

High mortality in foals associated with *Salmonella enterica* subsp. *enterica* Abortusequi infection in Italy

Journal of Veterinary Diagnostic Investigation
1–3
© 2018 The Author(s)
Reprints and permissions:
sagepub.com/journalsPermissions.nav
DOI: 10.1177/1040638717753965
jvdi.sagepub.com

Erika Grandolfo,¹ Antonio Parisi, Antonia Ricci, Eleonora Lorusso, Rocco de Siena, Adriana Trotta, Domenico Buonavoglia, Vito Martella, Marialaura Corrente

Abstract. *Salmonella enterica* subsp. *enterica* serovar Abortusequi is frequently reported as a cause of abortion in mares and neonatal septicemia and polyarthritis in Asian and African countries, but only sporadically in Europe and the United States. We report an outbreak of *S. Abortusequi* in foals in Italy, characterized by high mortality. In a herd of Murgese horses, 10 of 34 newborns died at birth and a further 7 died, after developing severe clinical signs, during the first 10 d of life. Tissue specimens from different organs of 2 dead foals, synovial fluids from 4 sick foals, and vaginal and rectal swabs from their dams were cultured. A total of 16 isolates, all as pure cultures, were obtained and identified as *Salmonella*. The isolates exhibited the same antimicrobial resistance pattern and the same sequence type, ST251, a type that has been associated with *S. Abortusequi*. Six of 16 isolates were serotyped and found to be *S. Abortusequi* 4,12:-:e,n,x. Equine practitioners should be aware of *S. Abortusequi* infection as a cause of neonatal mortality in foals.

Key words: Murgese foals; outbreak; *Salmonella* Abortusequi.

Salmonella enterica subsp. *enterica* serovar Abortusequi is a host-adapted serovar that is associated with abortion in mares and neonatal septicemia and polyarthritis.^{3,15,17} Moreover, equine salmonellosis is often reported as a secondary bacterial infection associated with equid herpesvirus 1 (EHV-1; species *Equid alphaherpesvirus 1*) infection.^{18,19} Although reported commonly from Asian and African countries, *S. Abortusequi* is only isolated sporadically in Europe, the United States, and Argentina.^{3,9,12–14,16}

We describe herein a severe disease outbreak in foals, characterized by high mortality, which was caused by *S. Abortusequi*. The herd was located in Altamura, Apulia, Italy, and consisted of 72 Murgese horses (40 mares, 7 stallions, and 25 yearlings). From January to April 2016, 34 foals were born from mares without clinical signs. Ten of the 34 newborns died at birth. A further 7 foals developed severe clinical signs including fever up to 41°C, lethargy, bloody diarrhea, and lameness, and died during the first 10 d of life (overall mortality rate of 50%). An additional 4 foals of the same herd, 15–20 d of age, exhibited similar clinical signs. Abortion or clinical signs were not reported in the mares in the months before the disease outbreak.

Autopsies of 2 dead foals revealed diffuse hemorrhagic inflammation of the cecum and large colon, hepatization of the lungs, and necrotic lesions of the kidneys and liver. The lymph nodes were swollen and hemorrhagic. Tissue specimens from mediastinal lymph nodes, lung, kidney, liver, spleen, and colon of the 2 dead foals, and synovial fluids of

4 sick foals, were collected. Vaginal and rectal swabs from their dams ($n = 6$) were also included. The tissues were cultured on MacConkey agar and 5% sheep blood agar and incubated at 37°C for 24 and 48 h, respectively.⁴ Gram-negative, lactose-negative organisms were cultured on triple sugar iron (Oxoid, Milan, Italy). All lactose-negative, H₂S-negative isolates were tested by means of a PCR assay targeting the *invA* gene.¹¹ Antimicrobial susceptibility testing was performed by the agar diffusion disk method,^{5,6} testing the following antimicrobials: chloramphenicol (30 µg), doxycycline (30 µg), tetracycline (30 µg), ampicillin (10 µg), amoxicillin–clavulanic acid (20 µg of amoxicillin + 10 µg of clavulanic acid), cefuroxime (30 µg), ceftazidime (30 µg), colistin (10 µg), gentamicin (10 µg), ciprofloxacin (10 µg), enrofloxacin (5 µg), streptomycin (10 µg), sulfamethoxazole (50 µg), and trimethoprim–sulfamethoxazole (1.25 µg of trimethoprim + 23.75 µg of sulfamethoxazole). The discs were all obtained from a single source (Liofilchem, Teramo,

Department of Veterinary Medicine, University of Bari “Aldo Moro”, Valenzano, Italy (Grandolfo, Lorusso, Trotta, Buonavoglia, Martella, Corrente); Experimental Zooprophyllactic Institute of Puglia and Basilicata, Putignano, Italy (Parisi); National/OIE Reference Laboratory for Salmonella, Experimental Zooprophyllactic Institute of Venice, Italy (Ricci); Veterinary practitioner, Italy (de Siena).

¹Corresponding author: Erika Grandolfo, Department of Veterinary Medicine, University of Bari “Aldo Moro”, Valenzano, Italy 70010. egrandolfo87@gmail.com

Italy). Clinical and Laboratory Standards Institute (CLSI) breakpoints⁵ for bacteria associated with infections of humans were used for the interpretation of disk diffusion results for cefuroxime and ceftazidime, whereas veterinary CLSI breakpoints⁶ were used for the interpretation of the remaining antimicrobials. Six of 16 isolates (obtained from lung and liver of 2 dead foals and synovial fluids of 2 sick foals) were serotyped at the National/OIE Reference Laboratory for *Salmonella* (Experimental Zooprophyllactic Institute of Venice, IT) according to the Kaufmann–White scheme.^{8,10} Strains were analyzed by multi-locus sequence typing (MLST).¹ The MLST sequence type was assigned through the Enterobase website (<https://goo.gl/3AfZP8>). DNA and RNA were extracted from lung and liver of foals and from the vaginal swabs (QIAamp *cad*or pathogen mini kit, Qiagen, Hilden, Germany) and tested by real-time PCR for EHV-1⁷ and by reverse-transcription PCR for equine arteritis virus (EAV).²

From the synovial fluids of 4 sick foals, and from the organs of 2 dead foals, 16 isolates were obtained and identified as *Salmonella* spp. by PCR. The isolates exhibited the same antimicrobial resistance pattern (i.e., susceptible to chloramphenicol, tetracycline, ampicillin, amoxicillin–clavulanic acid, cefuroxime, ceftazidime, gentamicin, enrofloxacin, and trimethoprim–sulfamethoxazole, and resistant to doxycycline, colistin, ciprofloxacin, streptomycin, and sulfamethoxazole). Six representative strains were serotyped as *S. Abortusequi* 4,12:-:e,n,x. All of the isolates from the foals were characterized as sequence type (ST)251 (<https://goo.gl/3AfZP8>). All of the samples from the foals were negative to the virologic screening (EHV-1 and EAV), and no other bacteria were isolated. The rectal and vaginal samples obtained from the mares were negative in the virologic screening, and *Salmonella* spp. were not detected. After the etiologic diagnosis and on the basis of the antimicrobial resistance patterns of the isolates, 4 symptomatic foals were treated with trimethoprim–sulfamethoxazole (20 mg/kg once a day) and gentamicin (6 mg/kg once a day) via the intramuscular route for 10 d. Three foals recovered fully without permanent sequelae; one foal had permanent signs of polyarthritis.

Several microbes can be involved in the etiology of genital and neonatal disorders in equids.^{17,18} Equine paratyphoid, caused by *S. Abortusequi*, is an infectious disease characterized by contagious abortions in most equids and is generally only reported sporadically in Europe.^{12–14} In our study, pure cultures of *Salmonella* were isolated from all of the samples ($n = 16$) from the foals subjected to our laboratory investigations, with all isolates shown to be of the same sequence type and 6 confirmed as *S. Abortusequi*. No preliminary signs of abortion were reported, and the source of the outbreak could not be identified confidently. The rectal and vaginal samples of the relevant dams were negative, and it was not possible to test the other mares. However, the

epidemiologic investigation indicated that a possible source of infection could have been 2 mares that had been introduced into the herd recently. Given that the horses lived and grazed freely, it is possible that one or more carriers had infected the other animals or the newborns at grazing or during delivery of some mares. Horses infected with *S. Abortusequi* may act as long-term carriers even after their recovery, and it has been suggested that recovered horses pose a risk as new sources of infection.¹⁷ It is difficult to distinguish between infections acquired in utero and infections that are acquired neonatally; the long-term consequences for foals depend on the severity of the disease.¹⁷

The isolates obtained from the outbreak were typed as ST251, a clonal type previously described for isolates of *S. Abortusequi* from Croatia, Argentina, and the United States (<https://goo.gl/3AfZP8>). Disease caused by *S. Abortusequi* has been reported in Italy previously,¹⁴ but the isolates were subjected to only serotyping and not genotyping, thus hindering a precise comparison. The Murghese horse is an autochthonous Italian breed, which is usually considered to be intrinsically resistant to most diseases, and vaccination was not a feature of herd management in our case. Equine practitioners and breeders should be aware of the risks posed by *S. Abortusequi* infection and employ appropriate preventive measures.

Declaration of conflicting interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

ORCID iD

Erika Grandolfo  <http://orcid.org/0000-0002-6829-344X>

References

1. Achtman M, et al. Multilocus sequence typing as a replacement for serotyping in *Salmonella enterica*. PLoS Pathog 2012;8:e1002776.
2. Balasuriya UB, et al. Detection of equine arteritis virus by real-time TaqMan reverse transcription-PCR assay. J Virol Methods 2002;101:21–28.
3. Bustos CP, et al. *Salmonella enterica* serovar Abortusequi as an emergent pathogen causing equine abortion in Argentina. J Equine Vet Sci 2016;39(Suppl):S58–S59.
4. Carter GR, Cole JR. Diagnostic Procedures in Veterinary Bacteriology and Mycology. 5th ed. Vol. 1. San Diego, CA: Academic Press, 1990.
5. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing. 27th ed. Wayne, PA: CLSI, 2017. CLSI supplement M100.

6. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals; approved guideline. 4th ed. Wayne, PA: CLSI, 2013. CLSI document VET01-S.
7. Elia G, et al. Detection of equine herpesvirus type 1 by real time PCR. *J Virol Methods* 2006;133:70–75.
8. Grimont PAD, Weill FX. Antigenic Formulae of the *Salmonella* Serovars. 9th ed. Paris, France: World Health Organization Collaborating Centre for Reference and Research on Salmonella, Institut Pasteur, 2007.
9. Hong B, et al. Equine abortion and stillbirth in central Kentucky during 1988 and 1989 foaling season. *J Vet Diagn Invest* 1993;5:560–566.
10. Issenhuth-Jeanjean S, et al. Supplement 2008–2010 (no. 48) to the White-Kauffmann-Le Minor scheme. *Res Microbiol* 2014;165:526–530.
11. Khan AA, et al. Detection of multidrug-resistant *Salmonella typhimurium* DT104 by multiplex polymerase chain reaction. *FEMS Microbiol Lett* 2000;182:355–360.
12. Llorente L, et al. Occurrence of multiple abortions due to *Salmonella enterica* serovar Abortusequi infection. *J Equine Vet Sci* 2016;39(Suppl):S58.
13. Madić J, et al. An outbreak of abortion in mares associated with *Salmonella* Abortusequi infection. *Equine Vet J* 1997;29:230–233.
14. Marenzoni ML, et al. Causes of equine abortion, stillbirth and neonatal death in central Italy. *Vet Rec* 2012;170:262.
15. Rodriguez A, et al. Prevalence of *Salmonella* in diverse environmental farm samples. *J Food Prot* 2006;69:2576–2580.
16. Spier SJ. Salmonellosis. *Vet Clin North Am Equine Pract* 1993;9:385–397.
17. Swerczek TW. Identifying the bacterial causes of abortion in mares. *Vet Med* 1991;86:1210–1216.
18. Swerczek TW. The most common viral causes of equine abortion. *Vet Med* 1991;86:1205–1208.
19. Tewari SS, et al. Equine herpesvirus 1 and neonatal foal mortality in northern India. *Rev Sci Tech* 1989;8:103–110.