The 2,6-Xylyl Moiety as a Privileged Scaffold of Pharmaceutical Significance

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The 2,6-xylyl moiety is an attractive scaffold extensively used in the field of medicinal chemistry. The local anesthetic (LA) Lidocaine (or lignocaine, Xylocaine®, Tab. 1) was discovered in 1943, and originally labelled as LL30. Thanks to the presence of the 2,6-xylyl moiety, unlike its congeners, it overcame the systemic toxicity of Tetracaine and Cocaine and had a longer duration of action than Procaine. "Xylocaine® has, for more than three decades, stood the test as a reliable and highly efficient local anesthetic" [1]. Some constrained analogues, bearing a pipecolylxylidide moiety, were then studied: Mepivacaine was introduced in the 1950s, followed by Bupivacaine, in 1965, that gained popularity because of its long duration of action. The closely related compound Ropivacaine was evaluated in clinical trials starting in 1990 and introduced in clinics in 1996. It was less neurotoxic and cardiotoxic than Bupivacaine [2,3]. Pyrrocaine is a LA drug bearing a pyrrolidine moiety used as the hydrochloride salt [4]. Etidocaine is a long-lasting LA used in gynecology and obstetrics, which alleged toxicity has restricted its clinical use. Recently, ionic gradient liposomes have been proposed to increase the upload and prolong the drug release, from liposomes [5]. Interestingly, the use of LAs during clinical cancer surgeries has suggested their potential role in cancer [6], to be used not as stand-alone anticancer drugs but preferably as chemotherapy synergists [7]. Specifically, Lidocaine showed antiproliferative effects in vitro on different types of cancer cells (breast, colon, tongue or melanoma) [8]: it selectively inhibits colon cancer cells proliferation without affecting tumor microenvironment (cancer-associated fibroblasts) [9,10]. Thus, its repositioning to anticancer has been proposed [11]. Recent studies on LAs metabolism and toxicity have been reported, also focusing on interaction with other drugs and food [12-19]. Mexiletine and tocainide (Fig. 2) are 2,6-xylyl-substituted derivatives belonging to IB class antiarrhythmic drugs. Their antiarrhythmic activity is due to the block of human cardiac voltage gated

sodium channels, namely hNav1.5 [20,21]. Moreover, as blockers of voltage-gated $Na⁺$ channels, they have been extensively studied for their antimyotonic activity [22,23], acting on human skeletal muscle voltage gated sodium channels (hNav1.4), and for the activity in neuropathic pain, exerted by blocking hNav1.7 and hNav1.8 channels, which are sodium channel subtypes predominantly expressed in peripheral nociceptive neurons [24]. Several analogues of these compounds, even more active than the parent compound, were described in the literature, both in their racemic and/or optically active forms [25-28]. The deuterated phenyl analogue of mexiletine, bearing the 2,6-xylyl portion, has been recently reported: it shows superior cardiovascular, metabolic and pharmacokinetic properties, and improved safety profiles [29]. Moreover, several metabolites of mexiletine have been studied for their antiarrhythmic and/or antimyotonic properties [30,31]. Interestingly, the metabolite m-hydroxymexiletine (MHM) was 2-fold more potent than the parent compound on the cardiac sodium channels showing more favorable toxicological properties than mexiletine since it did not impair motor coordination in contrast to mexiletine and showed no cytotoxicity on human hepatocellular liver carcinoma (HepG2) cells [32,33]. Lidoflazine is an old atypical calcium channel modulator as it is defined a non-dihydropyridine; it has been recently reconsidered [34,35]. Ranolazine (Ranexa™) is a drug approved by Food and Drug Administration (FDA) in 2006 for the treatment of chronic stable angina pectoris and as an antiarrhythmic [36]. This property is exerted by blocking late sodium currents over peak fast sodium currents. Ranolazine has also demonstrated hypoglycemic properties in pre-clinical and clinical studies, through the inhibition of glucagon secretion from pancreatic islets, as it blocks sodium channels (Nav1.3 isoform) in α -cells. It has been recently demonstrated also to be effective against cognitive impairment and depressive-like behavior [37]. Lidamidine is a derivative of guanylurea, which is a drug used to treat diarrhea, by affecting peripheral alpha-2 adrenoceptors [38]; it can also show anticoagulant and analgesic effects [39]. Lopinavir (ABT-678) is an extremely potent inhibitor of HIV-1 protease ($K_i = 1.3$ pM) that was licensed in combination with ritonavir (as KaletraTM, Abbott Laboratories) for anti-retroviral therapy in 2000 [40]. Its metabolism and doses are currently under study for the emerging Coronavirus

Disease 2019 (COVID-19), in association with the antiviral ritonavir [41,42]. Xylazine is a novel adulterant with fentanyl in fatal drug intoxications, which has public health, safety, and criminal investigative implications. It is a non-narcotic sedative used for analgesia and muscle relaxation uniquely in veterinary medicine [43].

The 2,6-xylyl moiety also characterize new synthetic compounds studied in the literature for antimicrobial activities. A 2,6-xylyl derivative was used to determine the X-ray crystal structure of bacterial Enterococcus faecalis thymidylate synthase (EfTS) [44,45]. Recently, antimicrobial activity was found for several analogues of triclocarban, a chlorinated highly effective and broad-spectrum antimicrobial and antiseptic agent that is often found in personal care products, despite its toxicity [46]. The new compounds belong to the class of diarylureas [47], known anticancer agents recently proposed for repositioning to antimicrobial agents [48]. Among the studied compounds, the most interesting were those bearing the 2,6-xylyl moiety: they were more active than triclocarban and, unlike triclocarban, they were not cytotoxic against the human mammary epithelial and embryonic kidney epithelial cells [49,50].

Finally, the 2,6-xylyl moiety is worthy of note in materials chemistry. Poly(2,6-dimethyl-1,4 phenylene)oxide (PPO) is a widespread high-performance commercially available polymer, being generally amorphous and exhibiting a high glass-transition temperature (T_g \approx 220 °C), which has been identified as a valid membrane material, used since ancient times [51]. Quaternized PPO copolymers have been recently described as anion exchange membranes for alkaline fuel cell application [52].

In conclusion, the 2,6-xylyl moiety may be considered an interesting and important scaffold in medicinal and materials chemistry. In the presence of this moiety, various pharmacological activities have been found, obtaining compounds that are also generally devoid of cytotoxicity.

Structure	Name	Activity [Ref]	Toxicity [Ref]
$\boldsymbol{\mathsf{H}}$ Y	Lidocaine	LA [1, 12]	$[8,11,53-55]$
H N − ∂	Mepivacaine	LA [2, 12]	$[2]$
$\boldsymbol{\mathsf{H}}$ N. ő	Ropivacaine	LA [3, 12]	[8, 55]
H	Bupivacaine	LA[3]	[8, 55]
$\frac{H}{N}$ ő	Pyrrocaine	LA [4, 12]	$[58]$
H ő	Etidocaine	LA[5]	$[5]$
$\frac{H}{N}$ NH ₂	Tocainide	Antiarrhythmic ^[21] Antimyotonic [23,26] Action in neuropathic pain [21]	$[59]$
NH ₂	Mexiletine	Antiarrhythmic [28,32] Antimyotonic ^[56] Action in neuropathic pain [57]	[28, 32]
HO. NH ₂	$\mathop{\rm MHM}\nolimits$	Antiarrhythmic ^[32] Antimyotonic ^[56]	$[32]$
	Lidoflazine	Antiarrhythmic ^[35]	$[60]$
H N. OH	Ranolazine	Treatment of chronic stable angina pectoris [36,37]	[61]
H N. н н .N. ö NH	Lidamidine	Antidiarrheal ^[38]	$[62]$

Table 1. Structures and activities of compounds bearing the 2,6-xylyl moiety described in the text

References

- 1. Gordh, T.; Gordh, T.E.; Lindqvist, K.; Warner, D.S. Lidocaine: the origin of a modern local anesthetic. Anesthesiologists, 2010, 113(6), 1433-1437.
- 2. Mosqueira, M.; Aykut, G.; Fink, R.H. Mepivacaine reduces calcium transients in isolated murine ventricular cardiomyocytes. BMC Anesthesiol., 2020, 20(1), 1-9.
- 3. Ruetsch, Y.A.; Boni, T.; Borgeat, A. From cocaine to ropivacaine: the history of local anesthetic drugs. Curr. Top. Med. Chem., 2001, 1(3), 175-182.
- 4. Marisetti, V.M.; Katari, N.K. Development and validation of RP-UPLC method for 2,6 dimethylaniline, its isomers, and related compounds using design of experiments. Chromatographia, 2021, 84(4), 359-369.
- 5. Oliveira, J.D.; Ribeiro, L.N.M.; Rodrigues da Silva, G.H.; Casadei, B.R.; Couto, V.M.; Martinez, E.F.; de Paula, E. Sustained release from ionic-gradient liposomes significantly decreases etidocaine cytotoxicity. Pharm. Res., 2018, 35.
- 6. Ni, J.; Xie, T.; Xiao, M.; Xiang, W.; Wang, L. Amide-linked local anesthetics preferentially target leukemia stem cell through inhibition of Wnt/β-catenin. Biochem. Biophys. Res. Commun., 2018, 503(2), 956-962.
- 7. Liu, H.; Dilger, J.P.; Lin, J. Effects of local anesthetics on cancer cells. Pharmacol. Ther. 2020, 212, 107558.
- 8. Chamaraux-Tran, T.N.; Mathelin, C.; Aprahamian, M.; Joshi, G.P.; et al. Antitumor effects of lidocaine on human breast cancer cells: An in vitro and in vivo experimental trial. Anticancer Res., 2018, 38, 95-105.
- 9. Tat, T.; Jurj, A.; Selicean, C.; Pasca, S.; Ionescu, D. Antiproliferative effects of propofol and lidocaine on the colon adenocarcinoma microenvironment. J. Buon., 2019, 24, 106-115.
- 10. Bundscherer, A.C.; Malsy, M.; Bitzinger, D.I.; Wiese, C.H.; et al. Effects of Lidocaine on HT-29 and SW480 colon cancer cells in vitro. Anticancer Res., 2017, 37(4), 1941-1945.
- 11. Zhou, D.; Wang, L.; Cui, Q.; Iftikhar, R.; Xia, Y.; Xu, P. Repositioning Lidocaine as an Anticancer Drug: The Role Beyond Anesthesia. Front. Cell Dev. Biol., 2020, 8, 565.
- 12. El Beheiry, H. Local Anesthetics. In Pharmacology in Clinical Neurosciences (pp. 95-102). 2020, Springer, Singapore.
- 13. Kubrova, E.; Su, M.; Galeano-Garces, C.; Galvan, M.L.; Jerez, S.; Dietz, A.B.; et al. Differences in cytotoxicity of lidocaine, ropivacaine, and bupivacaine on the viability and metabolic activity of human adipose-derived mesenchymal stem cells. Am. J. Phys. Med. Rehabil., 2021, 100(1), 82-91.
- 14. Punt, A.; Lautz, L.; Stoopen, G.; Pinckaers, N.; Rijkers, D.; Essers, M.; et al. In vitro metabolism of lidocaine in subcellular post-mitochondrial fractions and precision cut slices from cattle liver. Toxicol. in Vitro, 2021, 76, 105228.
- 15. Lin, S.; Jin, P.; Shao, C.; Lu, W.; Xiang, Q.; Jiang, Z.; et al. Lidocaine attenuates lipopolysaccharide-induced inflammatory responses and protects against endotoxemia in mice by suppressing HIF1 α -induced glycolysis. *Int. Immunopharmacol.*, **2020**, 80, 106150.
- 16. Wang, Y.; Ou-Yang, Q.G.; Huang, W.L.; Huang, H.L.; Zhuang, X.L.; Lin, Q.M.; et al. Investigation of the inhibitory effect of simvastatin on the metabolism of lidocaine both in vitro and in vivo. Drug Design, Develop. Ther., 2020, 14, 1739.
- 17. Kim, J.H.; Kang, D.W.; Choi, G.W.; Lee, S.B.; Lee, S.; Cho, H.Y. Evaluation of lidocaine and metabolite pharmacokinetics in hyaluronic acid injection. Pharmaceutics, 2021, 13(2), 203.
- 18. Al Nebaihi, H.M.; Al Batran, R.; Ussher, J.R.; Maayah, Z.H.; El-Kadi, A.O.; Brocks, D.R. Dietary-induced obesity, hepatic cytochrome P450, and lidocaine metabolism: comparative effects of high-fat diets in mice and rats and reversibility of effects with normalization of diet. J. Pharm. Sci., 2020, 109(2), 1199-1210.
- 19. Khamarova, M.V.; Davtyan, R.A.; Markov, A.; Goncharov, V.V. Effective anesthesia during various manipulations in maxillofacial surgery and in dental practice: overview of medicines. J.P.R.I., 2021, 91-96.
- 20. Catalano, A.; Carocci, A. Antiarrhythmic mexiletine: a review on synthetic routes to racemic and homochiral mexiletine and its enantioseparation. Curr. Med. Chem., 2016, 23(29), 3227-3244.
- 21. Carocci, A.; Corbo, F.; Lentini, G.; Cavalluzzi, M.M.; Franchini, C.; Catalano, A. A focus on the synthesis and pharmacokinetics of tocainide and its analogues. Curr. Med. Chem., 2018, 25(42), 5822-5834.
- 22. De Luca, A.; Talon, S.; De Bellis, M.; Desaphy, J.-F.; Franchini, C.; Lentini, G.; et al. Inhibition of skeletal muscle sodium currents by mexiletine analogues: Specific hydrophobic interactions rather than lipophilia per se account for drug therapeutic profile. Naunyn Schmiedeberg's Arch. Pharmacol., 2003, 367, 318-327.
- 23. Catalano, A.; Carocci, A.; Corbo, F.; Franchini, C.; Muraglia, M.; Scilimati, A.; et al. Constrained analogues of tocainide as potent skeletal muscle sodium channel blockers towards the development of antimyotonic agents. Eur. J. Med. Chem., 2008, 43, 2535-2540.
- 24. Drizin, I.; Gregg, R.J.; Scanio, M.J.; Shi, L.; Gross, M.F.; Atkinson, R.N.; et al. Discovery of potent furan piperazine sodium channel blockers for treatment of neuropathic pain. Bioorg. Med. Chem., 2008, 16, 6379-6386.
- 25. Catalano, A.; Franchini, C.; Carocci, A. Voltage-gated sodium channel blockers: synthesis of mexiletine analogues and homologues. Curr. Med. Chem., 2021, 28(8), 1535-1548.
- 26. Muraglia, M.; De Bellis, M.; Catalano, A.; Carocci, A.; Franchini, C.; Carrieri, A.; et al. Naryl-2,6-dimethylbenzamides, a new generation of tocainide analogues as blockers of skeletal muscle voltage-gated sodium channels. J. Med. Chem. 2014, 57, 2589-2600.
- 27. Carocci, A.; Catalano, A.; Bruno, C.; Lentini, G.; Franchini, C.; De Bellis, M.; et al. Synthesis and in vitro sodium channel blocking activity evaluation of novel homochiral mexiletine analogs. Chirality, 2010, 22(3), 299-307.
- 28. Carocci, A.; Roselli, M.; Budriesi, R.; Micucci, M.; Desaphy, J.F.; Altamura, C.; et al. Synthesis and evaluation of voltage-gated sodium channel blocking pyrroline derivatives endowed with both antiarrhythmic and antioxidant activities. *ChemMedChem.*, 2021, 16, 578–588.
- 29. Gomez-Galeno, J.; Okolotowicz, K.; Johnson, M.; McKeithan, W.L.; Mercola, M.; Cashman, J.R. Human-induced pluripotent stem cell-derived cardiomyocytes: Cardiovascular properties and metabolism and pharmacokinetics of deuterated mexiletine analogs. Pharmacol. Res. Perspect., 2021, 9(4), e00828.
- 30. De Bellis, M.; De Luca, A.; Rana, F.; Cavalluzzi, M.M.; Catalano, A.; Lentini, G.; et al. Evaluation of the pharmacological activity of the major mexiletine metabolites on skeletal muscle sodium currents. Br. J. Pharmacol. 2006, 149(3), 300-310.
- 31. Catalano, A.; Carocci, A.; Fracchiolla, G.; Franchini, C.; Lentini, G.; Tortorella, V.; et al. Stereospecific synthesis of "para-hydroxymexiletine" and sodium channel blocking activity evaluation. *Chirality*, **2004**, *16*(2), 72-78.
- 32. Catalano, A.; Desaphy, J.F.; Lentini, G.; Carocci, A.; Di Mola, A.; Bruno, C.; et al. Synthesis and toxicopharmacological evaluation of m-hydroxymexiletine, the first metabolite of mexiletine more potent than the parent compound on voltage-gated sodium channels. J. Med. Chem. 2012, 55(3), 1418-1422.
- 33. Catalano, A.; Budriesi, R.; Bruno, C.; Di Mola, A.; Defrenza, I.; Cavalluzzi, M.M.; et al. Searching for new antiarrhythmic agents: evaluation of *meta*-hydroxymexiletine enantiomers. Eur. J. Med. Chem., 2013, 65, 511-516.
- 34. Kochanek, P.M.; Manole, M.D.; Callaway, C.W. Strengthening the link between pre-clinical and clinical resuscitation research. Resuscitation, 2021, 158, 282-285.
- 35. Bisi, A.; Micucci, M.; Gobbi, S.; Belluti, F.; Budriesi, R.; Rampa, A. Cardiovascular profile of xanthone-based 1,4-dihydropyridines bearing a lidoflazine pharmacophore fragment. Molecules, 2018, 23, 3088.
- 36. Ghosh, G.C.; Ghosh, R.K.; Bandyopadhyay, D.; Chatterjee, K.; Aneja, A. Ranolazine: multifaceted role beyond coronary artery disease, a recent perspective. Heart Views, 2018, 19(3), 88.
- 37. Cassano, V.; Leo, A.; Tallarico, M.; Nesci, V.; Cimellaro, A.; Fiorentino, T.V.; et al. Metabolic and cognitive effects of ranolazine in type 2 diabetes mellitus: data from an in vivo model. Nutrients, 2020, 12, 382.
- 38. Awad, R.A.; Llorens, F.; Camelo, A.L.; Sánchez, M. A randomised double-blind placebocontrolled trial of lidamidine HCl in irritable bowel syndrome. Acta Gastroenterol. Latinoam., 2000, 30(3), 169-175.
- 39. Karimzadeh, M.; Manouchehri, N.; Saberi, D.; Niknam, K. FT-IR study and solvent-implicit and explicit effect on stepwise tautomerism of Guanylurea: M06-2X as a case of study. Spectrochim. Acta Part A: Mol. Biomol. Spectrosc., 2018, 199, 1-11.
- 40. Stoll, V.; Qin, W.; Stewart, K.D.; Jakob, C.; Park, C.; Walter, K.; et al. X-ray crystallographic structure of ABT-378 (lopinavir) bound to HIV-1 protease. Bioorg. Med. Chem., 2002, 10(8), 2803-2806.
- 41. Gregoire, M.; Le Turnier, P.; Gaborit, B.J.; Veyrac, G.; Lecomte, R.; Boutoille, D.; Canet, E.; Imbert, B.M.; Bellouard, R.; Raffi, F. Lopinavir pharmacokinetics in COVID-19 patients. J. Antimicrob. Chemother., 2020, 75, 2702–2704.
- 42. Lê, M.P.; Jaquet, P.; Patrier, J.; Wicky, P.H.; Le Hingrat, Q.; Veyrier, M.; et al. Pharmacokinetics of lopinavir/ritonavir oral solution to treat COVID-19 in mechanically ventilated ICU patients. J. Antimicrob. Chemother., 2020, 75(9), 2657-2660.
- 43. Nunez, J.; DeJoseph, M.E.; Gill, J.R. Xylazine, a veterinary tranquilizer, detected in 42 accidental fentanyl intoxication deaths. Am. J. Forensic Med. Pathol., 2021, 42(1), 9-11.
- 44. Catalano, A.; Luciani, R.; Carocci, A.; Cortesi, D.; Pozzi, C.; Borsari, C.; et al. X-ray crystal structures of *Enterococcus faecalis* thymidylate synthase with folate binding site inhibitors. Eur. J. Med. Chem., 2016, 123, 649-664.
- 45. Pozzi, C.; Ferrari, S.; Cortesi, D.; Luciani, R.; Stroud, R.M.; Catalano, A.; et al. The structure of Enterococcus faecalis thymidylate synthase provides clues about folate bacterial metabolism. Acta Crystallogr. D Biol. Crystallogr. 2012, 68, 1232-1241.
- 46. Iacopetta, D.; Catalano, A.; Ceramella, J.; Saturnino, C.; Salvagno, L.; Ielo, I.; et al. The different facets of triclocarban: a review. Molecules, 2021, 26, 2811.
- 47. Catalano, A.; Iacopetta, D.; Sinicropi, M.S.; Franchini, C. Diarylureas as antitumor agents. Appl. Sci., 2021, 11, 374.
- 48. Catalano, A.; Iacopetta, D.; Pellegrino, M.; Aquaro, S.; Franchini, C.; Sinicropi, M.S. Diarylureas: Repositioning from antitumor to antimicrobials or multi-target agents against new pandemics. Antibiotics, 2021, 10, 92.
- 49. Catalano, A.; Iacopetta, D.; Rosato, A.; Salvagno, L.; Ceramella, J.; Longo, F.; Sinicropi, M.S.; Franchini, C. Searching for small molecules as antibacterials: Non-cytotoxic diarylureas analogues of triclocarban. Antibiotics, 2021, 10, 204.
- 50. Catalano, A.; Rosato, A.; Salvagno, L.; Iacopetta, D.; Ceramella, J.; Fracchiolla, G.; et al. Benzothiazole-containing analogues of triclocarban with potent antibacterial activity. Antibiotics, 2021, 10, 803.
- 51. Toi, K.; Morel, G.; Paul, D.R. Gas sorption and transport in poly (phenylene oxide) and comparisons with other glassy polymers. J. Appl. Polym. Sci., 1982, 27(8), 2997-3005.
- 52. Liao, J.; Ruan, H.; Gao, X.; Chen, Q.; Shen, J. Exploring the acid enrichment application of piperidinium-functionalized cross-linked poly (2,6-dimethyl-1,4-phenylene oxide) anion exchange membranes in electrodialysis. J. Membr. Sci., 2021, 621, 118999.
- 53. Choi, W.; Ryu, H.; Fuwad, A.; Goh, S.; Zhou, C.; Shim, J.; et al. Quantitative Analysis of the Membrane Affinity of Local Anesthetics Using a Model Cell Membrane. Membranes, 2021, 11(8), 579.
- 54. Zhang, H.; Chen, X.; Zheng, T.; Lin, M.; Chen, P.; Liao, Y.; et al. Amitriptyline protects against lidocaine-induced neurotoxicity in SH-SY5Y cells via inhibition of BDNF-mediated autophagy. Neurotox. Res., 2021, 39(2), 133-145.
- 55. Chen, H.; Jin, Z.; Xia, F.; Fu, Z. Bupivacaine inhibits a small conductance calcium‐activated potassium type 2 channel in human embryonic kidney 293 cells. BMC Pharmacol. Toxicol., 2021, 22(1), 1-6.
- 56. Catalano, A.; Carocci, A.; Sinicropi, M.S. Mexiletine metabolites: a review. Curr. Med. Chem., 2015, 22(11), 1400-1413.
- 57. Romman, A.; Salama-Hanna, J.; Dwivedi, S. Mexiletine usage in a chronic pain clinic: indications, tolerability, and side effects. Pain Physic., 2018, 21(5), E573-9.
- 58. Masten, S.; Carson, B.L. Integrated Laboratory Systems, 2000.
- 59. De Bellis, M.; Carbonara, R.; Roussel, J.; Farinato, A.; Massari, A.; Pierno, S.; et al. Increased sodium channel use-dependent inhibition by a new potent analogue of tocainide greatly enhances in vivo antimyotonic activity. Neuropharmacology, 2017, 113, 206-216.
- 60. Mamoshina, P.; Rodriguez, B.; Bueno-Orovio, A. Toward a broader view of mechanisms of drug cardiotoxicity. Cell Rep. Med., 2021, 2, 100216.
- 61. M. Reed and D. Nicolas, "Ranolazine" in: StatPearls [Internet], StatPearls Publishing, Treasure Island (FL), 2019, https://www.ncbi.nlm.nih.gov/books/NBK507828/.
- 62. Chou, B.J.; Mir, G.N.; Brown, W.R.; Rapp, W.R.; Yelnosky, J. Toxicity studies on lidamidine hydrochloride (WHR-1142A), a novel antidiarrheal agent. Arzneimittel-forschung, 1978, 28(8a), 1471-1476.
- 63. Ruiz-Colón, K.; Chavez-Arias, C.; Díaz-Alcalá, J.E.; Martínez, M.A. Xylazine intoxication in humans and its importance as an emerging adulterant in abused drugs: A comprehensive review of the literature. Forensic Sci. Int., 2014, 240, 1-8.