14*, 671–683. [[CrossRef](#)] [[PubMed](#)]
53. Sundar, V.V.; Ong, S.H.; Easaw, M.E.P.; Chee, W.S.S. Sarcopenia with co-existent type 2 diabetes mellitus is associated with worse clinical outcomes among hospitalised cardiac patients. *Clin. Nutr. ESPEN* **2021**, *46*, 380–385. [[CrossRef](#)] [[PubMed](#)]
54. Foong, Y.C.; Chherawala, N.; Aitken, D.; Scott, D.; Winzenberg, T.; Jones, G. Accelerometer-determined physical activity, muscle mass, and leg strength in community-dwelling older adults. *J. Cachex-Sarcopenia Muscle* **2015**, *7*, 275–283. [[CrossRef](#)] [[PubMed](#)]
55. Foong, Y.C.; Aitken, D.; Winzenberg, T.; Otahal, P.; Scott, D.; Jones, G. The association between physical activity and reduced body fat lessens with age — Results from a cross-sectional study in community-dwelling older adults. *Exp. Gerontol.* **2014**, *55*, 107–112. [[CrossRef](#)] [[PubMed](#)]
56. Yaegashi, A.; Shirahata, A.; Kudo, S.; Kozuka, M. Effects and contents of nutrition education relating to sarcopenia and frailty for Japanese older adults: A systematic review. *Geriatr. Gerontol. Int.* **2021**, *21*, 1084–1092. [[CrossRef](#)] [[PubMed](#)]
57. The Look AHEAD Research Group; Earnest, C.P.; Church, T.S.; Lee, D.; Jacobs, D.R.; Lind, L.; Lind, P.M.; Normand, M.P.; Gibson, J.L.; Yeh, H.; et al. The Look AHEAD Study: A Description of the Lifestyle Intervention and the Evidence Supporting It. *Obesity* **2006**, *14*, 737–752. [[CrossRef](#)] [[PubMed](#)]
58. Hong, J.; Kim, J.; Kim, S.W.; Kong, H.-J. Effects of home-based tele-exercise on sarcopenia among community-dwelling elderly adults: Body composition and functional fitness. *Exp. Gerontol.* **2017**, *87*(Pt. A), 33–39. [[CrossRef](#)]
59. Guo, M.; Yao, J.; Li, J.; Zhang, J.; Wang, D.; Zuo, H.; Zhang, Y.; Xu, B.; Zhong, Y.; Shen, F.; et al. Irisin ameliorates age-associated sarcopenia and metabolic dysfunction. *J. Cachex-Sarcopenia Muscle* **2022**, *14*, 391–405. [[CrossRef](#)]
60. Kistner, T.M.; Pedersen, B.K.; Lieberman, D.E. Interleukin 6 as an energy allocator in muscle tissue. *Nat. Metab.* **2022**, *4*, 170–179. [[CrossRef](#)]
61. Ozaki, Y.; Ohashi, K.; Otaka, N.; Kawanishi, H.; Takikawa, T.; Fang, L.; Takahara, K.; Tatsumi, M.; Ishihama, S.; Takefuji, M.; et al. Myonectin protects against skeletal muscle dysfunction in male mice through activation of AMPK/PGC1 $\alpha$  pathway. *Nat. Commun.* **2023**, *14*, 4675. [[CrossRef](#)] [[PubMed](#)]
62. Minniti, G.; Pescinini-Salzedas, L.M.; Minniti, G.A.d.S.; Laurindo, L.F.; Barbalho, S.M.; Sinatoro, R.V.; Sloan, L.A.; Haber, R.S.d.A.; Araújo, A.C.; Quesada, K.; et al. Organokines, Sarcopenia, and Metabolic Repercussions: The Vicious Cycle and the Interplay with Exercise. *Int. J. Mol. Sci.* **2022**, *23*, 13452. [[CrossRef](#)] [[PubMed](#)]
63. Hansen, J.S.; Rutti, S.; Arous, C.; Clemmesen, J.O.; Secher, N.H.; Drescher, A.; Gonelle-Gispert, C.; Halban, P.A.; Pedersen, B.K.; Weigert, C.; et al. Circulating Follistatin Is Liver-Derived and Regulated by the Glucagon-to-Insulin Ratio. *J. Clin. Endocrinol. Metab.* **2016**, *101*, 550–560. [[CrossRef](#)] [[PubMed](#)]
64. Lee, S.-J.; Lee, Y.-S.; Zimmers, T.A.; Soleimani, A.; Matzuk, M.M.; Tsuchida, K.; Cohn, R.D.; Barton, E.R. Regulation of Muscle Mass by Follistatin and Activins. *Mol. Endocrinol.* **2010**, *24*, 1998–2008. [[CrossRef](#)] [[PubMed](#)]
65. Yin, H.; He, H.; Shen, X.; Tang, S.; Zhao, J.; Cao, X.; Han, S.; Cui, C.; Chen, Y.; Wei, Y.; et al. MicroRNA Profiling Reveals an Abundant miR-200a-3p Promotes Skeletal Muscle Satellite Cell Development by Targeting TGF- $\beta$ 2 and Regulating the TGF- $\beta$ 2/SMAD Signaling Pathway. *Int. J. Mol. Sci.* **2020**, *21*, 3274. [[CrossRef](#)] [[PubMed](#)]
66. Baig, M.H.; Ahmad, K.; Moon, J.S.; Park, S.-Y.; Lim, J.H.; Chun, H.J.; Qadri, A.F.; Hwang, Y.C.; Jan, A.T.; Ahmad, S.S.; et al. Myostatin and its Regulation: A Comprehensive Review of Myostatin Inhibiting Strategies. *Front. Physiol.* **2022**, *13*, 876078. [[CrossRef](#)] [[PubMed](#)]
67. Umezu, T.; Nakamura, S.; Sato, Y.; Kobayashi, T.; Ito, E.; Abe, T.; Kaneko, M.; Nomura, M.; Yoshimura, A.; Oya, A.; et al. Smad2 and Smad3 expressed in skeletal muscle promote immobilization-induced bone atrophy in mice. *Biochem. Biophys. Res. Commun.* **2021**, *582*, 111–117. [[CrossRef](#)]
68. Rolland, Y.; Dray, C.; Vellas, B.; Barreto, P.D.S. Current and investigational medications for the treatment of sarcopenia. *Metabolism* **2023**, *149*, 155597. [[CrossRef](#)]
69. Coelho-Júnior, H.J.; Calvani, R.; Tosato, M.; Landi, F.; Picca, A.; Marzetti, E. Protein intake and physical function in older adults: A systematic review and meta-analysis. *Ageing Res. Rev.* **2022**, *81*, 101731. [[CrossRef](#)]



70. Ali, S.; Corbi, G.; Medoro, A.; Intrieri, M.; Scapagnini, G.; Davinelli, S. Relationship between monounsaturated fatty acids and sarcopenia: A systematic review and meta-analysis of observational studies. *Aging Clin. Exp. Res.* **2023**, *35*, 1823–1834. [[CrossRef](#)]
71. Diao, H.; Yan, F.; He, Q.; Li, M.; Zheng, Q.; Zhu, Q.; Fang, F.; Cui, W. Association between Dietary Inflammatory Index and Sarcopenia: A Meta-Analysis. *Nutrients* **2023**, *15*, 219. [[CrossRef](#)] [[PubMed](#)]
72. Jalili, C.; Talebi, S.; Bagheri, R.; Ghanavati, M.; Camera, D.M.; Amirian, P.; Zarpoosh, M.; Dizaji, M.K.; Kermani, M.A.H.; Moradi, S. The Association between Dietary Inflammatory Index and Aging Biomarkers/Conditions: A Systematic Review and Dose-response Meta-analysis. *J. Nutr. Health Aging* **2023**, *27*, 378–390. [[CrossRef](#)] [[PubMed](#)]
73. Coelho-Júnior, H.J.; Trichopoulou, A.; Panza, F. Cross-sectional and longitudinal associations between adherence to Mediterranean diet with physical performance and cognitive function in older adults: A systematic review and meta-analysis. *Ageing Res. Rev.* **2021**, *70*, 101395. [[CrossRef](#)] [[PubMed](#)]
74. Gielen, E.; Beckwée, D.; Delaere, A.; De Breucker, S.; Vandewoude, M.; Bautmans, I.; Sarcopenia Guidelines Development Group of the Belgian Society of Gerontology and Geriatrics (BSGG). Nutritional interventions to improve muscle mass, muscle strength, and physical performance in older people: An umbrella review of systematic reviews and meta-analyses. *Nutr. Rev.* **2020**, *79*, 121–147. [[CrossRef](#)]
75. Beckwée, D.; Sarcopenia Guidelines Development Group of the Belgian Society of Gerontology and Geriatrics (BSGG); Delaere, A.; Aelbrecht, S.; Baert, V.; Beudart, C.; Bruyere, O.; de Saint-Hubert, M.; Bautmans, I. Exercise Interventions for the Prevention and Treatment of Sarcopenia. A Systematic Umbrella Review. *J. Nutr. Heal. Aging* **2019**, *23*, 494–502. [[CrossRef](#)] [[PubMed](#)]
76. Wu, P.-Y.; Huang, K.-S.; Chen, K.-M.; Chou, C.-P.; Tu, Y.-K. Exercise, Nutrition, and Combined Exercise and Nutrition in Older Adults with Sarcopenia: A Systematic Review and Network Meta-analysis. *Maturitas* **2020**, *145*, 38–48. [[CrossRef](#)] [[PubMed](#)]
77. Shen, Y.; Shi, Q.; Nong, K.; Li, S.; Yue, J.; Huang, J.; Dong, B.; Beauchamp, M.; Hao, Q. Exercise for sarcopenia in older people: A systematic review and network meta-analysis. *J. Cachex-Sarcopenia Muscle* **2023**, *14*, 1199–1211. [[CrossRef](#)]
78. Cienfuegos, S.; Corapi, S.; Gabel, K.; Ezpeleta, M.; Kalam, F.; Lin, S.; Pavlou, V.; Varady, K.A. Effect of Intermittent Fasting on Reproductive Hormone Levels in Females and Males: A Review of Human Trials. *Nutrients* **2022**, *14*, 2343. [[CrossRef](#)]
79. Whittaker, J.; Wu, K. Low-fat diets and testosterone in men: Systematic review and meta-analysis of intervention studies. *J. Steroid Biochem. Mol. Biol.* **2021**, *210*, 105878. [[CrossRef](#)]
80. Fantus, R.J.; Halpern, J.A.; Chang, C.; Keeter, M.K.; Bennett, N.E.; Helfand, B.; Brannigan, R.E. The Association between Popular Diets and Serum Testosterone among Men in the United States. *J. Urol.* **2020**, *203*, 398–404. [[CrossRef](#)]
81. Whittaker, J.; Harris, M. Low-carbohydrate diets and men’s cortisol and testosterone: Systematic review and meta-analysis. *Nutr. Health* **2022**, *28*, 543–554. [[CrossRef](#)] [[PubMed](#)]
82. Whittaker, J. High-protein diets and testosterone. *Nutr. Health* **2023**, *29*, 185–191. [[CrossRef](#)] [[PubMed](#)]
83. Schmitt, C.d.S.; da Costa, C.M.; Souto, J.C.S.; Chiogna, L.M.; Santos, Z.E.d.A.; Rhoden, E.L.; Neto, B.S. The effects of a low carbohydrate diet on erectile function and serum testosterone levels in hypogonadal men with metabolic syndrome: A randomized clinical trial. *BMC Endocr. Disord.* **2023**, *23*, 30. [[CrossRef](#)]
84. Furini, C.; Spaggiari, G.; Simoni, M.; Greco, C.; Santi, D. Ketogenic state improves testosterone serum levels—Results from a systematic review and meta-analysis. *Endocrine* **2022**, *79*, 273–282. [[CrossRef](#)] [[PubMed](#)]
85. Chidi-Ogbolu, N.; Baar, K. Effect of Estrogen on Musculoskeletal Performance and Injury Risk. *Front. Physiol.* **2019**, *9*, 1834. [[CrossRef](#)] [[PubMed](#)]
86. Lisco, G.; Triggiani, D.; Giagulli, V.A.; De Pergola, G.; Guastamacchia, E.; Piazzolla, G.; Jirillo, E.; Triggiani, V. Endocrine, Metabolic, and Immune Pathogenesis of Postmenopausal Osteoporosis. Is there a Therapeutic Role in Natural Products? *Endocrine Metab. Immune Disord. Drug Targets* **2023**, *23*, 1278–1290. [[CrossRef](#)] [[PubMed](#)]
87. Tiidus, P.M. Benefits of Estrogen Replacement for Skeletal Muscle Mass and Function in Post-Menopausal Females: Evidence from Human and Animal Studies. *Eurasian J. Med.* **2011**, *43*, 109–114. [[CrossRef](#)]
88. Latham, C.M.; Brightwell, C.R.; Keeble, A.R.; Munson, B.D.; Thomas, N.T.; Zagzoog, A.M.; Fry, C.S.; Fry, J.L. Vitamin D Promotes Skeletal Muscle Regeneration and Mitochondrial Health. *Front. Physiol.* **2021**, *12*, 660498. [[CrossRef](#)]
89. Luo, J.; Quan, Z.; Lin, S.; Cui, L. The association between blood concentration of 25-hydroxyvitamin D and sarcopenia: A meta-analysis. *Asia Pac. J. Clin. Nutr.* **2018**, *27*, 1258–1270. [[CrossRef](#)]
90. Besora-Moreno, M.; Llauradó, E.; Valls, R.M.; Tarro, L.; Pedret, A.; Solà, R. Antioxidant-rich foods, antioxidant supplements, and sarcopenia in old-young adults  $\geq 55$  years old: A systematic review and meta-analysis of observational studies and randomized controlled trials. *Clin. Nutr.* **2022**, *41*, 2308–2324. [[CrossRef](#)]
91. Gkekas, N.K.; Anagnostis, P.; Paraschou, V.; Stamiris, D.; Dellis, S.; Kenanidis, E.; Potoupnis, M.; Tsiroidis, E.; Goulis, D.G. The effect of vitamin D plus protein supplementation on sarcopenia: A systematic review and meta-analysis of randomized controlled trials. *Maturitas* **2021**, *145*, 56–63. [[CrossRef](#)] [[PubMed](#)]
92. Cheng, S.-H.; Chen, K.-H.; Chen, C.; Chu, W.-C.; Kang, Y.-N. The Optimal Strategy of Vitamin D for Sarcopenia: A Network Meta-Analysis of Randomized Controlled Trials. *Nutrients* **2021**, *13*, 3589. [[CrossRef](#)] [[PubMed](#)]
93. Nasimi, N.; Sohrabi, Z.; Nunes, E.A.; Sadeghi, E.; Jamshidi, S.; Gholami, Z.; Akbarzadeh, M.; Faghih, S.; Akhlaghi, M.; Phillips, S.M. Whey Protein Supplementation with or without Vitamin D on Sarcopenia-Related Measures: A Systematic Review and Meta-Analysis. *Adv. Nutr. Int. Rev. J.* **2023**, *14*, 762–773. [[CrossRef](#)] [[PubMed](#)]
94. Chang, M.C.; Choo, Y.J. Effects of Whey Protein, Leucine, and Vitamin D Supplementation in Patients with Sarcopenia: A Systematic Review and Meta-Analysis. *Nutrients* **2023**, *15*, 521. [[CrossRef](#)] [[PubMed](#)]

95. Otremski, I.; Lev-Ran, M.; Salama, R.; Edelstein, S. The metabolism of vitamin D3 in response to testosterone. *Calcif. Tissue Int.* **1997**, *60*, 485–487. [[CrossRef](#)] [[PubMed](#)]
96. Jensen, M.B. Vitamin D and male reproduction. *Nat. Rev. Endocrinol.* **2014**, *10*, 175–186. [[CrossRef](#)] [[PubMed](#)]
97. Wang, L.; Lu, H.; Wang, S.; Liu, H.; Guo, M.; Bai, H.; Zeng, W.; Zhang, T. Vitamin D Receptor affects male mouse fertility via regulation of lipid metabolism and testosterone biosynthesis in testis. *Gene* **2022**, *834*, 146589. [[CrossRef](#)]
98. Holt, R.; Yahyavi, S.K.; Kooij, L.; Poulsen, N.N.; Juul, A.; Jørgensen, N.; Jensen, M.B. Effects of vitamin D on sex steroids, luteinizing hormone, and testosterone to luteinizing hormone ratio in 307 infertile men. *Andrology* **2023**, *in press*. [[CrossRef](#)]
99. D'andrea, S.; Martorella, A.; Coccia, F.; Castellini, C.; Minaldi, E.; Totaro, M.; Parisi, A.; Francavilla, F.; Francavilla, S.; Barbonetti, A. Relationship of Vitamin D status with testosterone levels: A systematic review and meta-analysis. *Endocrine* **2020**, *72*, 49–61. [[CrossRef](#)]
100. Hagenfeldt, Y.; Linde, K.; Sjöberg, H.-E.; Zumkeller, W.; Arver, S. Testosterone increases serum 1,25-dihydroxyvitamin D and insulin-like growth factor-I in hypogonadal men. *Int. J. Androl.* **1992**, *15*, 93–102. [[CrossRef](#)]
101. Saki, F.; Kasaei, S.R.; Sadeghian, F.; Koohepyma, F.; Omrani, G.R. Investigating the effect of testosterone by itself and in combination with letrozole on 1,25-dihydroxy vitamin D and FGF23 in male rats. *J. Endocrinol. Investig.* **2018**, *42*, 19–25. [[CrossRef](#)] [[PubMed](#)]
102. Pilz, S.; Frisch, S.; Koertke, H.; Kuhn, J.; Dreier, J.; Obermayer-Pietsch, B.; Wehr, E.; Zittermann, A. Effect of Vitamin D Supplementation on Testosterone Levels in Men. *Horm. Metab. Res.* **2010**, *43*, 223–225. [[CrossRef](#)] [[PubMed](#)]
103. Lerchbaum, E.; Pilz, S.; Trummer, C.; Schwetz, V.; Pachernegg, O.; Heijboer, A.C.; Obermayer-Pietsch, B. Vitamin D and Testosterone in Healthy Men: A Randomized Controlled Trial. *J. Clin. Endocrinol. Metab.* **2017**, *102*, 4292–4302. [[CrossRef](#)] [[PubMed](#)]
104. Hochberg, Z.; Borochowitz, Z.; Benderli, A.; Vardi, P.; Oren, S.; Gougoux, A.; Spirer, Z.; Heyman, I.; Weisman, Y. Does 1,25-Dihydroxyvitamin D Participate in the Regulation of Hormone Release from Endocrine Glands? *J. Clin. Endocrinol. Metab.* **1985**, *60*, 57–61. [[CrossRef](#)] [[PubMed](#)]
105. Hofer, D.; Münzker, J.; Schwetz, V.; Ulbing, M.; Hutz, K.; Stiegler, P.; Zigeuner, R.; Pieber, T.R.; Müller, H.; Obermayer-Pietsch, B. Testicular Synthesis and Vitamin D Action. *J. Clin. Endocrinol. Metab.* **2014**, *99*, 3766–3773. [[CrossRef](#)] [[PubMed](#)]
106. ElSayed, N.A.; Aleppo, G.; Aroda, V.R.; Bannuru, R.R.; Brown, F.M.; Bruemmer, D.; Collins, B.S.; Hilliard, M.E.; Isaacs, D.; Johnson, E.L.; et al. 9. Pharmacologic Approaches to Glycemic Treatment: Standards of Care in Diabetes—2023. *Diabetes Care* **2022**, *46* (Suppl. 1), S140–S157. [[CrossRef](#)] [[PubMed](#)]
107. Yang, Y.; Liao, Z.; Xiao, Q. Metformin ameliorates skeletal muscle atrophy in Grx1 KO mice by regulating intramuscular lipid accumulation and glucose utilization. *Biochem. Biophys. Res. Commun.* **2020**, *533*, 1226–1232. [[CrossRef](#)] [[PubMed](#)]
108. Long, D.E.; Kosmac, K.; Dungan, C.M.; Bamman, M.M.; Peterson, C.A.; Kern, P.A. Potential Benefits of Combined Statin and Metformin Therapy on Resistance Training Response in Older Individuals. *Front. Physiol.* **2022**, *13*, 872745. [[CrossRef](#)]
109. Song, Y.; Wu, Z.; Zhao, P. The Function of Metformin in Aging-Related Musculoskeletal Disorders. *Front. Pharmacol.* **2022**, *13*, 865524. [[CrossRef](#)]
110. Toledo-Pérez, R.; Lopéz-Cervantes, S.P.; Hernández-Álvarez, D.; Mena-Montes, B.; Pedraza-Vázquez, G.; Sánchez-Garibay, C.; López-Diazguerrero, N.E.; Königsberg, M.; Luna-López, A. Metformin and tBHQ Treatment Combined with an Exercise Regime Prevents Osteosarcopenic Obesity in Middle-Aged Wistar Female Rats. *Oxidative Med. Cell. Longev.* **2021**, *2021*, 5294266. [[CrossRef](#)]
111. Wahdan-Alaswad, R.; Harrell, J.C.; Fan, Z.; Edgerton, S.M.; Liu, B.; Thor, A.D. Metformin attenuates transforming growth factor beta (TGF- $\beta$ ) mediated oncogenesis in mesenchymal stem-like/claudin-low triple negative breast cancer. *Cell Cycle* **2016**, *15*, 1046–1059. [[CrossRef](#)] [[PubMed](#)]
112. Lin, H.-M.; Lee, J.-H.; Yadav, H.; Kamaraju, A.K.; Liu, E.; Zhigang, D.; Vieira, A.; Kim, S.-J.; Collins, H.; Matschinsky, F.; et al. Transforming Growth Factor- $\beta$ /Smad3 Signaling Regulates Insulin Gene Transcription and Pancreatic Islet  $\beta$ -Cell Function. *J. Biol. Chem.* **2009**, *284*, 12246–12257. [[CrossRef](#)] [[PubMed](#)]
113. Honda, M.; Makino, T.; Zhao, X.; Matsuto, M.; Sakurai, H.; Takahashi, Y.; Shimizu, M.; Sato, R.; Yamauchi, Y. Pathophysiological levels of GDF11 activate Smad2/Smad3 signaling and induce muscle atrophy in human iPSC-derived myocytes. *Am. J. Physiol. Cell Physiol.* **2022**, *323*, C1402–C1409. [[CrossRef](#)] [[PubMed](#)]
114. Kwon, B.; Querfurth, H.W. Palmitate activates mTOR/p70S6K through AMPK inhibition and hypophosphorylation of raptor in skeletal muscle cells: Reversal by oleate is similar to metformin. *Biochimie* **2015**, *118*, 141–150. [[CrossRef](#)]
115. Kang, M.J.; Moon, J.W.; Lee, J.O.; Kim, J.H.; Jung, E.J.; Kim, S.J.; Oh, J.Y.; Wu, S.W.; Lee, P.R.; Park, S.H.; et al. Metformin induces muscle atrophy by transcriptional regulation of myostatin via HDAC6 and FoxO3a. *J. Cachexia-Sarcopenia Muscle* **2021**, *13*, 605–620. [[CrossRef](#)] [[PubMed](#)]
116. Cui, M.; Gang, X.; Wang, G.; Xiao, X.; Li, Z.; Jiang, Z.; Wang, G. A cross-sectional study: Associations between sarcopenia and clinical characteristics of patients with type 2 diabetes. *Medicine* **2020**, *99*, e18708. [[CrossRef](#)]
117. Ai, Y.; Xu, R.; Liu, L. The prevalence and risk factors of sarcopenia in patients with type 2 diabetes mellitus: A systematic review and meta-analysis. *Diabetol. Metab. Syndr.* **2021**, *13*, 93. [[CrossRef](#)]
118. Witham, M.D.; Granic, A.; Pearson, E.; Robinson, S.M.; Sayer, A.A. Repurposing Drugs for Diabetes Mellitus as Potential Pharmacological Treatments for Sarcopenia—A Narrative Review. *Drugs Aging* **2023**, *40*, 703–719. [[CrossRef](#)]
119. Ashcroft, F.M. Mechanisms of the Glycaemic Effects of Sulfonylureas. *Horm. Metab. Res.* **1996**, *28*, 456–463. [[CrossRef](#)]

120. Bak, J.F.; Schmitz, O.; Sørensen, N.S.; Pedersen, O. Postreceptor effects of sulfonylurea on skeletal muscle glycogen synthase activity in type II diabetic patients. *Diabetes* **1989**, *38*, 1343–1350. [[CrossRef](#)]
121. Zhang, X.; Zhao, Y.; Chen, S.; Shao, H. Anti-diabetic drugs and sarcopenia: Emerging links, mechanistic insights, and clinical implications. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 1368–1379. [[CrossRef](#)] [[PubMed](#)]
122. Bischoff, H. The mechanism of alpha-glucosidase inhibition in the management of diabetes. *Clin. Investig. Med.* **1995**, *18*, 303–311.
123. Jiang, L.-L.; Xu, X.-H.; Luo, M.-H.; Wang, H.-Y.; Ding, B.; Yan, R.-N.; Hu, Y.; Ma, J.-H. Association of Acarbose with Decreased Muscle Mass and Function in Patients with Type 2 Diabetes: A Retrospective, Cross-Sectional Study. *Diabetes Ther.* **2021**, *12*, 2955–2969. [[CrossRef](#)] [[PubMed](#)]
124. Wang, H.; Ni, Y.; Yang, S.; Li, H.; Li, X.; Feng, B. The Effects of Gliclazide, Metformin, and Acarbose on Body Composition in Patients with Newly Diagnosed Type 2 Diabetes Mellitus. *Curr. Ther. Res. Clin. Exp.* **2013**, *75*, 88–92. [[CrossRef](#)] [[PubMed](#)]
125. Thornberry, N.A.; Gallwitz, B. Mechanism of action of inhibitors of dipeptidyl-peptidase-4 (DPP-4). *Best Pract. Res. Clin. Endocrinol. Metab.* **2009**, *23*, 479–486. [[CrossRef](#)] [[PubMed](#)]
126. A Nauck, M.; Meier, J.J. The incretin effect in healthy individuals and those with type 2 diabetes: Physiology, pathophysiology, and response to therapeutic interventions. *Lancet Diabetes Endocrinol.* **2016**, *4*, 525–536. [[CrossRef](#)] [[PubMed](#)]
127. Shyamaladevi, B.; Dash, I.; Badrachalam, R.; Krishnan, M.; Panneerselvam, A.; Undru, S. An update on diagnosis and therapeutics for type-2 diabetes mellitus. *Bioinformation* **2023**, *19*, 295–298. [[CrossRef](#)]
128. Morieri, M.L.; Consoli, A.; Sesti, G.; Purrello, F.; Avogaro, A.; Fadini, G.P.; DARWIN-FUP Network. Comparative effectiveness of dapagliflozin vs. DPP-4 inhibitors on a composite endpoint of HbA1c, body weight and blood pressure reduction in the real world. *Diabetes Metab. Res. Rev.* **2020**, *37*, e3353. [[CrossRef](#)]
129. Neidert, L.; Al-Tarhuni, M.; Goldman, D.; Kluess, H.A.; Jackson, D.N. Endogenous dipeptidyl peptidase IV modulates skeletal muscle arteriolar diameter in rats. *Physiol. Rep.* **2018**, *6*, e13564. [[CrossRef](#)]
130. Giannocco, G.; Oliveira, K.C.; Crajoinas, R.O.; Venturini, G.; Salles, T.A.; Fonseca-Alaniz, M.H.; Maciel, R.M.; Girardi, A.C. Dipeptidyl peptidase IV inhibition upregulates GLUT4 translocation and expression in heart and skeletal muscle of spontaneously hypertensive rats. *Eur. J. Pharmacol.* **2013**, *698*, 74–86. [[CrossRef](#)]
131. Lv, J.; Li, Y.; Shi, S.; Xu, X.; Wu, H.; Zhang, B.; Song, Q. Skeletal muscle mitochondrial remodeling in heart failure: An update on mechanisms and therapeutic opportunities. *Biomed. Pharmacother.* **2022**, *155*, 113833. [[CrossRef](#)] [[PubMed](#)]
132. Boschmann, M.; Engeli, S.; Dobberstein, K.; Budziarek, P.; Strauss, A.; Boehnke, J.; Sweep, F.C.G.J.; Luft, F.C.; He, Y.; Foley, J.E.; et al. Dipeptidyl-Peptidase-IV Inhibition Augments Postprandial Lipid Mobilization and Oxidation in Type 2 Diabetic Patients. *J. Clin. Endocrinol. Metab.* **2009**, *94*, 846–852. [[CrossRef](#)] [[PubMed](#)]
133. Takada, S.; Masaki, Y.; Kinugawa, S.; Matsumoto, J.; Furihata, T.; Mizushima, W.; Kadoguchi, T.; Fukushima, A.; Homma, T.; Takahashi, M.; et al. Dipeptidyl peptidase-4 inhibitor improved exercise capacity and mitochondrial biogenesis in mice with heart failure via activation of glucagon-like peptide-1 receptor signalling. *Cardiovasc. Res.* **2016**, *111*, 338–347. [[CrossRef](#)] [[PubMed](#)]
134. Yamada, S.; Ogura, Y.; Inoue, K.; Tanabe, J.; Sugaya, T.; Ohata, K.; Nagai, Y.; Natsuki, Y.; Hoshino, S.; Watanabe, S.; et al. Effect of GLP-1 receptor agonist, liraglutide, on muscle in spontaneously diabetic torii fatty rats. *Mol. Cell. Endocrinol.* **2021**, *539*, 111472. [[CrossRef](#)] [[PubMed](#)]
135. Vainshtein, A.; Desjardins, E.M.A.; Armani, A.; Sandri, M.; Hood, D.A. PGC-1 $\alpha$  modulates denervation-induced mitophagy in skeletal muscle. *Skelet. Muscle* **2015**, *5*, 9. [[CrossRef](#)] [[PubMed](#)]
136. Liu, Y.; Xu, F.; Jiang, P. Effect of sitagliptin on expression of skeletal muscle peroxisome proliferator-activated receptor  $\gamma$  coactivator-1 $\alpha$  and irisin in a rat model of type 2 diabetes mellitus. *J. Int. Med Res.* **2020**, *48*, 300060519885569. [[CrossRef](#)] [[PubMed](#)]
137. Nahon, K.J.; Doornink, F.; Straat, M.E.; Botani, K.; Martinez-Tellez, B.; Abreu-Vieira, G.; van Klinken, J.B.; Voortman, G.J.; Friesema, E.C.H.; Ruiz, J.R.; et al. Effect of sitagliptin on energy metabolism and brown adipose tissue in overweight individuals with prediabetes: A randomised placebo-controlled trial. *Diabetologia* **2018**, *61*, 2386–2397. [[CrossRef](#)]
138. Scalzo, R.L.; Rafferty, D.; Schauer, I.; Huebschmann, A.G.; Cree-Green, M.; Reusch, J.E.; Regensteiner, J.G. Sitagliptin improves diastolic cardiac function but not cardiorespiratory fitness in adults with type 2 diabetes. *J. Diabetes Complicat.* **2019**, *33*, 561–566. [[CrossRef](#)]
139. Hällsten, K.; Virtanen, K.A.; Lönnqvist, F.; Sipilä, H.; Oksanen, A.; Viljanen, T.; Rönnemaa, T.; Viikari, J.; Knuuti, J.; Nuutila, P. Rosiglitazone but Not Metformin Enhances Insulin- and Exercise-Stimulated Skeletal Muscle Glucose Uptake in Patients with Newly Diagnosed Type 2 Diabetes. *Diabetes* **2002**, *51*, 3479–3485. [[CrossRef](#)]
140. Meyer, M.M.; Levin, K.; Grimmsmann, T.; Perwitz, N.; Eirich, A.; Beck-Nielsen, H.; Klein, H.H. Troglitazone Treatment Increases Protein Kinase B Phosphorylation in Skeletal Muscle of Normoglycemic Subjects at Risk for the Development of Type 2 Diabetes. *Diabetes* **2002**, *51*, 2691–2697. [[CrossRef](#)]
141. Kim, Y.-B.; Ciaraldi, T.P.; Kong, A.; Kim, D.; Chu, N.; Mohideen, P.; Mudaliar, S.; Henry, R.R.; Kahn, B.B. Troglitazone but not Metformin Restores Insulin-Stimulated Phosphoinositide 3-Kinase Activity and Increases p110 $\beta$  Protein Levels in Skeletal Muscle of Type 2 Diabetic Subjects. *Diabetes* **2002**, *51*, 443–448. [[CrossRef](#)] [[PubMed](#)]
142. Tonelli, J.; Li, W.; Kishore, P.; Pajvani, U.B.; Kwon, E.; Weaver, C.; Scherer, P.E.; Hawkins, M. Mechanisms of Early Insulin-Sensitizing Effects of Thiazolidinediones in Type 2 Diabetes. *Diabetes* **2004**, *53*, 1621–1629. [[CrossRef](#)] [[PubMed](#)]



143. Tan, G.D.; Debard, C.; Funahashi, T.; Humphreys, S.M.; Matsuzawa, Y.; Frayn, K.N.; Karpe, F.; Vidal, H. Changes in adiponectin receptor expression in muscle and adipose tissue of type 2 diabetic patients during rosiglitazone therapy. *Diabetologia* **2005**, *48*, 1585–1589. [[CrossRef](#)] [[PubMed](#)]
144. Tyagi, S.; Gupta, P.; Saini, A.S.; Kaushal, C.; Sharma, S. The peroxisome proliferator-activated receptor: A family of nuclear receptors role in various diseases. *J. Adv. Pharm. Technol. Res.* **2011**, *2*, 236–240. [[CrossRef](#)] [[PubMed](#)]
145. Dixon, E.D.; Nardo, A.D.; Claudel, T.; Trauner, M. The Role of Lipid Sensing Nuclear Receptors (PPARs and LXR) and Metabolic Lipases in Obesity, Diabetes and NAFLD. *Genes* **2021**, *12*, 645. [[CrossRef](#)] [[PubMed](#)]
146. Polyzos, S.A.; Vachliotis, I.D.; Mantzoros, C.S. Sarcopenia, sarcopenic obesity and nonalcoholic fatty liver disease. *Metabolism* **2023**, *147*, 155676. [[CrossRef](#)] [[PubMed](#)]
147. Stuart, C.A.; Howell, M.E.; Yin, D. Overexpression of GLUT5 in Diabetic Muscle Is Reversed by Pioglitazone. *Diabetes Care* **2007**, *30*, 925–931. [[CrossRef](#)]
148. McClelland, T.J.; Fowler, A.J.; Davies, T.W.; Pearse, R.; Prowle, J.; Puthuchery, Z. Can pioglitazone be used for optimization of nutrition in critical illness? A systematic review. *J. Parenter. Enter. Nutr.* **2023**, *47*, 459–475. [[CrossRef](#)]
149. Fiorentino, T.V.; Monroy, A.; Kamath, S.; Sotero, R.; Cas, M.D.; Daniele, G.; Chavez, A.O.; Abdul-Ghani, M.; Hribal, M.L.; Sesti, G.; et al. Pioglitazone corrects dysregulation of skeletal muscle mitochondrial proteins involved in ATP synthesis in type 2 diabetes. *Metabolism* **2020**, *114*, 154416. [[CrossRef](#)]
150. Warshauer, J.T.; Lopez, X.; Gordillo, R.; Hicks, J.; Holland, W.L.; Anuwe, E.; Blankfard, M.B.; Scherer, P.E.; Lingvay, I. Effect of pioglitazone on plasma ceramides in adults with metabolic syndrome. *Diabetes Metab. Res. Rev.* **2015**, *31*, 734–744. [[CrossRef](#)]
151. Punthakee, Z.; Alméras, N.; Després, J.-P.; Dagenais, G.R.; Anand, S.S.; Hunt, D.L.; Sharma, A.M.; Jung, H.; Yusuf, S.; Gerstein, H.C. Impact of rosiglitazone on body composition, hepatic fat, fatty acids, adipokines and glucose in persons with impaired fasting glucose or impaired glucose tolerance: A sub-study of the DREAM trial. *Diabet. Med.* **2014**, *31*, 1086–1092. [[CrossRef](#)] [[PubMed](#)]
152. Yokota, T.; Kinugawa, S.; Hirabayashi, K.; Suga, T.; Takada, S.; Omokawa, M.; Kadoguchi, T.; Takahashi, M.; Fukushima, A.; Matsushima, S.; et al. Pioglitazone improves whole-body aerobic capacity and skeletal muscle energy metabolism in patients with metabolic syndrome. *J. Diabetes Investig.* **2016**, *8*, 535–541. [[CrossRef](#)] [[PubMed](#)]
153. Smith, S.R.; de Jonge, L.; Volaufova, J.; Li, Y.; Xie, H.; Bray, G.A. Effect of pioglitazone on body composition and energy expenditure: A randomized controlled trial. *Metabolism* **2005**, *54*, 24–32. [[CrossRef](#)] [[PubMed](#)]
154. Bi, Y.; Zhang, B.; Xu, W.; Yang, H.; Feng, W.; Li, C.; Tong, G.; Li, M.; Wang, X.; Shen, S.; et al. Effects of exenatide, insulin, and pioglitazone on liver fat content and body fat distributions in drug-naive subjects with type 2 diabetes. *Acta Diabetol.* **2014**, *51*, 865–873. [[CrossRef](#)]
155. Vettor, R.; Milan, G.; Franzin, C.; Sanna, M.; De Coppi, P.; Rizzuto, R.; Federspil, G. The origin of intermuscular adipose tissue and its pathophysiological implications. *Am. J. Physiol. Endocrinol. Metab.* **2009**, *297*, E987–E998. [[CrossRef](#)]
156. De Coppi, P.; Milan, G.; Scarda, A.; Boldrin, L.; Centobene, C.; Piccoli, M.; Pozzobon, M.; Pilon, C.; Pagano, C.; Gamba, P.; et al. Rosiglitazone modifies the adipogenic potential of human muscle satellite cells. *Diabetologia* **2006**, *49*, 1962–1973. [[CrossRef](#)]
157. Balas, B.; Belfort, R.; Harrison, S.A.; Darland, C.; Finch, J.; Schenker, S.; Gastaldelli, A.; Cusi, K. Pioglitazone treatment increases whole body fat but not total body water in patients with non-alcoholic steatohepatitis. *J. Hepatol.* **2007**, *47*, 565–570. [[CrossRef](#)]
158. Bray, G.A.; Smith, S.R.; Banerji, M.A.; Tripathy, D.; Clement, S.C.; Buchanan, T.A.; Henry, R.R.; Kitabchi, A.E.; Mudaliar, S.; Musi, N.; et al. Effect of pioglitazone on body composition and bone density in subjects with prediabetes in the ACT NOW trial. *Diabetes, Obes. Metab.* **2013**, *15*, 931–937. [[CrossRef](#)]
159. Slim, R.; Ben Salem, C.; Zamy, M.; Biour, M. Pioglitazone-Induced Acute Rhabdomyolysis. *Diabetes Care* **2009**, *32*, e84. [[CrossRef](#)]
160. Hsia, D.S.; Grove, O.; Cefalu, W.T. An update on sodium-glucose co-transporter-2 inhibitors for the treatment of diabetes mellitus. *Curr. Opin. Endocrinol. Diabetes Obes.* **2017**, *24*, 73–79. [[CrossRef](#)]
161. Abdul-Ghani, M.A.; DeFronzo, R.A.; Norton, L. Novel Hypothesis to Explain Why SGLT2 Inhibitors Inhibit Only 30–50% of Filtered Glucose Load in Humans. *Diabetes* **2013**, *62*, 3324–3328. [[CrossRef](#)] [[PubMed](#)]
162. Ferrannini, G.; Hach, T.; Crowe, S.; Sanghvi, A.; Hall, K.D.; Ferrannini, E. Energy Balance After Sodium–Glucose Cotransporter 2 Inhibition. *Diabetes Care* **2015**, *38*, 1730–1735. [[CrossRef](#)] [[PubMed](#)]
163. Schork, A.; Saynisch, J.; Vosseler, A.; Jaghutriz, B.A.; Heyne, N.; Peter, A.; Häring, H.-U.; Stefan, N.; Fritsche, A.; Artunc, F. Effect of SGLT2 inhibitors on body composition, fluid status and renin–angiotensin–aldosterone system in type 2 diabetes: A prospective study using bioimpedance spectroscopy. *Cardiovasc. Diabetol.* **2019**, *18*, 46. [[CrossRef](#)] [[PubMed](#)]
164. Fathi, A.; Vickneson, K.; Singh, J.S. SGLT2-inhibitors; more than just glycosuria and diuresis. *Heart Fail. Rev.* **2020**, *26*, 623–642. [[CrossRef](#)] [[PubMed](#)]
165. Xu, L.; Nagata, N.; Nagashimada, M.; Zhuge, F.; Ni, Y.; Chen, G.; Mayoux, E.; Kaneko, S.; Ota, T. SGLT2 Inhibition by Empagliflozin Promotes Fat Utilization and Browning and Attenuates Inflammation and Insulin Resistance by Polarizing M2 Macrophages in Diet-induced Obese Mice. *EBioMedicine* **2017**, *20*, 137–149. [[CrossRef](#)]
166. Monzo, L.; Ferrari, I.; Cicogna, F.; Tota, C.; Cice, G.; Girerd, N.; Calò, L. Sodium-glucose co-transporter 2 inhibitors in heart failure: An updated evidence-based practical guidance for clinicians. *Eur. Heart J. Suppl.* **2023**, *25* (Suppl. C), C309–C315. [[CrossRef](#)]
167. Sargeant, J.A.; Henson, J.; King, J.A.; Yates, T.; Khunti, K.; Davies, M.J. A Review of the Effects of Glucagon-Like Peptide-1 Receptor Agonists and Sodium-Glucose Cotransporter 2 Inhibitors on Lean Body Mass in Humans. *Endocrinol. Metab.* **2019**, *34*, 247–262. [[CrossRef](#)]

168. Sugiyama, S.; Jinnouchi, H.; Kurinami, N.; Hieshima, K.; Yoshida, A.; Jinnouchi, K.; Nishimura, H.; Suzuki, T.; Miyamoto, F.; Kajiwara, K.; et al. Dapagliflozin Reduces Fat Mass without Affecting Muscle Mass in Type 2 Diabetes. *J. Atheroscler. Thromb.* **2018**, *25*, 467–476. [[CrossRef](#)]
169. Pan, R.; Zhang, Y.; Wang, R.; Xu, Y.; Ji, H.; Zhao, Y. Effect of SGLT-2 inhibitors on body composition in patients with type 2 diabetes mellitus: A meta-analysis of randomized controlled trials. *PLoS ONE* **2022**, *17*, e0279889. [[CrossRef](#)]
170. Kalaitzoglou, E.; Fowlkes, J.L.; Popescu, I.; Thrailkill, K.M. Diabetes pharmacotherapy and effects on the musculoskeletal system. *Diabetes Metab. Res. Rev.* **2018**, *35*, e3100. [[CrossRef](#)]
171. Zizola, C.; Schulze, P.C. Metabolic and structural impairment of skeletal muscle in heart failure. *Heart Fail. Rev.* **2012**, *18*, 623–630. [[CrossRef](#)] [[PubMed](#)]
172. Takada, S.; Sabe, H.; Kinugawa, S. Treatments for skeletal muscle abnormalities in heart failure: Sodium-glucose transporter 2 and ketone bodies. *Am. J. Physiol. Heart Circ. Physiol.* **2022**, *322*, H117–H128. [[CrossRef](#)] [[PubMed](#)]
173. Voorrips, S.N.; Saucedo-Orozco, H.; Sánchez-Aguilera, P.I.; De Boer, R.A.; Van der Meer, P.; Westenbrink, B.D. Could SGLT2 Inhibitors Improve Exercise Intolerance in Chronic Heart Failure? *Int. J. Mol. Sci.* **2022**, *23*, 8631. [[CrossRef](#)] [[PubMed](#)]
174. Trujillo, J.M.; Nuffer, W.; Smith, B.A. GLP-1 receptor agonists: An updated review of head-to-head clinical studies. *Ther. Adv. Endocrinol. Metab.* **2021**, *12*, 2042018821997320. [[CrossRef](#)] [[PubMed](#)]
175. Drucker, D.J. Mechanisms of Action and Therapeutic Application of Glucagon-like Peptide-1. *Cell Metab.* **2018**, *27*, 740–756. [[CrossRef](#)] [[PubMed](#)]
176. Blundell, J.; Finlayson, G.; Axelsen, M.; Flint, A.; Gibbons, C.; Kvist, T.; Hjerpsted, J.B. Effects of once-weekly semaglutide on appetite, energy intake, control of eating, food preference and body weight in subjects with obesity. *Diabetes Obes. Metab.* **2017**, *19*, 1242–1251. [[CrossRef](#)]
177. Lisco, G.; De Tullio, A.; Guastamacchia, E.; Triggiani, V. Fixed-Ratio Combinations of Basal Insulin and GLP-1RA in the Management of Type 2 Diabetes Mellitus: Highlights from the Literature. *Endocr. Metab. Immune Disord. Drug Targets* **2021**, *21*, 626–646. [[CrossRef](#)]
178. Liu, J.-S.; Su, S.-C.; Kuo, F.-C.; Li, P.-F.; Huang, C.-L.; Ho, L.-J.; Chen, K.-C.; Liu, Y.-C.; Lin, C.-P.; Cheng, A.-C.; et al. The efficacy and safety of combined GLP-1RA and basal insulin therapy among inadequately controlled T2D with premixed insulin therapy. *Medicine* **2023**, *102*, e33167. [[CrossRef](#)]
179. Lisco, G.; De Tullio, A.; Disoteo, O.; De Geronimo, V.; Piazzolla, G.; De Pergola, G.; Giagulli, V.A.; Jirillo, E.; Guastamacchia, E.; Sabbà, C.; et al. Basal insulin intensification with GLP-1RA and dual GIP and GLP-1RA in patients with uncontrolled type 2 diabetes mellitus: A rapid review of randomized controlled trials and meta-analysis. *Front. Endocrinol.* **2022**, *13*, 920541. [[CrossRef](#)]
180. Feng, W.; Bi, Y.; Li, P.; Yin, T.; Gao, C.; Shen, S.; Gao, L.; Yang, D.; Zhu, D. Effects of liraglutide, metformin and gliclazide on body composition in patients with both type 2 diabetes and non-alcoholic fatty liver disease: A randomized trial. *J. Diabetes Investig.* **2018**, *10*, 399–407. [[CrossRef](#)]
181. Gibbons, C.; Blundell, J.; Hoff, S.T.; Dahl, K.; Bauer, R.; Bækdal, T. Effects of oral semaglutide on energy intake, food preference, appetite, control of eating and body weight in subjects with type 2 diabetes. *Diabetes Obes. Metab.* **2020**, *23*, 581–588. [[CrossRef](#)] [[PubMed](#)]
182. Bouchi, R.; Nakano, Y.; Fukuda, T.; Takeuchi, T.; Murakami, M.; Minami, I.; Izumiyama, H.; Hashimoto, K.; Yoshimoto, T.; Ogawa, Y. Reduction of visceral fat by liraglutide is associated with ameliorations of hepatic steatosis, albuminuria, and micro-inflammation in type 2 diabetic patients with insulin treatment: A randomized control trial. *Endocr. J.* **2017**, *64*, 269–281. [[CrossRef](#)] [[PubMed](#)]
183. Volpe, S.; Lisco, G.; Racaniello, D.; Fanelli, M.; Colaianni, V.; Voza, A.; Triggiani, V.; Sabbà, C.; Tortorella, C.; De Pergola, G.; et al. Once-Weekly Semaglutide Induces an Early Improvement in Body Composition in Patients with Type 2 Diabetes: A 26-Week Prospective Real-Life Study. *Nutrients* **2022**, *14*, 2414. [[CrossRef](#)] [[PubMed](#)]
184. Volpe, S.; Lisco, G.; Fanelli, M.; Racaniello, D.; Colaianni, V.; Lavarra, V.; Triggiani, D.; Crudele, L.; Triggiani, V.; Sabbà, C.; et al. Oral semaglutide improves body composition and preserves lean mass in patients with type 2 diabetes: A 26-week prospective real-life study. *Front. Endocrinol.* **2023**, *14*, 1240263. [[CrossRef](#)] [[PubMed](#)]
185. Piazzolla, G.; Voza, A.; Volpe, S.; Bergamasco, A.; Triggiani, V.; Lisco, G.; Falconieri, M.; Tortorella, C.; Solfrizzi, V.; Sabbà, C. Effectiveness and clinical benefits of new anti-diabetic drugs: A real life experience. *Open Med.* **2022**, *17*, 1203–1215. [[CrossRef](#)] [[PubMed](#)]
186. Love, K.M.; Liu, J.; Regensteiner, J.G.; Reusch, J.E.; Liu, Z. GLP-1 and insulin regulation of skeletal and cardiac muscle microvascular perfusion in type 2 diabetes. *J. Diabetes* **2020**, *12*, 488–498. [[CrossRef](#)] [[PubMed](#)]
187. Iwai, S.; Kaji, K.; Nishimura, N.; Kubo, T.; Tomooka, F.; Shibamoto, A.; Suzuki, J.; Tsuji, Y.; Fujinaga, Y.; Kitagawa, K.; et al. Glucagon-like peptide-1 receptor agonist, semaglutide attenuates chronic liver disease-induced skeletal muscle atrophy in diabetic mice. *Biochim. Biophys. Acta Mol. Basis Dis.* **2023**, *1869*, 166770. [[CrossRef](#)]
188. Xiang, J.; Qin, L.; Zhong, J.; Xia, N.; Liang, Y. GLP-1RA Liraglutide and Semaglutide Improves Obesity-Induced Muscle Atrophy via SIRT1 Pathway. *Diabetes Metab. Syndr. Obes. Targets Ther.* **2023**, *16*, 2433–2446. [[CrossRef](#)]
189. Guarnotta, V.; Bianco, M.J.; Vigneri, E.; Panto', F.; Sasso, B.L.; Ciaccio, M.; Pizzolanti, G.; Giordano, C. Effects of GLP-1 receptor agonists on myokine levels and pro-inflammatory cytokines in patients with type 2 diabetes mellitus. *Nutr. Metab. Cardiovasc. Dis.* **2021**, *31*, 3193–3201. [[CrossRef](#)]



190. Townsend, L.K.; Steinberg, G.R. AMPK and the Endocrine Control of Metabolism. *Endocr. Rev.* **2023**, *44*, 910–933. [[CrossRef](#)]
191. Khin, P.P.; Hong, Y.; Yeon, M.; Lee, D.H.; Lee, J.H.; Jun, H.-S. Dulaglutide improves muscle function by attenuating inflammation through OPA-1-TLR-9 signaling in aged mice. *Aging* **2021**, *13*, 21962–21974. [[CrossRef](#)] [[PubMed](#)]
192. Deng, F.; Wu, W.; Fan, X.; Zhong, X.; Wang, N.; Wang, Y.; Pan, T.; Du, Y. Dulaglutide Protects Mice against Diabetic Sarcopenia-Mediated Muscle Injury by Inhibiting Inflammation and Regulating the Differentiation of Myoblasts. *Int. J. Endocrinol.* **2023**, *2023*, 9926462. [[CrossRef](#)]
193. Giagulli, V.A.; Carbone, M.D.; Ramunni, M.I.; Licchelli, B.; De Pergola, G.; Sabbà, C.; Guastamacchia, E.; Triggiani, V. Adding liraglutide to lifestyle changes, metformin and testosterone therapy boosts erectile function in diabetic obese men with overt hypogonadism. *Andrology* **2015**, *3*, 1094–1103. [[CrossRef](#)]
194. Lisco, G.; Bartolomeo, N.; De Tullio, A.; De Pergola, G.; Guastamacchia, E.; Jirillo, E.; Piazzolla, G.; Triggiani, V.; Giagulli, V.A. Long-acting glucagon-like peptide 1 receptor agonists boost erectile function in men with type 2 diabetes mellitus complaining of erectile dysfunction: A retrospective cohort study. *Andrology* **2023**, *in press*. [[CrossRef](#)]
195. Zhao, F.; Zhou, Q.; Cong, Z.; Hang, K.; Zou, X.; Zhang, C.; Chen, Y.; Dai, A.; Liang, A.; Ming, Q.; et al. Structural insights into multiplexed pharmacological actions of tirzepatide and peptide 20 at the GIP, GLP-1 or glucagon receptors. *Nat. Commun.* **2022**, *13*, 1057. [[CrossRef](#)]
196. Coskun, T.; Urva, S.; Roell, W.C.; Qu, H.; Loghin, C.; Moyers, J.S.; O'farrell, L.S.; Briere, D.A.; Sloop, K.W.; Thomas, M.K.; et al. LY3437943, a novel triple glucagon, GIP, and GLP-1 receptor agonist for glycemic control and weight loss: From discovery to clinical proof of concept. *Cell Metab.* **2022**, *34*, 1234–1247.e9. [[CrossRef](#)]
197. Urva, S.; Coskun, T.; Loh, M.T.; Du, Y.; Thomas, M.K.; Gurbuz, S.; Haupt, A.; Benson, C.T.; Hernandez-Illas, M.; A D'alessio, D.; et al. LY3437943, a novel triple GIP, GLP-1, and glucagon receptor agonist in people with type 2 diabetes: A phase 1b, multicentre, double-blind, placebo-controlled, randomised, multiple-ascending dose trial. *Lancet* **2022**, *400*, 1869–1881. [[CrossRef](#)]
198. Heise, T.; DeVries, J.H.; Urva, S.; Li, J.; Pratt, E.J.; Thomas, M.K.; Mather, K.J.; Karanikas, C.A.; Dunn, J.; Haupt, A.; et al. Tirzepatide Reduces Appetite, Energy Intake, and Fat Mass in People With Type 2 Diabetes. *Diabetes Care* **2023**, *46*, 998–1004. [[CrossRef](#)]
199. Rosenstock, J.; Wysham, C.; Frías, J.P.; Kaneko, S.; Lee, C.J.; Landó, L.F.; Mao, H.; Cui, X.; A Karanikas, C.A.; Thieu, V.T. Efficacy and safety of a novel dual GIP and GLP-1 receptor agonist tirzepatide in patients with type 2 diabetes (SURPASS-1): A double-blind, randomised, phase 3 trial. *Lancet* **2021**, *398*, 143–155. [[CrossRef](#)] [[PubMed](#)]
200. Frías, J.P.; Davies, M.J.; Rosenstock, J.; Pérez Manghi, F.C.; Fernández Landó, L.; Bergman, B.K.; Liu, B.; Cui, X.; Brown, K. Tirzepatide versus Semaglutide Once Weekly in Patients with Type 2 Diabetes. *N. Engl. J. Med.* **2021**, *385*, 503–515. [[CrossRef](#)] [[PubMed](#)]
201. Ludvik, B.; Giorgino, F.; Jódar, E.; Frias, J.P.; Landó, L.F.; Brown, K.; Bray, R.; Rodríguez, Á. Once-weekly tirzepatide versus once-daily insulin degludec as add-on to metformin with or without SGLT2 inhibitors in patients with type 2 diabetes (SURPASS-3): A randomised, open-label, parallel-group, phase 3 trial. *Lancet* **2021**, *398*, 583–598. [[CrossRef](#)] [[PubMed](#)]
202. Dahl, D.; Onishi, Y.; Norwood, P.; Huh, R.; Bray, R.; Patel, H.; Rodríguez, Á. Effect of Subcutaneous Tirzepatide vs. Placebo Added to Titrated Insulin Glargine on Glycemic Control in Patients With Type 2 Diabetes: The SURPASS-5 Randomized Clinical Trial. *JAMA* **2022**, *327*, 534–545. [[CrossRef](#)] [[PubMed](#)]
203. Rosenstock, J.; Frías, J.P.; Rodbard, H.W.; Tofé, S.; Sears, E.; Huh, R.; Landó, L.F.; Patel, H. Tirzepatide vs. Insulin Lispro Added to Basal Insulin in Type 2 Diabetes: The SURPASS-6 Randomized Clinical Trial. *JAMA* **2023**, *330*, 1631. [[CrossRef](#)] [[PubMed](#)]
204. Del Prato, S.; E Kahn, S.; Pavo, I.; Weerakkody, G.J.; Yang, Z.; Doups, J.; Aizenberg, D.; Wynne, A.G.; Riesmeyer, J.S.; Heine, R.J.; et al. Tirzepatide versus insulin glargine in type 2 diabetes and increased cardiovascular risk (SURPASS-4): A randomised, open-label, parallel-group, multicentre, phase 3 trial. *Lancet* **2021**, *398*, 1811–1824. [[CrossRef](#)] [[PubMed](#)]
205. de Mesquita, Y.L.L.; Calvi, I.P.; Marques, I.R.; Cruz, S.A.; Padrao, E.M.H.; Carvalho, P.E.d.P.; da Silva, C.H.A.; Cardoso, R.; Moura, F.A.; Rafalskiy, V.V. Efficacy and safety of the dual GIP and GLP-1 receptor agonist tirzepatide for weight loss: A meta-analysis of randomized controlled trials. *Int. J. Obes.* **2023**, *47*, 883–892. [[CrossRef](#)] [[PubMed](#)]
206. Tan, B.; Pan, X.-H.; Chew, H.S.J.; Goh, R.S.J.; Lin, C.; Anand, V.V.; Lee, E.C.Z.; Chan, K.E.; Kong, G.; Ong, C.E.Y.; et al. Efficacy and safety of tirzepatide for treatment of overweight or obesity. A systematic review and meta-analysis. *Int. J. Obes.* **2023**, *47*, 677–685. [[CrossRef](#)]
207. Jastreboff, A.M.; Jastreboff, A.M.; Jastreboff, A.M.; Aronne, L.J.; Aronne, L.J.; Aronne, L.J.; Ahmad, N.N.; Ahmad, N.N.; Ahmad, N.N.; Wharton, S.; et al. Tirzepatide Once Weekly for the Treatment of Obesity. *N. Engl. J. Med.* **2022**, *387*, 205–216. [[CrossRef](#)]
208. Frias, J.P.; Jastreboff, A.M.; le Roux, C.W.; Sattar, N.; Aizenberg, D.; Mao, H.; Zhang, S.; Ahmad, N.N.; Bunck, M.C.; Benabbad, I.; et al. Tirzepatide once weekly for the treatment of obesity in people with type 2 diabetes (SURMOUNT-2): A double-blind, randomised, multicentre, placebo-controlled, phase 3 trial. *Lancet* **2023**, *402*, 613–626. [[CrossRef](#)]
209. Wadden, T.A.; Chao, A.M.; Machineri, S.; Kushner, R.; Ard, J.; Srivastava, G.; Halpern, B.; Zhang, S.; Chen, J.; Bunck, M.C.; et al. Tirzepatide after intensive lifestyle intervention in adults with overweight or obesity: The SURMOUNT-3 phase 3 trial. *Nat. Med.* **2023**, *29*, 2909–2918. [[CrossRef](#)]
210. O'brien, P.E.; Hindle, A.; Brennan, L.; Skinner, S.; Burton, P.; Smith, A.; Crosthwaite, G.; Brown, W. Long-Term Outcomes After Bariatric Surgery: A Systematic Review and Meta-analysis of Weight Loss at 10 or More Years for All Bariatric Procedures and a Single-Centre Review of 20-Year Outcomes After Adjustable Gastric Banding. *Obes. Surg.* **2018**, *29*, 3–14. [[CrossRef](#)]

211. Buchwald, H.; Avidor, Y.; Braunwald, E.; Jensen, M.D.; Pories, W.; Fahrbach, K.; Schoelles, K. Bariatric Surgery: A systematic review and meta-analysis. *JAMA* **2004**, *292*, 1724–1737. [[CrossRef](#)]
212. Sarma, S.; Palcu, P. Weight loss between glucagon-like peptide-1 receptor agonists and bariatric surgery in adults with obesity: A systematic review and meta-analysis. *Obesity* **2022**, *30*, 2111–2121. [[CrossRef](#)]
213. Yabe, D.; Kawamori, D.; Seino, Y.; Oura, T.; Takeuchi, M. Change in pharmacodynamic variables following once-weekly tirzepatide treatment versus dulaglutide in Japanese patients with type 2 diabetes (SURPASS J-mono substudy). *Diabetes Obes. Metab.* **2022**, *25*, 398–406. [[CrossRef](#)]
214. Lazzaroni, E.; Ben Nasr, M.; Loretelli, C.; Pastore, I.; Plebani, L.; Lunati, M.E.; Vallone, L.; Bolla, A.M.; Rossi, A.; Montefusco, L.; et al. Anti-diabetic drugs and weight loss in patients with type 2 diabetes. *Pharmacol. Res.* **2021**, *171*, 105782. [[CrossRef](#)]
215. Hankosky, E.R.; Wang, H.; Neff, L.M.; Kan, H.; Wang, F.; Ahmad, N.N.; Stefanski, A.; Garvey, W.T. Tirzepatide reduces the predicted risk of developing type 2 diabetes in people with obesity or overweight: Post hoc analysis of the SURMOUNT-1 trial. *Diabetes, Obes. Metab.* **2023**, *25*, 3748–3756. [[CrossRef](#)]
216. Fujita, S.; Rasmussen, B.B.; Cadenas, J.G.; Grady, J.J.; Volpi, E.; Rundqvist, H.C.; Esbjörnsson, M.; Rooyackers, O.; Österlund, T.; Moberg, M.; et al. Effect of insulin on human skeletal muscle protein synthesis is modulated by insulin-induced changes in muscle blood flow and amino acid availability. *Am. J. Physiol. Endocrinol. Metab.* **2006**, *291*, E745–E754. [[CrossRef](#)]
217. Rhoads, R.P.; Baumgard, L.H.; El-Kadi, S.W.; Zhao, L.D. PHYSIOLOGY AND ENDOCRINOLOGY SYMPOSIUM: Roles for insulin-supported skeletal muscle growth1,2. *J. Anim. Sci.* **2016**, *94*, 1791–1802. [[CrossRef](#)]
218. Somwar, R.; Sweeney, G.; Ramlal, T.; Klip, A. Stimulation of glucose and amino acid transport and activation of the insulin signaling pathways by insulin lispro in L6 skeletal muscle cells. *Clin. Ther.* **1998**, *20*, 125–140. [[CrossRef](#)]
219. Finocchietto, P.; Barreyro, F.; Holod, S.; Peralta, J.; Franco, M.C.; Méndez, C.; Converso, D.P.; Estévez, A.; Carreras, M.C.; Poderoso, J.J. Control of Muscle Mitochondria by Insulin Entails Activation of Akt2-mtNOS Pathway: Implications for the Metabolic Syndrome. *PLoS ONE* **2008**, *3*, e1749. [[CrossRef](#)] [[PubMed](#)]
220. Chu, W.L.; Chai, J.K.; Wang, X.T.; Han, S.F.; Liu, L.Y. Effects of insulin therapy on skeletal muscle wasting in severely scalded rats and its related mechanism. *Zhonghua Shao Shang Za Zhi* **2019**, *35*, 333–340. (In Chinese) [[CrossRef](#)] [[PubMed](#)]
221. Hong, O.; Choi, Y.; Kwon, H.; Jeong, H.; Son, J.; Lee, S.; Kim, S.; Yoon, K.; Yoo, S.J. Long-term insulin treatment leads to a change in myosin heavy chain fiber distribution in OLETF rat skeletal muscle. *J. Cell. Biochem.* **2019**, *120*, 2404–2412. [[CrossRef](#)]
222. Poulsen, M.K.; Henriksen, J.E.; Hother-Nielsen, O.; Beck-Nielsen, H. The Combined Effect of Triple Therapy With Rosiglitazone, Metformin, and Insulin Aspart in Type 2 Diabetic Patients. *Diabetes Care* **2003**, *26*, 3273–3279. [[CrossRef](#)]
223. Osaka, T.; Hamaguchi, M.; Fukui, M. Favorable Appendicular Skeletal Muscle Mass Changes in Older Patients with Type 2 Diabetes Receiving GLP-1 Receptor Agonist and Basal Insulin Co-Therapy. *Clin. Med. Insights Endocrinol. Diabetes* **2023**, *16*, 11795514231161885. [[CrossRef](#)]
224. Christoffersen, B.; Sanchez-Delgado, G.; John, L.M.; Ryan, D.H.; Raun, K.; Ravussin, E. Beyond appetite regulation: Targeting energy expenditure, fat oxidation, and lean mass preservation for sustainable weight loss. *Obesity* **2022**, *30*, 841–857. [[CrossRef](#)]
225. Yang, X.; Wang, L.; Zhang, L.; Zhai, X.; Sheng, X.; Quan, H.; Lin, H. Exercise mitigates Dapagliflozin-induced skeletal muscle atrophy in STZ-induced diabetic rats. *Diabetol. Metab. Syndr.* **2023**, *15*, 171, Erratum in *Diabetol. Metab. Syndr.* **2023**, *15*, 171. [[CrossRef](#)]
226. Severinsen, M.C.K.; Pedersen, B.K. Muscle–Organ Crosstalk: The Emerging Roles of Myokines. *Endocr. Rev.* **2020**, *41*, 594–609. [[CrossRef](#)]
227. Massimino, E.; Izzo, A.; Riccardi, G.; Della Pepa, G. The Impact of Glucose-Lowering Drugs on Sarcopenia in Type 2 Diabetes: Current Evidence and Underlying Mechanisms. *Cells* **2021**, *10*, 1958. [[CrossRef](#)]
228. Zheng, S.; Chen, N.; Kang, X.; Hu, Y.; Shi, S. Irisin alleviates FFA induced  $\beta$ -cell insulin resistance and inflammatory response through activating PI3K/AKT/FOXO1 signaling pathway. *Endocrine* **2022**, *M75*, 740–751. [[CrossRef](#)]
229. Zhang, Y.; Wang, L.; Kang, H.; Lin, C.-Y.; Fan, Y. Unlocking the Therapeutic Potential of Irisin: Harnessing Its Function in Degenerative Disorders and Tissue Regeneration. *Int. J. Mol. Sci.* **2023**, *24*, 6551. [[CrossRef](#)]
230. Wang, J.; Zhao, Y.T.; Zhang, L.; Dubielecka, P.M.; Qin, G.; Chin, Y.E.; Gower, A.C.; Zhuang, S.; Liu, P.Y.; Zhao, T.C. Irisin deficiency exacerbates diet-induced insulin resistance and cardiac dysfunction in type II diabetes in mice. *Am. J. Physiol. Cell Physiol.* **2023**, *325*, C1085–C1096. [[CrossRef](#)] [[PubMed](#)]
231. Colpitts, B.H.; Rioux, B.V.; Eadie, A.L.; Brunt, K.R.; Sénéchal, M. Irisin response to acute moderate intensity exercise and high intensity interval training in youth of different obesity statuses: A randomized crossover trial. *Physiol. Rep.* **2022**, *10*, e15198. [[CrossRef](#)]
232. Rad, M.M.; Bijeh, N.; Hosseini, S.R.A.; Saeb, A.R. The effect of two concurrent exercise modalities on serum concentrations of FGF21, irisin, follistatin, and myostatin in men with type 2 diabetes mellitus. *Arch. Physiol. Biochem.* **2020**, *129*, 424–433. [[CrossRef](#)]
233. Miazgowski, T.; Kaczmarkiewicz, A.; Miazgowski, B.; Kopeć, J. Cardiometabolic health, visceral fat and circulating irisin levels: Results from a real-world weight loss study. *J. Endocrinol. Investig.* **2021**, *44*, 1243–1252. [[CrossRef](#)]
234. Nadimi, H.; Djazayeri, A.; Javanbakht, M.H.; Dehpour, A.; Ghaedi, E.; Derakhshanian, H.; Mohammadi, H.; Zarei, M.; Djalali, M. The Effect of Vitamin D Supplementation on Serum and Muscle Irisin Levels, and FNDC5 Expression in Diabetic Rats. *Rep. Biochem. Mol. Biol.* **2019**, *8*, 236–243.

235. Safarpour, P.; Daneshi-Maskooni, M.; Vafa, M.; Nourbakhsh, M.; Janani, L.; Maddah, M.; Amiri, F.-S.; Mohammadi, F.; Sadeghi, H. Vitamin D supplementation improves SIRT1, Irisin, and glucose indices in overweight or obese type 2 diabetic patients: A double-blind randomized placebo-controlled clinical trial. *BMC Fam. Pract.* **2020**, *21*, 26. [[CrossRef](#)]
236. Wang, Y.; Gu, Y.; Huang, J.; Wu, H.; Meng, G.; Zhang, Q.; Liu, L.; Zhang, S.; Wang, X.; Zhang, J.; et al. Serum vitamin D status and circulating irisin levels in older adults with sarcopenia. *Front. Nutr.* **2022**, *9*, 1051870. [[CrossRef](#)]
237. Kamenov, Z.; Assyov, Y.; Angelova, P.; Gateva, A.; Tsakova, A. Irisin and Testosterone in Men with Metabolic Syndrome. *Horm. Metab. Res.* **2017**, *49*, 755–759. [[CrossRef](#)]
238. Ahmad, I.H.; Mostafa, E.R.M.; Mohammed, S.A.; Shipl, W.; Soliman, A.A.; Said, M. Correlations between serum testosterone and irisin levels in a sample of Egyptian men with metabolic syndrome; (case-control study). *Arch. Physiol. Biochem.* **2023**, *129*, 180–185. [[CrossRef](#)]
239. Zügel, M.; Qiu, S.; Laszlo, R.; Bosnyák, E.; Weigt, C.; Müller, D.; Diel, P.; Steinacker, J.M.; Schumann, U. The role of sex, adiposity, and gonadectomy in the regulation of irisin secretion. *Endocrine* **2016**, *54*, 101–110. [[CrossRef](#)]
240. Radellini, S.; Guarnotta, V.; Sciabica, V.; Pizzolanti, G.; Giordano, C. Metabolic Profile in a Cohort of Young Sicilian Patients with Klinefelter's Syndrome: The Role of Irisin. *Int. J. Endocrinol.* **2022**, *2022*, 3780741. [[CrossRef](#)]
241. Assyov, Y.; Gateva, A.; Karamfilova, V.; Gatev, T.; Nedeva, I.; Velikova, T.; Kamenov, Z.A. Impact of testosterone treatment on circulating irisin in men with late-onset hypogonadism and metabolic syndrome. *Aging Male* **2020**, *23*, 1381–1387. [[CrossRef](#)]
242. Yardimci, A.; Ulker, N.; Bulmus, O.; Sahin, E.; Alver, A.; Gungor, I.H.; Turk, G.; Artas, G.; Tektemur, N.K.; Ozcan, M.; et al. Irisin Improves High-Fat Diet-Induced Sexual Dysfunction in Obese Male Rats. *Neuroendocrinology* **2022**, *112*, 1087–1103. [[CrossRef](#)]
243. Mu, Y.; Dai, H.-G.; Luo, L.-B.; Yang, J. Irisin alleviates obesity-related spermatogenesis dysfunction via the regulation of the AMPK $\alpha$  signalling pathway. *Reprod. Biol. Endocrinol.* **2021**, *19*, 135. [[CrossRef](#)]
244. Tekin, S.; Beytur, A.; Erden, Y.; Beytur, A.; Cigremis, Y.; Vardi, N.; Turkoz, Y.; Tekedereli, I.; Sandal, S. Effects of intracerebroventricular administration of irisin on the hypothalamus–pituitary–gonadal axis in male rats. *J. Cell. Physiol.* **2018**, *234*, 8815–8824. [[CrossRef](#)]
245. Pereira, S.d.C.; Benoit, B.; Junior, F.C.A.d.A.; Chanon, S.; Vieille-Marchiset, A.; Pesenti, S.; Ruzzin, J.; Vidal, H.; Toscano, A.E. Fibroblast growth factor 19 as a countermeasure to muscle and locomotion dysfunctions in experimental cerebral palsy. *J. Cachex-Sarcopenia Muscle* **2021**, *12*, 2122–2133. [[CrossRef](#)]
246. Guo, A.; Li, K.; Xiao, Q. Fibroblast growth factor 19 alleviates palmitic acid-induced mitochondrial dysfunction and oxidative stress via the AMPK/PGC-1 $\alpha$  pathway in skeletal muscle. *Biochem. Biophys. Res. Commun.* **2020**, *526*, 1069–1076. [[CrossRef](#)]
247. Benoit, B.; Meugnier, E.; Castelli, M.; Chanon, S.; Vieille-Marchiset, A.; Durand, C.; Bendridi, N.; Pesenti, S.; Monternier, P.-A.; Durieux, A.-C.; et al. Fibroblast growth factor 19 regulates skeletal muscle mass and ameliorates muscle wasting in mice. *Nat. Med.* **2017**, *23*, 990–996. [[CrossRef](#)]
248. Soytaş, R.B.; Suzan, V.; Arman, P.; Gedik, T.E.; Unal, D.; Cengiz, M.; Bolayirli, I.M.; Erdinçler, D.S.; Doventas, A.; Yavuzer, H. Association of FGF-19 and FGF-21 levels with primary sarcopenia. *Geriatr. Gerontol. Int.* **2021**, *21*, 959–962. [[CrossRef](#)]
249. Guo, A.; Li, K.; Tian, H.; Fan, Z.; Chen, Q.; Yang, Y.; Yu, J.; Wu, Y.; Xiao, Q. FGF19 protects skeletal muscle against obesity-induced muscle atrophy, metabolic derangement and abnormal irisin levels via the AMPK/SIRT-1/PGC- $\alpha$  pathway. *J. Cell. Mol. Med.* **2021**, *25*, 3585–3600. [[CrossRef](#)]
250. Benoit, B.; Beau, A.; Bres, É.; Chanon, S.; Pinteur, C.; Vieille-Marchiset, A.; Jalabert, A.; Zhang, H.; Garg, P.; Strigini, M.; et al. Treatment with fibroblast growth factor 19 increases skeletal muscle fiber size, ameliorates metabolic perturbations and hepatic inflammation in 5/6 nephrectomized mice. *Sci. Rep.* **2023**, *13*, 5520. [[CrossRef](#)]
251. Bailey, C.J.; Flatt, P.R.; Conlon, J.M. An update on peptide-based therapies for type 2 diabetes and obesity. *Peptides* **2023**, *161*, 170939. [[CrossRef](#)]
252. Morvan, F.; Rondeau, J.-M.; Zou, C.; Minetti, G.; Scheufler, C.; Scharenberg, M.; Jacobi, C.; Brebbia, P.; Ritter, V.; Toussaint, G.; et al. Blockade of activin type II receptors with a dual anti-ActRIIA/IIB antibody is critical to promote maximal skeletal muscle hypertrophy. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 12448–12453. [[CrossRef](#)]
253. Ghanim, H.; Dhindsa, S.; Batra, M.; Green, K.; Abuaysheh, S.; Kuhadiya, N.D.; Makdissi, A.; Chaudhuri, A.; Dandona, P. Effect of Testosterone on FGF2, MRF4, and Myostatin in Hypogonadotropic Hypogonadism: Relevance to Muscle Growth. *J. Clin. Endocrinol. Metab.* **2019**, *104*, 2094–2102. [[CrossRef](#)]
254. Liu, W.; Thomas, S.G.; Asa, S.L.; Gonzalez-Cadavid, N.; Bhasin, S.; Ezzat, S. Myostatin Is a Skeletal Muscle Target of Growth Hormone Anabolic Action. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 5490–5496. [[CrossRef](#)]
255. Campbell, C.; McMillan, H.J.; Mah, J.K.; Tarnopolsky, M.; Selby, K.; McClure, T.; Wilson, D.M.; Sherman, M.L.; Escolar, D.; Attie, K.M. Myostatin inhibitor ACE-031 treatment of ambulatory boys with Duchenne muscular dystrophy: Results of a randomized, placebo-controlled clinical trial. *Muscle Nerve* **2016**, *55*, 458–464. [[CrossRef](#)]

256. Barrett, D.; Bilic, S.; Chyung, Y.; Cote, S.M.; Iarrobino, R.; Kacena, K.; Kalra, A.; Long, K.; Nomikos, G.; Place, A.; et al. A Randomized Phase 1 Safety, Pharmacokinetic and Pharmacodynamic Study of the Novel Myostatin Inhibitor Apitegromab (SRK-015): A Potential Treatment for Spinal Muscular Atrophy. *Adv. Ther.* **2021**, *38*, 3203–3222. [[CrossRef](#)]
257. Becker, C.; Lord, S.R.; A Studenski, S.; Warden, S.J.; A Fielding, R.; Recknor, C.P.; Hochberg, M.C.; Ferrari, S.L.; Blain, H.; Binder, E.F.; et al. Myostatin antibody (LY2495655) in older weak fallers: A proof-of-concept, randomised, phase 2 trial. *Lancet Diabetes Endocrinol.* **2015**, *3*, 948–957. [[CrossRef](#)]

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