

1 **Epidemiological and genetic evaluation of HEV in swine slaughtered in Sicily region (Italy)**

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25 **ABSTRACT**

26 Hepatitis E virus (HEV) is the etiological agent of acute viral hepatitis, a disease transmitted by the
27 oral-faecal route. In Europe, zoonotic transmission of HEV-3 genotype is associated with the
28 consumption of raw or slightly cooked meat of pigs and wild boars that are considered the main
29 reservoirs.

30 This work aims to assess the occurrence of swines' HEV RNA liver samples and rectal swabs
31 slaughtered in Sicily using biomolecular methods. HEV-RNA was detected in 17.5% (21/120) liver
32 samples analyzed and in 3.7% (3/81) rectal swabs examined. All positive samples were predicted as
33 genotype 3 and subtype 3c (75%). These data suggest a potential HEV transmission to humans
34 through close contact with pig breeders, veterinarians, slaughterhouse personnel, and pork meat
35 product consumption. Moreover, there are few scientific data evaluating the HEV spread around
36 pigs and humans in Sicily. Therefore, further studies are necessary to correlate humans with swine
37 serotypes and to assess the HEV presence and persistence in food and the risk during the
38 slaughtering process. These surveys allow to clarify the role of the swine species as a potential
39 source of infection for other domestic or wild animals and humans and to establish possible control
40 measures throughout the food chain.

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42 **Keywords**

43 Zoonosis; Molecular methods; Epidemiological survey; Genotype 3; Phylogenetic analysis

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49 **1.INTRODUCTION**

50 Hepatitis E virus (HEV) is the etiological agent of acute viral hepatitis, a human disease caused by
51 contaminated food or water consumption and transmitted by the faecal-oral route. In healthy
52 people, it occurs as a self-limiting acute hepatitis that shows case fatality rates between 1% and 5%
53 (Pavia et al., 2021). In pregnant women and immunocompromised individuals, HEV develops
54 fulminant hepatic failure or chronic liver damage that reaches rates up to 20% (Doceul et al., 2016;
55 Ricci et al., 2017). The burden of this infection worldwide is unknown, and it is estimated that, over
56 the past decade, there were more than 21,000 acute clinical HEV cases with 28 deaths in the
57 European Union (EU) (World Health Organization (WHO), 2021). However, this data may be
58 underestimated because human HEV disease is not notified in all EU countries. Moreover, the latter
59 have different viral surveillance systems which prevents the comparison of reported HEV cases.

60 HEV is a hepatotropic, small non-enveloped, and single-stranded RNA virus which belongs to the
61 family *Hepeviridae*, genus *Orthohepevirus*, species *Orthohepevirus A*. The latter includes five
62 different genotypes infecting humans but, of them, HEV-3 and HEV-4 were detected also in animals
63 (pigs, wild boar, deer, cows, goats, rabbits) and considered zoonotic. In fact, sero-epidemic studies
64 have shown a high prevalence of anti-HEV antibodies in the healthy population of many countries
65 (Ricci et al., 2017), suggesting zoonotic transmission of HEV due to its presence in animals, such as
66 wild boars and pigs, which are considered the reservoirs (Di Cola et al., 2021).

67 To date, HEV is recognized as an emerging foodborne zoonosis and in Europe, HEV-3 represents the
68 most common genotype detected in humans and animals while HEV-4 has been detected in humans
69 and less in pigs (Chelli et al., 2021). During the last decade, Canada, Europe, Japan, and the United
70 States reported HEV cases often related to people traveling in HEV endemic areas but, to date,
71 people with no history of traveling abroad tested positive for HEV too. In Europe, HEV-3 was
72 reported as the predominant genotype, widespread in pork, wild boar, and deer (Dalton et al., 2016;

73 De Sabato et al., 2018; Ricci et al., 2017). It was responsible for sporadic and clustered human cases
74 associated with the consumption of pigs and wild boars' meat used in food products such as liver,
75 offal products, sausages etc. In particular, HEV 3c, 3e, and 3f subtypes have been reported in both
76 animals and humans worldwide (De Sabato et al., 2020b). In Italy, the HEV-3f subtype is widespread
77 among humans and pigs followed by HEV-3e while HEV-3c is the most frequent subtype detected in
78 wild boars, but occasionally in pigs (De Sabato et al., 2020a; Pierini et al., 2021). The detection of
79 identical HEV sequences in food and patients highlighted that the consumption of processed wild
80 boar and pig meat could be related to a high risk of either HEV-3 or HEV-4 human infection (Di Cola
81 et al., 2021).

82 In particular, pigs represent a major risk of HEV transmission to humans when their contaminated
83 meat products are consumed raw or undercooked. There are several risk factors related to swine
84 HEV infection such as the pigs' age and, the type of farming. **Firstly**, a higher HEV-RNA prevalence
85 has been detected in younger pigs (mean age: 4-5 months) than older (Di Bartolo et al., 2011).
86 **Secondly**, different studies described conflicting data regarding HEV occurrence in extensive or
87 intensive farming production (Lopez-Lopez et al., 2018). Moreover, professional exposure to
88 infected animals such as wild boars or swine represents another important HEV transmission route.
89 Several studies reported that breeders, veterinarians, and slaughterhouse personnel may be
90 infected by HEV 1.5 times more than the general population suggesting an occupational risk (de
91 Schryver et al., 2015; Mrzljak et al., 2021).

92 More studies are needed to evaluate the potential risk linked to the **consumption of pork products**
93 and also to increase and improve the epidemiological surveillance information. These goals will be
94 useful to better understand and reduce the zoonotic transmission of HEV caused by infected pigs.

95 To date, In Sicily, there aren't studies focusing on the HEV prevalence in swine and no human HEV
96 cases have been reported since 2017, even if in Sicily region the pig meat products' consumption is
97 one of the highest in Italy ("ISTAT", "SEIEVA").

98 Our study aimed to assess the HEV presence and genetic variability in liver samples and rectal swabs,
99 of swine slaughtered, in the eastern area of the Sicilia region, to provide epidemiological data
100 currently not available.

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102 **2. MATERIALS AND METHODS**

103 **2.1. Sampling**

104 Between October 2021 and February 2022, 120 liver samples of pigs slaughtered in Sicily and only
105 81 rectal swabs from the same animals, were collected (Table 1). During the official veterinary
106 control conducted by the local health authority, livers (30-40 cm³) and rectal swabs were collected
107 and transported under refrigerated conditions to the laboratory and stored at -80 °C until use.

108 **2.2. Sample preparation and nucleic acid extraction**

109 Virus extraction was performed following the method described by Di Pasquale et al. (2019) with
110 appropriate modifications. Briefly, 1 g of liver was shredded with a sterile blade and was added to a
111 tube containing 3.5 ml of TRIZOL Reagent (Life Technologies, Monza, Italy) and 1 sterile tungsten
112 carbide bead (3mm diameter). Samples were homogenized for 5 min at 25 hz/s in Tissue Lyser II
113 (Qiagen, GmbH, Hilden, Germany). After mechanical disruption of the livers, samples were
114 incubated at Room Temperature (RT) for 15 min and centrifuged at 8000 x g for 20 min at 4 °C. Then
115 the supernatant was added with 0.7 ml of chloroform and vortexed for 15 sec and incubated for 15
116 min at RT. The aqueous phase obtained by centrifugation at 8000 x g for 15 min at 4°C, was
117 recovered and stored at – 80°C until use. In addition, the rectal swab samples were diluted in 1 ml

118 of PBS and centrifuged at 15700g for 1 min; the homogenate was recovered and stored at -80°C
119 until use.

120 **2.3. Touchdown RT-PCR and Touchdown hemi-nested PCR**

121 The touchdown RT-PCR (TD/RT-PCR) and Touchdown hemi-nested PCR assays were carried out as
122 described in detail by Lorusso et al. (2022). Briefly, the RNA extract was retrotranscribed by RT-PCR
123 touchdown (TD/RT-PCR) and subsequently amplified with Touchdown hemi-nested PCR (TD/hn-
124 PCR) in order to increase sensitivity. The primers (HEV-FA228, HEV-R4598 and HEV-R4565) amplified
125 a fragment of 346 bp belonging to RdRp of ORF1 (Drexler et al. 2012).

126 **2.4. Detection of amplified products**

127 TD/hn-PCR-amplified products were displayed by electrophoresis on 1.5% (w/v) agarose NA
128 (Pharmacia, Uppsala, Sweden) gel in 1X TBE buffer containing 0.089 M Tris, 0.089 M boric acid, 0.002
129 M EDTA, pH 8.0 (USB, Cleveland, OH, USA), and stained with Green Gel Safe 10000X Nucleic Acid
130 Stain (5 l/100 ml) (Fisher Molecular Biology, USA). The Gene Ruler™ 100 bp DNA Ladder molecular
131 weight marker (MBI Fermentas, Vilnius, Lithuania) was used.

132 Image acquisition was performed with Gel Doc™ EZ imager Bio-rad. Cycle sequencing was carried
133 out using BigDye Terminator Cycle chemistry (Applied Biosystems, Foster City, California, US).

134 **2.5. Data analysis**

135 Sicilian consensus sequences were obtained using the Geneious software version 10.0.5 (Biomatters
136 Ltd, New Zealand) and molecular data were combined with the dataset of sequences generated in
137 Calabria region by Lorusso et al. (2022). The two datasets were merged and dereplicated to identify
138 identical sequences (unique haplotypes) by using mothur v 1.45.3 (Schloss et al., 2009). Genotypes
139 and subtypes **of fragment of HEV sequences** were predicted through the online HEVnet tool (Mulder

140 et al., 2019). Molecular data, including the RNA-dependent RNA polymerase gene fragment sampled
141 in the Italian regions from several hosts, were downloaded from GenBank (February 2022).
142 Sequences were aligned using MAFFT (Kato et al., 2018) and the alignment was manually checked
143 using SeaView v.4.0 (Gouy et al., 2010). A Maximum Likelihood tree was built in FastTree (Price et al.,
144 2010) using GTR gamma model and HEV-4 strain was used as an outgroup (AB197673). Visualization
145 and annotation of the tree were performed in iTOL (“iTOL: Interactive Tree Of Life,” n.d.) (Letunic
146 and Bork, 2019). Matrices of pairwise p-distances (the number of differences divided by the total
147 number of sites) were generated for nucleotide and protein sequences included in the tree using
148 MEGA 11 (Tamura et al., 2021) and amino acid multialignment was generated and visualized using
149 SeaView v.4.0 (Gouy et al., 2010). A map of sampling sites was built using leaflet R package
150 (“leafletR: Interactive Web-Maps Based on the Leaflet JavaScript Library. R package version 0.4-0”).

151 **3. RESULTS**

152 The HEV-RNA was detected in 17.5% of liver samples (21/120) and in 3.7% of rectal swabs (3/81) by
153 the amplification of a 346 bp fragment. All pigs tested positive for HEV-RNA in rectal swabs gave a
154 negative result in the liver samples of the same animals and vice versa (Table 1). The generation of
155 unique sequences reduced the dataset from 24 sequences to 4 haplotypes (non-redundant unique
156 sequences, hap_1, hap_5, hap_6, hap_7) (Table 2). The phylogenetic approach of the HEVnet tool
157 identified all haplotypes belonged to the hepatitis E virus with high support (>90%) whereas
158 prediction of the subtype HEV-3c was inferred with low support (about 70%) for two haplotypes
159 (Hap_1 and Hap_5) and no subtype was inferred for Hap_6 and Hap_7 (Table 2). Evolutionary
160 prediction by the HEVnet showed all haplotypes matched FJ705359 as the closest reference
161 sequence that was previously found both in human and wild boar and in several geographical areas
162 (France, Germany, Netherlands, Sweden, UK, Thailand, Canada). Based on a total of 103 sequences,
163 after the removal of redundancies, the tree-building included 87 nucleotide sequences of 291 bp

164 (including the four haplotypes and the outgroup) (Figure 1). In the tree, our haplotypes were
165 grouped together in a clade and the German reference sampled in wild boar (FJ705359) was
166 confirmed to be the most related to the Sicilian haplotypes. Sister sequences to the clade also are
167 represented by two references, one collected in Central Italy from wild boar (MT840360) by Aprea
168 et al. (2020) and a second sampled in Bulgaria from swine (MZ519907) by Palombieri et al. (2021).
169 Details of p-distances matrices and amino acid multi-alignment are available in the Supplementary
170 Data file and Supp Fig 1.

171 4. DISCUSSION

172 The results achieved in our study highlight the HEV incidence in pigs (*Sus scrofa domesticus*)
173 slaughtered in the Sicily region, especially in the eastern part of the island, with an overall HEV-3
174 RNA presence of 17.5% (21/120) in liver samples and 3.7% (3/81) in rectal swabs. These findings
175 indicated that HEV infection is widely spread in Sicilian pigs, and they are consistent with data
176 obtained in southern Italy where HEV was detected in 16.5% of pigs' liver samples (Chelli et al.,
177 2021). However, the point prevalence value estimated in liver samples is higher if compared with
178 previous studies in Italy detecting HEV-RNA between 0 and 5% (Di Bartolo et al., 2008; Mughini-Gras
179 et al., 2017; Pavoni et al., 2015) and also, with recent European scientific works conducted in France,
180 Slovenia and Spain where up to 2.8% of liver samples tested positive for HEV (Chelli et al., 2021; Di
181 Bartolo et al., 2012). However, in Europe an higher prevalence was detected in Netherlands where
182 up to 11% of swine liver samples tested positive for HEV-RNA (Boxman et al., 2022).

183 In our study, liver and fecal samples were collected from the same pigs but no one animal tested
184 positive for HEV-RNA in both sample types. This occurrence could be related to liver sampling errors.
185 The acute phase of infection is generally referred as to the start of HEV shedding in feces. Based on
186 the infection dose and route, HEV shedding in pigs can last from 7 to 50 days (Meester et al., 2021;
187 Salines et al., 2017). However HEV RNA could be detected in the liver, bile, and other organs after

188 the end of shedding and viremia and consequently, pigs can transmit HEV to humans if infected
189 organs are consumed raw or slightly cooked (Meester et al., 2021).

190 Our study describes an HEV-RNA incidence in rectal swabs in line with results obtained by Chelli et
191 al. (2021) and Di Martino et al. (2010) where HEV-RNA was detected in 1.9% and 7.3% of Italian pigs'
192 faecal samples respectively. Nevertheless, these outcomes are lower than data detected in other
193 Italian studies that presented a higher HEV-RNA prevalence in swine rectal swabs' reaching
194 percentages from 24.8 up to 100% (Chelli et al., 2021). Our results could be justified considering
195 that most positive samples came from the same farms areas as described in Table 1. Moreover, the
196 type of farm (intensive or extensive) could have also influenced the exposure of animals to the virus,
197 increasing HEV infection in these species (Lopez-Lopez et al., 2018). Unfortunately, this information
198 was not available and future studies are needed to search for a correlation about the type of pig
199 farming with the major risk to be infected for swine and professional workers. Furthermore, in our
200 study, all pigs were 9- to 12-month-old and, consequently, it was impossible to correlate the HEV
201 positivity trends with the pigs' age.

202 Generally, HEV has been detected at high rates in animals of different age groups in Italian
203 slaughterhouses and the HEV-3f subtype has been the most frequent in humans and pigs, followed
204 by the HEV-3e, HEV-3c (De Sabato et al., 2020b). On the other hand, in our study, 18/24 (75%) HEV3
205 positive liver samples were predicted as subtype 3c with bootstrap $\geq 70\%$, even if the fragment
206 employed for the molecular analysis was short. This evidence suggests that subtype 3c is potentially
207 dominant in pigs around the Sicilian territory. Moreover, based on the results obtained in our study,
208 it is interesting to note that haplotype 1 was detected in pigs bred in different eastern Sicilian
209 farms representing one of the most widely spread haplotype. Furthermore, previous study (Lorusso
210 et al., 2022) identified the same haplotype also in liver samples of Calabrian wild boars where, in
211 the same way, it represented the main haplotype circulating. This evidence could suggest direct

212 transmission between wild animals and pigs raised on semi-free farms and/or environmental
213 contamination, as described by Montagnaro et al. (2015) and Wu et al. (2011) reporting
214 environmental interaction between wild boars and swine.

215 The limitation of sampling areas and the number of farms in which samples were taken does not
216 allow a complete evaluation of HEV prevalence in the Sicily region and a wider sampling involving
217 all the Sicilian provinces, will be necessary to carry out an epidemiological characterization and
218 define the riskiest areas. Furthermore, this study suggests that workers operating in the swine food
219 chain could be infected by the zoonotic HEV strains during industrial and domestic slaughtering
220 processes. Thus, the slaughterhouse could represent an epidemiological focal point to determine
221 HEV transmission in a geographical area. It might also support national analysis to establish risk
222 management measures.

223 Furthermore, more studies are needed to search for a correlation between swine and human HEV
224 serotypes to confirm and define the role played by this species as a reservoir and potential source
225 of infection for both domestic and wild animals, and not least for humans.

226

227 5. CONCLUSION

228 Our study highlights the high circulation of HEV among pigs slaughtered in the eastern part of Sicily;
229 this data could be extended to the whole region involving all Sicilian sampling areas and the Italian
230 peninsula. Therefore, further investigation necessary to understand the circulation of HEV
231 genotypes between the pigs farmed and consequently to establish swine HEV prevalence and the
232 real risk of infection for personnel operating in the swine food chain. The Competent Authorities
233 should enforce the risk assessment systems to establish measures for the improvement of public
234 health. Moreover, they could implement biosafety rules during the swine farming and slaughtering
235 process to contain the risk of HEV infection.

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239

240 **REFERENCES**

241 Aprea, G., Scattolini, S., D'Angelantonio, D., Chiaverini, A., Di Lollo, V., Olivieri, S., Marcacci, M.,
242 Mangone, I., Salucci, S., Antoci, S., Cammà, C., Di Pasquale, A., Migliorati, G., Pomilio, F., 2020.
243 Whole Genome Sequencing Characterization of HEV3-e and HEV3-f Subtypes among the Wild
244 Boar Population in the Abruzzo Region, Italy: First Report. *Microorganisms* 8, 1393.
245 <https://doi.org/10.3390/microorganisms8091393>

246 Boxman, I.L.A., Verhoef, L., Dop, P.Y., Vennema, H., Dirks, A.M., Opsteegh, M., 2022. *International*
247 *Journal of Food Microbiology* High prevalence of acute hepatitis E virus infection in pigs in
248 Dutch slaughterhouses 379. <https://doi.org/10.1016/j.ijfoodmicro.2022.109830>

249 Chelli, E., Suffredini, E., De Santis, P., De Medici, D., Di Bella, S., D'amato, S., Gucciardi, F., Guercio,
250 A., Ostanello, F., Perrone, V., Purpari, G., Scavia, G.S., Schembri, P., Varcasia, B.M., Di Bartolo,
251 I., 2021. Hepatitis E Virus Occurrence in Pigs Slaughtered in Italy. *Anim. an open access J.*
252 *from MDPI* 11, 1–14. <https://doi.org/10.3390/ANI11020277>

253 Dalton, H.R., Kamar, N., Izopet, J., 2016. Hepatitis E virus. *Clin. Virol.* 1209–1229.
254 <https://doi.org/10.1128/9781555819439.ch50>

255 De Sabato, L., Amoroso, M.G., Ianiro, G., Esposito, C., De Grossi, L., Fusco, G., Barone, A., Martini,
256 E., Ostanello, F., Di Bartolo, I., 2020a. Detection of Hepatitis E Virus in Livers and Muscle
257 Tissues of Wild Boars in Italy. *Food Environ. Virol.* 12, 1–8. [https://doi.org/10.1007/s12560-](https://doi.org/10.1007/s12560-019-09405-0)
258 [019-09405-0](https://doi.org/10.1007/s12560-019-09405-0)

259 De Sabato, L., Di Bartolo, I., Lapa, D., Capobianchi, M.R., Garbuglia, A.R., 2020b. Molecular
260 Characterization of HEV Genotype 3 in Italy at Human/Animal Interface. *Front. Microbiol.* 11,
261 137. <https://doi.org/10.3389/FMICB.2020.00137/BIBTEX>

262 De Sabato, L., Lemey, P., Vrancken, B., Bonfanti, L., Ceglie, L., Vaccari, G., Di Bartolo, I., 2018.
263 Proposal for a new subtype of the zoonotic genotype 3 Hepatitis E virus: HEV-3I. *Virus Res.*
264 248, 1–4. <https://doi.org/10.1016/J.VIRUSRES.2018.02.010>

265 de Schryver, A., de Schrijver, K., François, G., Hambach, R., van Sprundel, M., Tabibi, R., Colosio, C.,
266 2015. Hepatitis E virus infection: An emerging occupational risk? *Occup. Med. (Chic. Ill)*. 65,
267 667–672. <https://doi.org/10.1093/OCCMED/KQV154>

268 Di Bartolo, I. Di, Diez-Valcarce, M., Vasickova, P., Kralik, P., Hernandez, M., Angeloni, G., Ostanello,
269 F., Bouwknecht, M., Rodríguez-Lázaro, D., Pavlik, I., Ruggeri, F.M., 2012. Hepatitis E Virus in
270 Pork Production Chain in Czech Republic, Italy, and Spain, 2010 - Volume 18, Number 8—
271 August 2012 - *Emerging Infectious Diseases journal - CDC. Emerg. Infect. Dis.* 18, 1282–1289.
272 <https://doi.org/10.3201/EID1808.111783>

273 Di Bartolo, I., Martelli, F., Inglese, N., Pourshaban, M., Caprioli, A., Ostanello, F., Ruggeri, F.M.,
274 2008. Widespread diffusion of genotype 3 hepatitis E virus among farming swine in Northern
275 Italy. *Vet. Microbiol.* 132, 47–55. <https://doi.org/10.1016/J.VETMIC.2008.04.028>

276 Di Bartolo, I., Ponterio, E., Castellini, L., Ostanello, F., Ruggeri, F.M., 2011. Viral and antibody HEV
277 prevalence in swine at slaughterhouse in Italy. *Vet. Microbiol.* 149, 330–338.
278 <https://doi.org/10.1016/j.vetmic.2010.12.007>

279 Di Cola, G., Fantilli, A.C., Pisano, M.B., Ré, V.E., 2021. Foodborne transmission of hepatitis A and
280 hepatitis E viruses: A literature review. *Int. J. Food Microbiol.* 338, 108986.

281 <https://doi.org/10.1016/j.ijfoodmicro.2020.108986>

282 Di Martino, B., Di Profio, F., Martella, V., Di Felice, E., Francesco, C.E.D., Ceci, C., Marsilio, F., 2010.
283 Detection of hepatitis E virus in slaughtered pigs in Italy. *Arch. Virol.* 155, 103–106.
284 <https://doi.org/10.1007/S00705-009-0544-0/FIGURES/1>

285 Di Pasquale, S., De Santis, P., La Rosa, G., Di Domenico, K., Iaconelli, M., Micarelli, G., Martini, E.,
286 Bilei, S., De Medici, D., Suffredini, E., 2019. Quantification and genetic diversity of Hepatitis E
287 virus in wild boar (*Sus scrofa*) hunted for domestic consumption in Central Italy. *Food*
288 *Microbiol.* 82, 194–201. <https://doi.org/10.1016/j.fm.2019.02.005>

289 Doceul, V., Bagdassarian, E., Demange, A., Pavio, N., 2016. Zoonotic hepatitis E virus: Classification,
290 animal reservoirs and transmission routes. *Viruses* 8, 1–24.
291 <https://doi.org/10.3390/v8100270>

292 Drexler, J.F., Seelen, A., Corman, V.M., Fumie Tateno, A., Cottontail, V., Melim Zerbinati, R., Gloza-
293 Rausch, F., Klose, S.M., Adu-Sarkodie, Y., Opong, S.K., Kalko, E.K. V., Osterman, A., Rasche,
294 A., Adam, A., Muller, M.A., Ulrich, R.G., Leroy, E.M., Lukashev, A.N., Drosten, C., 2012. Bats
295 Worldwide Carry Hepatitis E Virus-Related Viruses That Form a Putative Novel Genus within
296 the Family Hepeviridae. *J. Virol.* 86, 9134–9147. <https://doi.org/10.1128/jvi.00800-12>

297 Gouy, M., Guindon, S., Gascuel, O., 2010. Sea view version 4: A multiplatform graphical user
298 interface for sequence alignment and phylogenetic tree building. *Mol. Biol. Evol.* 27, 221–224.
299 <https://doi.org/10.1093/molbev/msp259>

300 ISTAT, n.d.

301 iTOL: Interactive Tree Of Life [WWW Document], n.d. URL <https://itol.embl.de/> (accessed 2.23.22).

302 Katoh, K., Rozewicki, J., Yamada, K.D., 2018. MAFFT online service: Multiple sequence alignment,
303 interactive sequence choice and visualization. *Brief. Bioinform.* 20, 1160–1166.
304 <https://doi.org/10.1093/bib/bbx108>

305 leafletR: Interactive Web-Maps Based on the Leaflet JavaScript Library. R package version 0.4-0
306 [WWW Document], n.d. URL <http://cran.r-project.org/package=leafletR> (accessed 1.3.22).

307 Letunic, I., Bork, P., 2019. Interactive Tree Of Life (iTOL) v4: recent updates and new
308 developments. *Nucleic Acids Res.* 47. <https://doi.org/10.1093/NAR/GKZ239>

309 Lopez-Lopez, P., Risalde, M. de los A., Frias, M., García-Bocanegra, I., Brieva, T., Caballero-Gomez,
310 J., Camacho, A., Fernández-Molera, V., Machuca, I., Gomez-Villamandos, J.C., Rivero, A.,
311 Rivero-Juarez, A., 2018. Risk factors associated with hepatitis E virus in pigs from different
312 production systems. *Vet. Microbiol.* 224, 88–92.
313 <https://doi.org/10.1016/J.VETMIC.2018.08.020>

314 Lorusso, P., Bonerba, E., Pandiscia, A., Mottola, A., Di Pinto, A., Piredda, R., Terio, V., 2022.
315 Occurrence of hepatitis E virus (HEV) in Calabrian wild boars. *Int. J. Food Microbiol.* 371.
316 <https://doi.org/10.1016/J.IJFOODMICRO.2022.109671>

317 Meester, M., Tobias, T.J., Bouwknecht, M., Kusters, N.E., Stegeman, J.A., van der Poel, W.H.M.,
318 2021. Infection dynamics and persistence of hepatitis E virus on pig farms – a review. *Porc.*
319 *Heal. Manag.* 7, 1–16. <https://doi.org/10.1186/s40813-021-00189-z>

320 Montagnaro, S., De Martinis, C., Sasso, S., Ciarcia, R., Damiano, S., Auletta, L., Iovane, V., Zottola,
321 T., Pagnini, U., 2015. Viral and Antibody Prevalence of Hepatitis E in European Wild Boars (*Sus*
322 *scrofa*) and Hunters at Zoonotic Risk in the Latium Region. *J. Comp. Pathol.* 153, 1–8.
323 <https://doi.org/10.1016/J.JCPA.2015.04.006>

- 324 Mrzljak, A., Balen, I., Barbic, L., Ilic, M., Vilibic-Cavlek, T., 2021. Hepatitis E virus in professionally
325 exposed: A reason for concern? *World J. Hepatol.* 13, 723–730.
326 <https://doi.org/10.4254/WJH.V13.I7.723>
- 327 Mughini-Gras, L., Angeloni, G., Salata, C., Vonesch, N., D’Amico, W., Campagna, G., Natale, A.,
328 Zuliani, F., Ceglie, L., Monne, I., Vascellari, M., Capello, K., Di Martino, G., Inglese, N., Palù, G.,
329 Tomao, P., Bonfanti, L., 2017. Hepatitis E virus infection in North Italy: high seroprevalence in
330 swine herds and increased risk for swine workers. *Epidemiol. Infect.* 145, 3375.
331 <https://doi.org/10.1017/S0950268817002485>
- 332 Mulder, A.C., Kroneman, A., Franz, E., Vennema, H., Tulen, A.D., Takkinen, J., Hofhuis, A., Adlhoch,
333 C., Aberle, S., Subissi, L., Suin, V., Midgley, S., Kuznetsova, T., Izopet, J., Pavio, N., Baechlein,
334 C., Baylisa, S.A., Corman, V.M., Fabera, M., Johne, R., Kamp, C., Wenzel, J.J., Coughlan, S., Di
335 Bartolo, I., Bruni, R., Ciccaglione, A.R., Garbuglia, A.R., Suffredini, E., Boxman, I., Hogema, B.,
336 van der Poel, W., Zaaijra, H., Sousaa, R. de, Velebit, B., Avellóna, A., Buti, M., Girones, R.,
337 Quer, J., Widén, F., Norder, H., Nyström, K., Bachofen, C., Sahli, R., Ijaza, S., Treagus, S., Kulka,
338 M., Rizzi, V., 2019. HEVnet: A one health, collaborative, interdisciplinary network and
339 sequence data repository for enhanced hepatitis e virus molecular typing, characterisation
340 and epidemiological investigations. *Eurosurveillance* 24, 1–6. [https://doi.org/10.2807/1560-](https://doi.org/10.2807/1560-7917.ES.2019.24.10.1800407)
341 [7917.ES.2019.24.10.1800407](https://doi.org/10.2807/1560-7917.ES.2019.24.10.1800407)
- 342 Palombieri, A., Tsachev, I., Sarchese, V., Fruci, P., Profio, F. Di, Pepovich, R., Baymakova, M.,
343 Marsilio, F., Martella, V., Martino, B. Di, 2021. *veterinary sciences* A Molecular Study on
344 Hepatitis E Virus (HEV) in Pigs in Bulgaria.
- 345 Pavia, G., Gioffrè, A., Pirolo, M., Visaggio, D., Clausi, M.T., Gherardi, M., Samele, P., Ciambrone, L.,
346 Di Natale, R., Spatari, G., Visca, P., Casalnuovo, F., 2021. Seroprevalence and phylogenetic

347 characterization of hepatitis E virus in pig farms in Southern Italy. *Prev. Vet. Med.* 194.
348 <https://doi.org/10.1016/J.PREVETMED.2021.105448>

349 Pavoni, E., Barbieri, I., Bertasi, B., Lombardi, G., Cordioli, P., Losio, M.N., 2015. Detection and
350 Molecular Characterisation of Swine Hepatitis E Virus in Brescia Province, Italy. *Ital. J. food*
351 *Saf.* 4. <https://doi.org/10.4081/IJFS.2015.4587>

352 Pierini, I., Di Bartolo, I., Manuali, E., Pirani, S., Bazzucchi, M., Moscati, L., De Mia, G.M.,
353 Giammarioli, M., 2021. Hepatitis E virus (HEV) genotype 3 diversity: Identification of a novel
354 HEV subtype in wild boar in Central Italy. *Transbound. Emerg. Dis.* 68, 2121–2129.
355 <https://doi.org/10.1111/tbed.13860>

356 Price, M.N., Dehal, P.S., Arkin, A.P., 2010. FastTree 2--approximately maximum-likelihood trees for
357 large alignments. *PLoS One* 5. <https://doi.org/10.1371/JOURNAL.PONE.0009490>

358 Ricci, A., Allende, A., Bolton, D., Chemaly, M., Davies, R., Fernandez Escamez, P.S., Herman, L.,
359 Koutsoumanis, K., Lindqvist, R., Nørrung, B., Robertson, L., Ru, G., Sanaa, M., Simmons, M.,
360 Skandamis, P., Snary, E., Speybroeck, N., Ter Kuile, B., Threlfall, J., Wahlström, H., Di Bartolo,
361 I., Johne, R., Pavio, N., Rutjes, S., van der Poel, W., Vasickova, P., Hempen, M., Messens, W.,
362 Rizzi, V., Latronico, F., Girones, R., 2017. Public health risks associated with hepatitis E virus
363 (HEV) as a food-borne pathogen. *EFSA J.* 15. <https://doi.org/10.2903/J.EFSA.2017.4886>

364 Salines, M., Andraud, M., Rose, N., 2017. From the epidemiology of hepatitis E virus (HEV) within
365 the swine reservoir to public health risk mitigation strategies : a comprehensive review. *Vet.*
366 *Res.* 1–15. <https://doi.org/10.1186/s13567-017-0436-3>

367 Schloss, P.D., Westcott, S.L., Ryabin, T., Hall, J.R., Hartmann, M., Hollister, E.B., Lesniewski, R.A.,
368 Oakley, B.B., Parks, D.H., Robinson, C.J., Sahl, J.W., Stres, B., Thallinger, G.G., Van Horn, D.J.,

369 Weber, C.F., 2009. Introducing mothur: open-source, platform-independent, community-
370 supported software for describing and comparing microbial communities. *Appl. Environ.*
371 *Microbiol.* 75, 7537–7541. <https://doi.org/10.1128/AEM.01541-09>

372 SEIEVA [WWW Document], n.d. URL <https://www.epicentro.iss.it/en/hepatitis/data-seieva#e>
373 (accessed 8.2.22).

374 Tamura, K., Stecher, G., Kumar, S., 2021. MEGA11: Molecular Evolutionary Genetics Analysis
375 Version 11. *Mol. Biol. Evol.* 38, 3022–3027. <https://doi.org/10.1093/MOLBEV/MSAB120>

376 World Health Organization (WHO), 2021. Hepatitis E [WWW Document]. URL
377 <https://www.who.int/news-room/fact-sheets/detail/hepatitis-e> (accessed 12.14.21).

378 Wu, N., Abril, C., Hinic, V., Brodard, I., Thür, B., Fattebert, J., Hüsey, D., Ryser-Degiorgis, M.P., 2011.
379 Free-ranging wild boar: A disease threat to domestic PIGS in Switzerland? *J. Wildl. Dis.* 47,
380 868–879. <https://doi.org/10.7589/0090-3558-47.4.868>

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TABLES

393 **TABLE 1.** Liver and rectal swab samples of pigs tested positive for HEV-RNA and belonged to
 394 different areas of the Sicily region

Sampling areas ^a	HEV pos./total no. of livers	HEV pos./total no. of rectal swab	Age
Santa Venerina (CT)	6/21	0	9 – 12 months
Giarre (CT)	0/6	0	9 – 12 months
Piedimento Etneo (CT)	3/12	0	9 – 12 months
Modica (RG)	3/18 ^b	3/18 ^b	9 – 12 months
Barcellona Pozzo Di Gotto (ME)	6/15	0/15	9 – 12 months
Santa Maria di Licodia (CT)	0/9	0/9	9 – 12 months
Maniace (CT)	3/18	0/18	9 – 12 months
Belpasso (CT)	0/3	0/3	9 – 12 months
Bronte (CT)	0/3	0/3	9 – 12 months
Francia	0/15 ^c	0/15 ^c	9 – 12 months
HEV POS. TOT. NO.	21/120	3/ 81	

395 **a** Every sampling area includes one farm except Modica and Piedimento Etneo presenting two farms each.

396 **b** Positive liver samples are not from the same pig as rectal swabs.

397 **c** Farmed in France and slaughtered in Sicily.

398

399 **TABLE 2. Dataset’s summary of liver and rectal swab samples of pigs tested positive for HEV-RNA and**
 400 **belonged to different areas of the Sicily region**

401 Samples with identical HEV nucleotide sequences (haplotypes), the genotype and subtypes’ assignments
 402 inferred by HEVnet, were reported for each haplotype.

403

Haplotype	Samples		Genotype	Subtype	Reference
	Liver ID ^a	Rectal swabs ID ^a			
Hap_1 15 positive samples	MODICA_2, MODICA_3 MODICA_4 SANTA_VENERINA_1, SANTA_VENERINA_2, SANTA_VENERINA_3, MANIACE_5 MANIACE_6 MANIACE_7 GOTTO_1 GOTTO_2 GOTTO_6 GOTTO_9 GOTTO_13 GOTTO_15		HEV3	HEV3c	Germany – Sus scrofa (FJ705359)
Hap_5 3 positive samples		MODICA_1S MODICA_3S MODICA_6S	HEV3	HEV3c	Germany – Sus scrofa (FJ705359)
Hap_6 3 positive samples	SANTA_VENERINA_5 SANTA_VENERINA_6 SANTA_VENERINA_7		HEV3	Could not assign	Germany – Sus scrofa (FJ705359)
Hap_7 3 positive samples	PIEDIMENTO_ETNEO_2 PIEDIMENTO_ETNEO_7 PIEDIMENTO_ETNEO_8		HEV3	Could not assign	Germany – Sus scrofa (FJ705359)

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a: The ID was generated using the municipality of pigs’ farming with a progressive number.

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FIGURES

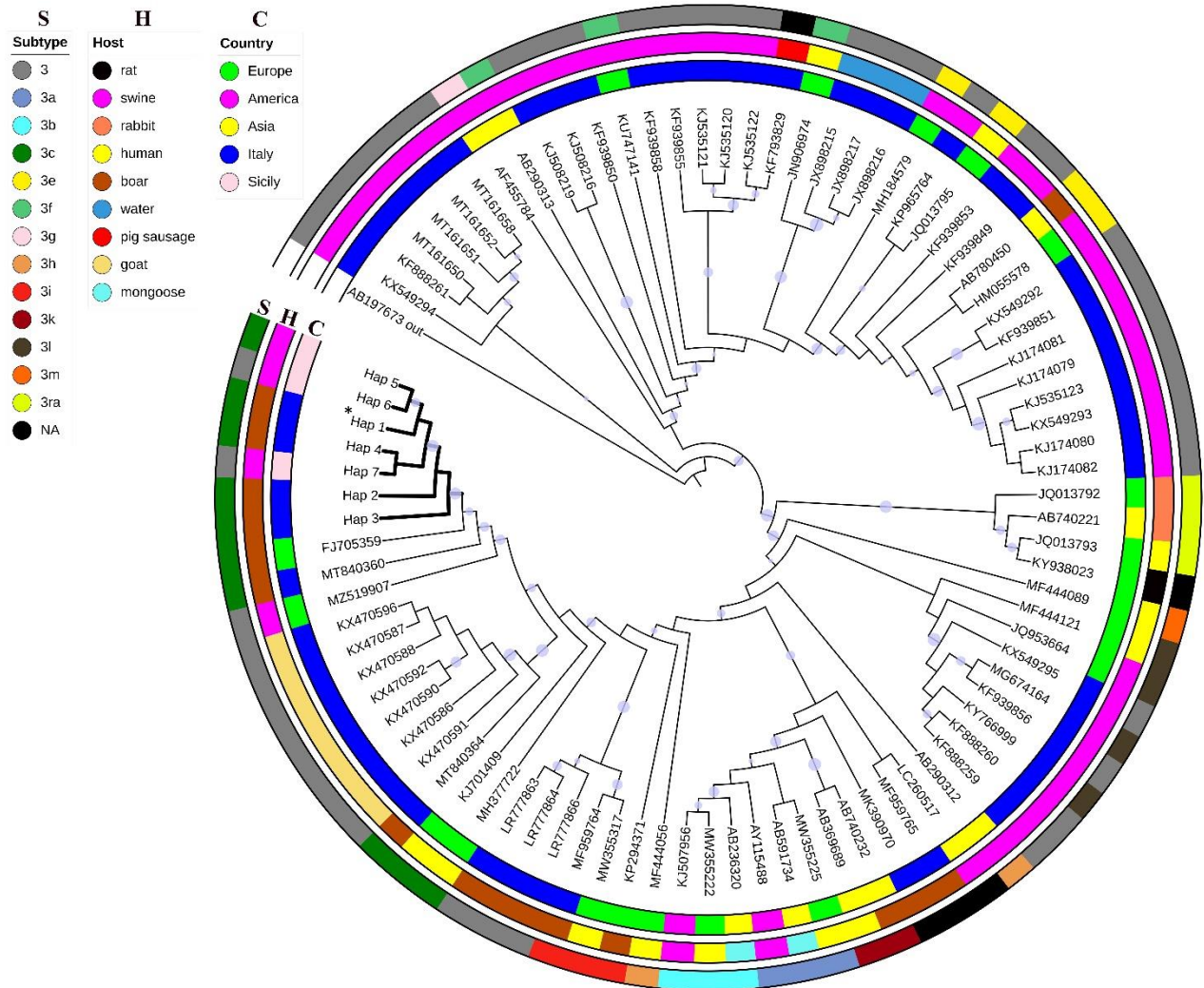


FIGURE 1. Maximum Likelihood tree of HEV 3 genotype using the RNA-dependent RNA polymerase.

The tree included our four haplotypes of 291 bp (Hap 1; Hap 5; Hap 6; Hap 7) detected in Sicily, three haplotypes (Hap 2; Hap 3; Hap 4) detected in Calabria region (Lorusso et al., 2022) and all samples available from other Italian regions and sources. Representative nucleotide sequences of other geographic areas were also included, and one HEV-4 strain was used as outgroup (AB197673). Bootstraps >70% were showed as symbols (dots) proportional to the values. Metadata associated

to nucleotide sequences were showed by three colour code panels: Subtype (S), Host (H) and Country (C).

*Haplotype 1 was detected also in pigs.

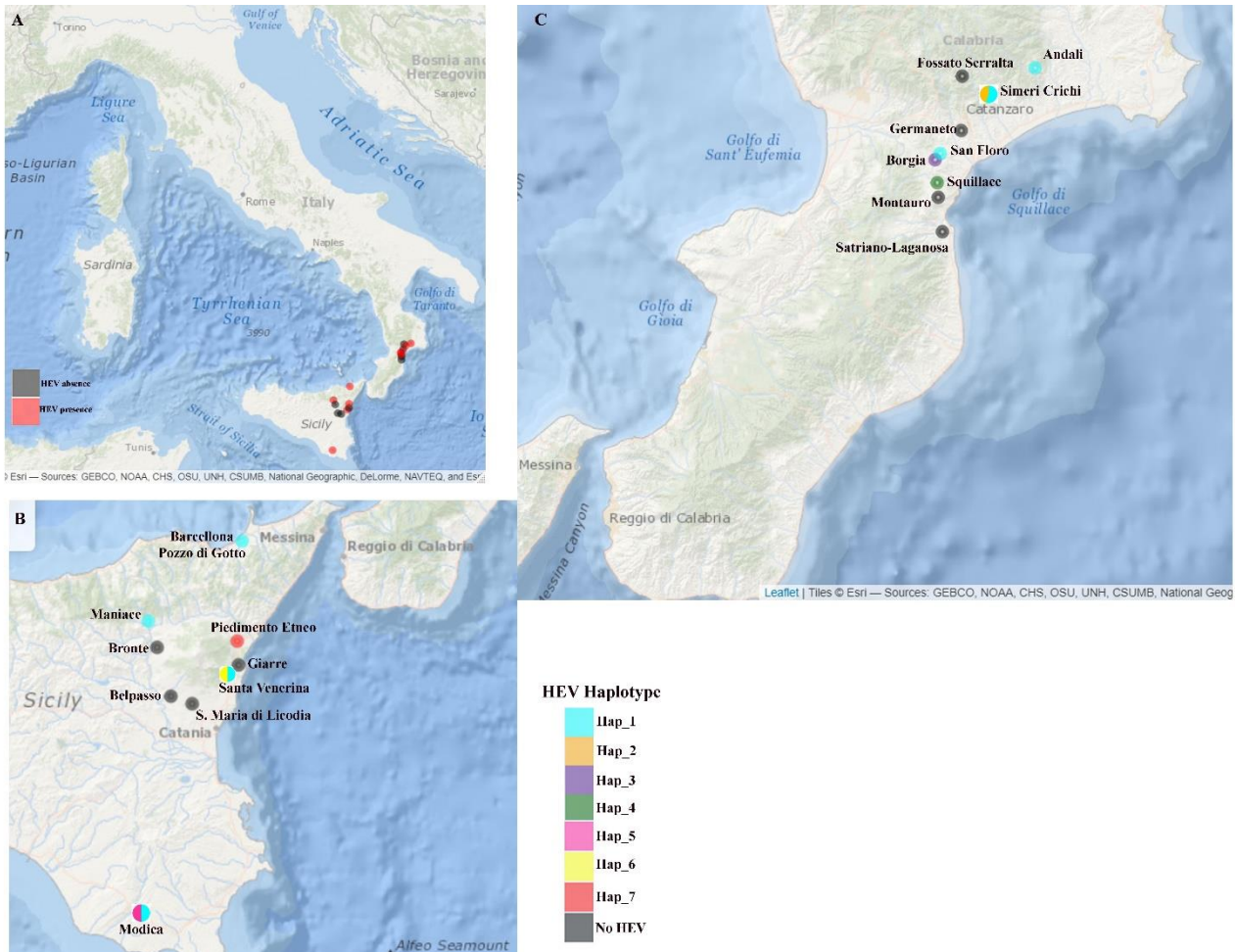


FIGURE 2.

A) Samples' georeferentiation and HEV results.

The map shows the georeferentiation of liver and rectal swabs' samples collected in Calabria and Sicily's municipalities. The HEV occurrence was also described: the red dots represent the areas where HEV was detected; grey dots represent areas with HEV absence.

B) Samples' map and haplotypes detected in Sicily region.

Map of sampling and corresponding haplotypes found in each municipality. Grey dots indicate sites where HEV was not detected.

C) Samples' map and haplotypes detected in Calabrian region.

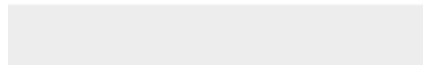
Map of sampling and corresponding haplotypes found in each municipality. Grey dots indicate sites where HEV was not detected.

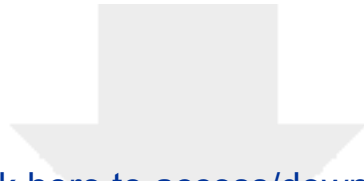


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Supplementary Interactive Plot Data (CSV)

Supplementary data 1.xlsx

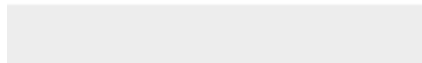




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Supplementary Interactive Plot Data (CSV)

SUPP. FIG. 1.docx



Conflict of Interest and Authorship Conformation Form

Please check the following as appropriate:

All authors have participated in (a) conception and design, or analysis and interpretation of the data; (b) drafting the article or revising it critically for important intellectual content; and (c) approval of the final version.

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